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(54) Title: FIBROBLAST GROWTH FACTOR-23 MOLECULES AND USES THEREOF

(57) Abstract: The present invention provides Fibroblast Growth Factor-23 (FGF-23) polypeptides and nucleic acid molecules encoding the same. The invention also provides selective binding agents, vectors, host cells, and methods for producing FGF-23 polypeptides. The invention further provides pharmaceutical compositions and methods for the diagnosis, treatment, amelioration, and/or prevention of diseases, disorders, and conditions associated with FGF-23 polypeptides.



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FIBROBLAST GROWTH FACTOR-23 MOLECULES AND USES THEREOF

Field of the Invention

5 The present invention relates to Fibroblast Growth Factor-23 (FGF-23) polypeptides and nucleic acid molecules encoding the same. The invention also relates to selective binding agents, vectors, host cells, and methods for producing FGF-23 polypeptides. The invention further relates to pharmaceutical compositions and methods for the diagnosis, treatment, amelioration, and/or
10 prevention of diseases, disorders, and conditions associated with FGF-23 polypeptides.

Background of the Invention

 Technical advances in the identification, cloning, expression, and
15 manipulation of nucleic acid molecules and the deciphering of the human genome have greatly accelerated the discovery of novel therapeutics. Rapid nucleic acid sequencing techniques can now generate sequence information at unprecedented rates and, coupled with computational analyses, allow the assembly of overlapping sequences into partial and entire genomes and the identification of
20 polypeptide-encoding regions. A comparison of a predicted amino acid sequence against a database compilation of known amino acid sequences allows one to determine the extent of homology to previously identified sequences and/or structural landmarks. The cloning and expression of a polypeptide-encoding region of a nucleic acid molecule provides a polypeptide product for structural
25 and functional analyses. The manipulation of nucleic acid molecules and encoded polypeptides may confer advantageous properties on a product for use as a therapeutic.

 In spite of the significant technical advances in genome research over the past decade, the potential for the development of novel therapeutics based on the
30 human genome is still largely unrealized. Many genes encoding potentially beneficial polypeptide therapeutics or those encoding polypeptides, which may

act as "targets" for therapeutic molecules, have still not been identified.

Accordingly, it is an object of the invention to identify novel polypeptides, and nucleic acid molecules encoding the same, which have diagnostic or therapeutic benefit.

5

Summary of the Invention

The present invention relates to novel FGF-23 nucleic acid molecules and encoded polypeptides.

The invention provides for an isolated nucleic acid molecule comprising a
10 nucleotide sequence selected from the group consisting of:

- (a) the nucleotide sequence as set forth in SEQ ID NO: 1;
- (b) the nucleotide sequence of the DNA insert in ATCC Deposit No. PTA-1617;
- (c) a nucleotide sequence encoding the polypeptide as set forth in SEQ
15 ID NO: 2;
- (d) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (c); and
- (e) a nucleotide sequence complementary to any of (a) - (c).

20 The invention also provides for an isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:

- (a) a nucleotide sequence encoding a polypeptide which is at least about 70 percent identical to the polypeptide as set forth in SEQ ID NO: 2, wherein the encoded polypeptide has an activity of the polypeptide set forth in
25 SEQ ID NO: 2;

- (b) a nucleotide sequence encoding an allelic variant or splice variant of the nucleotide sequence as set forth in SEQ ID NO: 1, the nucleotide sequence of the DNA insert in ATCC Deposit No. PTA-1617, or (a);

- (c) a region of the nucleotide sequence of SEQ ID NO: 1, the DNA
30 insert in ATCC Deposit No. PTA-1617, (a), or (b) encoding a polypeptide fragment of at least about 25 amino acid residues, wherein the polypeptide

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fragment has an activity of the encoded polypeptide as set forth in SEQ ID NO: 2, or is antigenic;

(d) a region of the nucleotide sequence of SEQ ID NO: 1, the DNA insert in ATCC Deposit No. PTA-1617, or any of (a) - (c) comprising a fragment
5 of at least about 16 nucleotides;

(e) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (d); and

(f) a nucleotide sequence complementary to any of (a) - (d).

10 The invention further provides for an isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:

(a) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one conservative amino acid substitution, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

15 (b) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid insertion, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(c) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid deletion, wherein the encoded polypeptide
20 has an activity of the polypeptide set forth in SEQ ID NO: 2;

(d) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 which has a C- and/or N- terminal truncation, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(e) a nucleotide sequence encoding a polypeptide as set forth in SEQ
25 ID NO: 2 with at least one modification selected from the group consisting of amino acid substitutions, amino acid insertions, amino acid deletions, C-terminal truncation, and N-terminal truncation, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(f) a nucleotide sequence of any of (a) - (e) comprising a fragment of
30 at least about 16 nucleotides;

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- (g) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (f); and
- (h) a nucleotide sequence complementary to any of (a) - (e).

5 The present invention provides for an isolated polypeptide comprising an amino acid sequence selected from the group consisting of:

- (a) the amino acid sequence as set forth in SEQ ID NO: 2; and
- (b) the amino acid sequence encoded by the DNA insert in ATCC Deposit No. PTA-1617.

10

The invention also provides for an isolated polypeptide comprising the amino acid sequence selected from the group consisting of:

- (a) the amino acid sequence as set forth in SEQ ID NO: 3, optionally further comprising an amino-terminal methionine;
- 15 (b) an amino acid sequence for an ortholog of SEQ ID NO: 2;
- (c) an amino acid sequence which is at least about 70 percent identical to the amino acid sequence of SEQ ID NO: 2, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;
- (d) a fragment of the amino acid sequence set forth in SEQ ID NO: 2
- 20 comprising at least about 25 amino acid residues, wherein the fragment has an activity of the polypeptide set forth in SEQ ID NO: 2, or is antigenic; and
- (e) an amino acid sequence for an allelic variant or splice variant of the amino acid sequence as set forth in SEQ ID NO: 2, the amino acid sequence encoded by the DNA insert in ATCC Deposit No. PTA-1617, or (a) - (c).

25

The invention further provides for an isolated polypeptide comprising the amino acid sequence selected from the group consisting of:

- (a) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one conservative amino acid substitution, wherein the polypeptide has an activity
- 30 of the polypeptide set forth in SEQ ID NO: 2;

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(b) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one amino acid insertion, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(c) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one amino acid deletion, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(d) the amino acid sequence as set forth in SEQ ID NO: 2 which has a C- and/or N- terminal truncation, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2; and

(e) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one modification selected from the group consisting of amino acid substitutions, amino acid insertions, amino acid deletions, C-terminal truncation, and N-terminal truncation, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2.

15

Also provided are fusion polypeptides comprising FGF-23 amino acid sequences.

The present invention also provides for an expression vector comprising the isolated nucleic acid molecules as set forth herein, recombinant host cells comprising the recombinant nucleic acid molecules as set forth herein, and a method of producing an FGF-23 polypeptide comprising culturing the host cells and optionally isolating the polypeptide so produced.

A transgenic non-human animal comprising a nucleic acid molecule encoding an FGF-23 polypeptide is also encompassed by the invention. The FGF-23 nucleic acid molecules are introduced into the animal in a manner that allows expression and increased levels of an FGF-23 polypeptide, which may include increased circulating levels. Alternatively, the FGF-23 nucleic acid molecules are introduced into the animal in a manner that prevents expression of endogenous FGF-23 polypeptide (*i.e.*, generates a transgenic animal possessing an FGF-23 polypeptide gene knockout). The transgenic non-human animal is preferably a mammal, and more preferably a rodent, such as a rat or a mouse.

Also provided are derivatives of the FGF-23 polypeptides of the present invention.

Additionally provided are selective binding agents such as antibodies and peptides capable of specifically binding the FGF-23 polypeptides of the invention.

5 Such antibodies and peptides may be agonistic or antagonistic.

Pharmaceutical compositions comprising the nucleotides, polypeptides, or selective binding agents of the invention and one or more pharmaceutically acceptable formulation agents are also encompassed by the invention. The pharmaceutical compositions are used to provide therapeutically effective
10 amounts of the nucleotides or polypeptides of the present invention. The invention is also directed to methods of using the polypeptides, nucleic acid molecules, and selective binding agents.

The FGF-23 polypeptides and nucleic acid molecules of the present invention may be used to treat, prevent, ameliorate, and/or detect diseases and
15 disorders, including those recited herein.

The present invention also provides a method of assaying test molecules to identify a test molecule that binds to an FGF-23 polypeptide. The method comprises contacting an FGF-23 polypeptide with a test molecule to determine the extent of binding of the test molecule to the polypeptide. The method further
20 comprises determining whether such test molecules are agonists or antagonists of an FGF-23 polypeptide. The present invention further provides a method of testing the impact of molecules on the expression of FGF-23 polypeptide or on the activity of FGF-23 polypeptide.

Methods of regulating expression and modulating (*i.e.*, increasing or
25 decreasing) levels of an FGF-23 polypeptide are also encompassed by the invention. One method comprises administering to an animal a nucleic acid molecule encoding an FGF-23 polypeptide. In another method, a nucleic acid molecule comprising elements that regulate or modulate the expression of an FGF-23 polypeptide may be administered. Examples of these methods include
30 gene therapy, cell therapy, and anti-sense therapy as further described herein.

In another aspect of the present invention, the FGF-23 polypeptides may be used for identifying receptors thereof ("FGF-23 polypeptide receptors"). Various forms of "expression cloning" have been extensively used to clone receptors for protein ligands. See, e.g., Simonsen and Lodish, 1994, *Trends Pharmacol. Sci.* 15:437-41 and Tartaglia *et al.*, 1995, *Cell* 83:1263-71. The isolation of an FGF-23 polypeptide receptor is useful for identifying or developing novel agonists and antagonists of the FGF-23 polypeptide signaling pathway. Such agonists and antagonists include soluble FGF-23 polypeptide receptors, anti-FGF-23 polypeptide receptor-selective binding agents (such as antibodies and derivatives thereof), small molecules, and antisense oligonucleotides, any of which can be used for treating one or more disease or disorder, including those disclosed herein.

Brief Description of the Figures

Figures 1A-1B illustrate the nucleotide sequence of the human FGF-23 gene (SEQ ID NO: 1) and the deduced amino acid sequence of human FGFR polypeptide (SEQ ID NO: 2). The predicted signal peptide is indicated (underlined);

Figures 2A-2G illustrate the amino acid sequence alignment of human FGF-1 (hu FGF-1; SEQ ID NO: 4), human FGF-2 (hu FGF-2; SEQ ID NO: 5), human FGF-3 (hu FGF-3; SEQ ID NO: 6), human FGF-4 (hu FGF-4; SEQ ID NO: 7), human FGF-5 (hu FGF-5; SEQ ID NO: 8), human FGF-6 (hu FGF-6; SEQ ID NO: 9), human FGF-7 (hu FGF-7; SEQ ID NO: 10), human FGF-8 (hu FGF-8; SEQ ID NO: 11), human FGF-9 (hu FGF-9; SEQ ID NO: 12), human FGF-10 (hu FGF-10; SEQ ID NO: 13), human FGF-11 (hu FGF-11; SEQ ID NO: 14), human FGF-12 (hu FGF-12; SEQ ID NO: 15), human FGF-13 (hu FGF-13; SEQ ID NO: 16), human FGF-14 (hu FGF-14; SEQ ID NO: 17), human FGF-16 (hu FGF-16; SEQ ID NO: 18), human FGF-17 (hu FGF-17; SEQ ID NO: 19), human FGF-18 (hu FGF-18; SEQ ID NO: 20), human FGF-19 (hu FGF-19; SEQ ID NO: 21), human FGF-23 (hu FGF-23; SEQ ID NO: 22), murine FGF-1 (mu FGF-1; SEQ ID NO:

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23), murine FGF-2 (mu FGF-2; SEQ ID NO: 24), murine FGF-3 (mu FGF-3; SEQ ID NO: 25), murine FGF-4 (mu FGF-4; SEQ ID NO: 26), murine FGF-5 (mu FGF-5; SEQ ID NO: 27), murine FGF-6 (mu FGF-6; SEQ ID NO: 28), murine FGF-7 (mu FGF-7; SEQ ID NO: 29), murine FGF-8 (mu FGF-8; SEQ ID NO: 30), murine FGF-9 (mu FGF-9; SEQ ID NO: 31), murine FGF-10 (mu FGF-10; SEQ ID NO: 32), murine FGF-11 (mu FGF-11; SEQ ID NO: 33), murine FGF-12 (mu FGF-12; SEQ ID NO: 34), murine FGF-13 (mu FGF-13; SEQ ID NO: 35), murine FGF-14 (mu FGF-14; SEQ ID NO: 36), murine FGF-15 (mu FGF-15; SEQ ID NO: 37), rat FGF-16 (rat FGF-16; SEQ ID NO: 38), murine FGF-17 (mu FGF-17; SEQ ID NO: 39);

Figure 3 illustrates the expression of FGF-23 mRNA as detected by *in situ* hybridization in the brain and cardiac muscle (heart) of a normal adult mouse (H&E = hematoxylin and eosin counterstaining; ISH = *in situ* hybridization);

15

Figure 4 illustrates the expression of FGF-23 mRNA as detected by *in situ* hybridization in the subcapsular region of the lymph node (lymph node), thymic medulla (thymus), lacunae of cortical bone from the tibia (tibia), and trabecular bone in the head (head) of a non-expressing transgenic mouse (H&E = hematoxylin and eosin counterstaining; ISH = *in situ* hybridization);

20

Figure 5 illustrates the expression of FGF-23 mRNA as detected by *in situ* hybridization in the liver, spleen, thymic medulla (thymus), and megakaryocytes in the bone marrow (bone marrow) of a high expressing transgenic mouse (H&E = hematoxylin and eosin counterstaining; ISH = *in situ* hybridization);

25

Figure 6 illustrates the expression of FGF-23 mRNA as detected by *in situ* hybridization in the smooth muscle tissue near the prostrate (smooth muscle), muscle tissue of the jaw (muscle), chondrocytes in the tibia (tibia), and chondrocytes in the vertebrae (vertebrae) of a high expressing transgenic mouse (H&E = hematoxylin and eosin counterstaining; ISH = *in situ* hybridization).

30

Detailed Description of the Invention

The section headings used herein are for organizational purposes only and are not to be construed as limiting the subject matter described. All references
5 cited in this application are expressly incorporated by reference herein.

Definitions

The terms "FGF-23 gene" or "FGF-23 nucleic acid molecule" or "FGF-23 polynucleotide" refer to a nucleic acid molecule comprising or consisting of a
10 nucleotide sequence as set forth in SEQ ID NO: 1, a nucleotide sequence encoding the polypeptide as set forth in SEQ ID NO: 2, a nucleotide sequence of the DNA insert in ATCC Deposit No. PTA-1617, and nucleic acid molecules as defined herein.

The term "FGF-23 polypeptide allelic variant" refers to one of several
15 possible naturally occurring alternate forms of a gene occupying a given locus on a chromosome of an organism or a population of organisms.

The term "FGF-23 polypeptide splice variant" refers to a nucleic acid molecule, usually RNA, which is generated by alternative processing of intron sequences in an RNA transcript of FGF-23 polypeptide amino acid sequence as
20 set forth in SEQ ID NO: 2.

The term "isolated nucleic acid molecule" refers to a nucleic acid molecule of the invention that (1) has been separated from at least about 50 percent of proteins, lipids, carbohydrates, or other materials with which it is naturally found when total nucleic acid is isolated from the source cells, (2) is not
25 linked to all or a portion of a polynucleotide to which the "isolated nucleic acid molecule" is linked in nature, (3) is operably linked to a polynucleotide which it is not linked to in nature, or (4) does not occur in nature as part of a larger polynucleotide sequence. Preferably, the isolated nucleic acid molecule of the present invention is substantially free from any other contaminating nucleic acid
30 molecule(s) or other contaminants that are found in its natural environment that would interfere with its use in polypeptide production or its therapeutic,

diagnostic, prophylactic or research use.

The term "nucleic acid sequence" or "nucleic acid molecule" refers to a DNA or RNA sequence. The term encompasses molecules formed from any of the known base analogs of DNA and RNA such as, but not limited to 4-
5 acetylcytosine, 8-hydroxy-N6-methyladenosine, aziridinyl-cytosine, pseudoisocytosine, 5-(carboxyhydroxymethyl) uracil, 5-fluorouracil, 5-bromouracil, 5-carboxymethylaminomethyl-2-thiouracil, 5-carboxymethylaminomethyluracil, dihydrouracil, inosine, N6-iso-pentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanine, 1-methylinosine, 2,2-
10 dimethyl-guanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-methyladenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyamino-methyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarbonyl-methyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid methylester, uracil-5-
15 oxyacetic acid, oxybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, N-uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, pseudouracil, queosine, 2-thiocytosine, and 2,6-diaminopurine.

The term "vector" is used to refer to any molecule (*e.g.*, nucleic acid, plasmid, or virus) used to transfer coding information to a host cell.
20

The term "expression vector" refers to a vector that is suitable for transformation of a host cell and contains nucleic acid sequences that direct and/or control the expression of inserted heterologous nucleic acid sequences. Expression includes, but is not limited to, processes such as transcription,
25 translation, and RNA splicing, if introns are present.

The term "operably linked" is used herein to refer to an arrangement of flanking sequences wherein the flanking sequences so described are configured or assembled so as to perform their usual function. Thus, a flanking sequence operably linked to a coding sequence may be capable of effecting the replication,
30 transcription and/or translation of the coding sequence. For example, a coding sequence is operably linked to a promoter when the promoter is capable of

directing transcription of that coding sequence. A flanking sequence need not be contiguous with the coding sequence, so long as it functions correctly. Thus, for example, intervening untranslated yet transcribed sequences can be present between a promoter sequence and the coding sequence and the promoter sequence
5 can still be considered "operably linked" to the coding sequence.

The term "host cell" is used to refer to a cell which has been transformed, or is capable of being transformed with a nucleic acid sequence and then of expressing a selected gene of interest. The term includes the progeny of the parent cell, whether or not the progeny is identical in morphology or in genetic
10 make-up to the original parent, so long as the selected gene is present.

The term "FGF-23 polypeptide" refers to a polypeptide comprising the amino acid sequence of SEQ ID NO: 2 and related polypeptides. Related polypeptides include FGF-23 polypeptide fragments, FGF-23 polypeptide orthologs, FGF-23 polypeptide variants, and FGF-23 polypeptide derivatives,
15 which possess at least one activity of the polypeptide as set forth in SEQ ID NO: 2. FGF-23 polypeptides may be mature polypeptides, as defined herein, and may or may not have an amino-terminal methionine residue, depending on the method by which they are prepared.

The term "FGF-23 polypeptide fragment" refers to a polypeptide that
20 comprises a truncation at the amino-terminus (with or without a leader sequence) and/or a truncation at the carboxyl-terminus of the polypeptide as set forth in SEQ ID NO: 2. The term "FGF-23 polypeptide fragment" also refers to amino-terminal and/or carboxyl-terminal truncations of FGF-23 polypeptide orthologs, FGF-23 polypeptide derivatives, or FGF-23 polypeptide variants, or to amino-
25 terminal and/or carboxyl-terminal truncations of the polypeptides encoded by FGF-23 polypeptide allelic variants or FGF-23 polypeptide splice variants. FGF-23 polypeptide fragments may result from alternative RNA splicing or from *in vivo* protease activity. Membrane-bound forms of an FGF-23 polypeptide are also contemplated by the present invention. In preferred embodiments, truncations
30 and/or deletions comprise about 10 amino acids, or about 20 amino acids, or about 50 amino acids, or about 75 amino acids, or about 100 amino acids, or more than

about 100 amino acids. The polypeptide fragments so produced will comprise about 25 contiguous amino acids, or about 50 amino acids, or about 75 amino acids, or about 100 amino acids, or about 150 amino acids, or about 200 amino acids, or more than about 200 amino acids. Such FGF-23 polypeptide fragments
5 may optionally comprise an amino-terminal methionine residue. It will be appreciated that such fragments can be used, for example, to generate antibodies to FGF-23 polypeptides.

The term "FGF-23 polypeptide ortholog" refers to a polypeptide from another species that corresponds to FGF-23 polypeptide amino acid sequence as set forth in SEQ ID NO: 2. For example, mouse and human FGF-23 polypeptides
10 are considered orthologs of each other.

The term "FGF-23 polypeptide variants" refers to FGF-23 polypeptides comprising amino acid sequences having one or more amino acid sequence substitutions, deletions (such as internal deletions and/or FGF-23 polypeptide
15 fragments), and/or additions (such as internal additions and/or FGF-23 fusion polypeptides) as compared to the FGF-23 polypeptide amino acid sequence set forth in SEQ ID NO: 2 (with or without a leader sequence). Variants may be naturally occurring (*e.g.*, FGF-23 polypeptide allelic variants, FGF-23 polypeptide orthologs, and FGF-23 polypeptide splice variants) or artificially constructed.
20 Such FGF-23 polypeptide variants may be prepared from the corresponding nucleic acid molecules having a DNA sequence that varies accordingly from the DNA sequence as set forth in SEQ ID NO: 1. In preferred embodiments, the variants have from 1 to 3, or from 1 to 5, or from 1 to 10, or from 1 to 15, or from 1 to 20, or from 1 to 25, or from 1 to 50, or from 1 to 75, or from 1 to 100, or
25 more than 100 amino acid substitutions, insertions, additions and/or deletions, wherein the substitutions may be conservative, or non-conservative, or any combination thereof.

The term "FGF-23 polypeptide derivatives" refers to the polypeptide as set forth in SEQ ID NO: 2, FGF-23 polypeptide fragments, FGF-23 polypeptide
30 orthologs, or FGF-23 polypeptide variants, as defined herein, that have been chemically modified. The term "FGF-23 polypeptide derivatives" also refers to

the polypeptides encoded by FGF-23 polypeptide allelic variants or FGF-23 polypeptide splice variants, as defined herein, that have been chemically modified.

The term "mature FGF-23 polypeptide" refers to an FGF-23 polypeptide
5 lacking a leader sequence. A mature FGF-23 polypeptide may also include other modifications such as proteolytic processing of the amino-terminus (with or without a leader sequence) and/or the carboxyl-terminus, cleavage of a smaller polypeptide from a larger precursor, N-linked and/or O-linked glycosylation, and the like. An exemplary mature CHL polypeptide is depicted by the amino acid
10 sequence of SEQ ID NO: 3.

The term "FGF-23 fusion polypeptide" refers to a fusion of one or more amino acids (such as a heterologous protein or peptide) at the amino- or carboxyl-terminus of the polypeptide as set forth in SEQ ID NO: 2, FGF-23 polypeptide fragments, FGF-23 polypeptide orthologs, FGF-23 polypeptide variants, or FGF-
15 23 derivatives, as defined herein. The term "FGF-23 fusion polypeptide" also refers to a fusion of one or more amino acids at the amino- or carboxyl-terminus of the polypeptide encoded by FGF-23 polypeptide allelic variants or FGF-23 polypeptide splice variants, as defined herein.

The term "biologically active FGF-23 polypeptides" refers to FGF-23
20 polypeptides having at least one activity characteristic of the polypeptide comprising the amino acid sequence of SEQ ID NO: 2. In addition, an FGF-23 polypeptide may be active as an immunogen; that is, the FGF-23 polypeptide contains at least one epitope to which antibodies may be raised.

The term "isolated polypeptide" refers to a polypeptide of the present
25 invention that (1) has been separated from at least about 50 percent of polynucleotides, lipids, carbohydrates, or other materials with which it is naturally found when isolated from the source cell, (2) is not linked (by covalent or noncovalent interaction) to all or a portion of a polypeptide to which the "isolated polypeptide" is linked in nature, (3) is operably linked (by covalent or
30 noncovalent interaction) to a polypeptide with which it is not linked in nature, or (4) does not occur in nature. Preferably, the isolated polypeptide is substantially

free from any other contaminating polypeptides or other contaminants that are found in its natural environment that would interfere with its therapeutic, diagnostic, prophylactic or research use.

The term "identity," as known in the art, refers to a relationship between the sequences of two or more polypeptide molecules or two or more nucleic acid molecules, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between nucleic acid molecules or polypeptides, as the case may be, as determined by the match between strings of two or more nucleotide or two or more amino acid sequences. "Identity" measures the percent of identical matches between the smaller of two or more sequences with gap alignments (if any) addressed by a particular mathematical model or computer program (*i.e.*, "algorithms").

The term "similarity" is a related concept, but in contrast to "identity," "similarity" refers to a measure of relatedness which includes both identical matches and conservative substitution matches. If two polypeptide sequences have, for example, 10/20 identical amino acids, and the remainder are all non-conservative substitutions, then the percent identity and similarity would both be 50%. If in the same example, there are five more positions where there are conservative substitutions, then the percent identity remains 50%, but the percent similarity would be 75% (15/20). Therefore, in cases where there are conservative substitutions, the percent similarity between two polypeptides will be higher than the percent identity between those two polypeptides.

The term "naturally occurring" or "native" when used in connection with biological materials such as nucleic acid molecules, polypeptides, host cells, and the like, refers to materials which are found in nature and are not manipulated by man. Similarly, "non-naturally occurring" or "non-native" as used herein refers to a material that is not found in nature or that has been structurally modified or synthesized by man.

The terms "effective amount" and "therapeutically effective amount" each refer to the amount of an FGF-23 polypeptide or FGF-23 nucleic acid molecule

used to support an observable level of one or more biological activities of the FGF-23 polypeptides as set forth herein.

The term "pharmaceutically acceptable carrier" or "physiologically acceptable carrier" as used herein refers to one or more formulation materials
5 suitable for accomplishing or enhancing the delivery of the FGF-23 polypeptide, FGF-23 nucleic acid molecule, or FGF-23 selective binding agent as a pharmaceutical composition.

The term "antigen" refers to a molecule or a portion of a molecule capable of being bound by a selective binding agent, such as an antibody, and additionally
10 capable of being used in an animal to produce antibodies capable of binding to an epitope of that antigen. An antigen may have one or more epitopes.

The term "selective binding agent" refers to a molecule or molecules having specificity for an FGF-23 polypeptide. As used herein, the terms, "specific" and "specificity" refer to the ability of the selective binding agents to
15 bind to human FGF-23 polypeptides and not to bind to human non-FGF-23 polypeptides. It will be appreciated, however, that the selective binding agents may also bind orthologs of the polypeptide as set forth in SEQ ID NO: 2, that is, interspecies versions thereof, such as mouse and rat FGF-23 polypeptides.

The term "transduction" is used to refer to the transfer of genes from one
20 bacterium to another, usually by a phage. "Transduction" also refers to the acquisition and transfer of eukaryotic cellular sequences by retroviruses.

The term "transfection" is used to refer to the uptake of foreign or exogenous DNA by a cell, and a cell has been "transfected" when the exogenous DNA has been introduced inside the cell membrane. A number of transfection
25 techniques are well known in the art and are disclosed herein. *See, e.g., Graham et al., 1973, Virology 52:456; Sambrook et al., Molecular Cloning, A Laboratory Manual (Cold Spring Harbor Laboratories, 1989); Davis et al., Basic Methods in Molecular Biology (Elsevier, 1986); and Chu et al., 1981, Gene 13:197.* Such techniques can be used to introduce one or more exogenous DNA moieties into
30 suitable host cells.

The term "transformation" as used herein refers to a change in a cell's genetic characteristics, and a cell has been transformed when it has been modified to contain a new DNA. For example, a cell is transformed where it is genetically modified from its native state. Following transfection or transduction, the transforming DNA may recombine with that of the cell by physically integrating into a chromosome of the cell, may be maintained transiently as an episomal element without being replicated, or may replicate independently as a plasmid. A cell is considered to have been stably transformed when the DNA is replicated with the division of the cell.

10

Relatedness of Nucleic Acid Molecules and/or Polypeptides

It is understood that related nucleic acid molecules include allelic or splice variants of the nucleic acid molecule of SEQ ID NO: 1, and include sequences which are complementary to any of the above nucleotide sequences. Related nucleic acid molecules also include a nucleotide sequence encoding a polypeptide comprising or consisting essentially of a substitution, modification, addition and/or deletion of one or more amino acid residues compared to the polypeptide in SEQ ID NO: 2. Such related FGF-23 polypeptides may comprise, for example, an addition and/or a deletion of one or more N-linked or O-linked glycosylation sites or an addition and/or a deletion of one or more cysteine residues.

20

Related nucleic acid molecules also include fragments of FGF-23 nucleic acid molecules which encode a polypeptide of at least about 25 contiguous amino acids, or about 50 amino acids, or about 75 amino acids, or about 100 amino acids, or more than 100 amino acid residues of the FGF-23 polypeptide of SEQ ID NO: 2.

25

In addition, related FGF-23 nucleic acid molecules also include those molecules which comprise nucleotide sequences which hybridize under moderately or highly stringent conditions as defined herein with the fully complementary sequence of the FGF-23 nucleic acid molecule of SEQ ID NO: 1, or of a molecule encoding a polypeptide, which polypeptide comprises the amino acid sequence as shown in SEQ ID NO: 2, or of a nucleic acid fragment as

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defined herein, or of a nucleic acid fragment encoding a polypeptide as defined herein. Hybridization probes may be prepared using the FGF-23 sequences provided herein to screen cDNA, genomic or synthetic DNA libraries for related sequences. Regions of the DNA and/or amino acid sequence of FGF-23 polypeptide that exhibit significant identity to known sequences are readily determined using sequence alignment algorithms as described herein and those regions may be used to design probes for screening.

The term "highly stringent conditions" refers to those conditions that are designed to permit hybridization of DNA strands whose sequences are highly complementary, and to exclude hybridization of significantly mismatched DNAs. Hybridization stringency is principally determined by temperature, ionic strength, and the concentration of denaturing agents such as formamide. Examples of "highly stringent conditions" for hybridization and washing are 0.015 M sodium chloride, 0.0015 M sodium citrate at 65-68°C or 0.015 M sodium chloride, 0.0015 M sodium citrate, and 50% formamide at 42°C. See Sambrook, Fritsch & Maniatis, *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory, 1989); Anderson *et al.*, *Nucleic Acid Hybridisation: A Practical Approach* Ch. 4 (IRL Press Limited).

More stringent conditions (such as higher temperature, lower ionic strength, higher formamide, or other denaturing agent) may also be used – however, the rate of hybridization will be affected. Other agents may be included in the hybridization and washing buffers for the purpose of reducing non-specific and/or background hybridization. Examples are 0.1% bovine serum albumin, 0.1% polyvinyl-pyrrolidone, 0.1% sodium pyrophosphate, 0.1% sodium dodecylsulfate, NaDodSO₄, (SDS), ficoll, Denhardt's solution, sonicated salmon sperm DNA (or another non-complementary DNA), and dextran sulfate, although other suitable agents can also be used. The concentration and types of these additives can be changed without substantially affecting the stringency of the hybridization conditions. Hybridization experiments are usually carried out at pH 6.8-7.4; however, at typical ionic strength conditions, the rate of hybridization is

nearly independent of pH. See Anderson *et al.*, *Nucleic Acid Hybridisation: A Practical Approach* Ch. 4 (IRL Press Limited).

Factors affecting the stability of DNA duplex include base composition, length, and degree of base pair mismatch. Hybridization conditions can be adjusted by one skilled in the art in order to accommodate these variables and allow DNAs of different sequence relatedness to form hybrids. The melting temperature of a perfectly matched DNA duplex can be estimated by the following equation:

$$T_m(^{\circ}\text{C}) = 81.5 + 16.6(\log[\text{Na}^+]) + 0.41(\% \text{G+C}) - 600/N - 0.72(\% \text{formamide})$$

where N is the length of the duplex formed, [Na⁺] is the molar concentration of the sodium ion in the hybridization or washing solution, %G+C is the percentage of (guanine+cytosine) bases in the hybrid. For imperfectly matched hybrids, the melting temperature is reduced by approximately 1°C for each 1% mismatch.

The term "moderately stringent conditions" refers to conditions under which a DNA duplex with a greater degree of base pair mismatching than could occur under "highly stringent conditions" is able to form. Examples of typical "moderately stringent conditions" are 0.015 M sodium chloride, 0.0015 M sodium citrate at 50-65°C or 0.015 M sodium chloride, 0.0015 M sodium citrate, and 20% formamide at 37-50°C. By way of example, "moderately stringent conditions" of 50°C in 0.015 M sodium ion will allow about a 21% mismatch.

It will be appreciated by those skilled in the art that there is no absolute distinction between "highly stringent conditions" and "moderately stringent conditions." For example, at 0.015 M sodium ion (no formamide), the melting temperature of perfectly matched long DNA is about 71°C. With a wash at 65°C (at the same ionic strength), this would allow for approximately a 6% mismatch. To capture more distantly related sequences, one skilled in the art can simply lower the temperature or raise the ionic strength.

A good estimate of the melting temperature in 1M NaCl* for oligonucleotide probes up to about 20nt is given by:

$$T_m = 2^{\circ}\text{C per A-T base pair} + 4^{\circ}\text{C per G-C base pair}$$

*The sodium ion concentration in 6X salt sodium citrate (SSC) is 1M. *See Suggs et al., Developmental Biology Using Purified Genes* 683 (Brown and Fox, eds., 1981).

High stringency washing conditions for oligonucleotides are usually at a
5 temperature of 0-5°C below the T_m of the oligonucleotide in 6X SSC, 0.1% SDS.

In another embodiment, related nucleic acid molecules comprise or consist of a nucleotide sequence that is at least about 70 percent identical to the nucleotide sequence as shown in SEQ ID NO: 1, or comprise or consist essentially of a nucleotide sequence encoding a polypeptide that is at least about 70 percent
10 identical to the polypeptide as set forth in SEQ ID NO: 2. In preferred embodiments, the nucleotide sequences are about 75 percent, or about 80 percent, or about 85 percent, or about 90 percent, or about 95, 96, 97, 98, or 99 percent identical to the nucleotide sequence as shown in SEQ ID NO: 1, or the nucleotide sequences encode a polypeptide that is about 75 percent, or about 80 percent, or
15 about 85 percent, or about 90 percent, or about 95, 96, 97, 98, or 99 percent identical to the polypeptide sequence as set forth in SEQ ID NO: 2. Related nucleic acid molecules encode polypeptides possessing at least one activity of the polypeptide set forth in SEQ ID NO: 2.

Differences in the nucleic acid sequence may result in conservative and/or
20 non-conservative modifications of the amino acid sequence relative to the amino acid sequence of SEQ ID NO: 2.

Conservative modifications to the amino acid sequence of SEQ ID NO: 2 (and the corresponding modifications to the encoding nucleotides) will produce a polypeptide having functional and chemical characteristics similar to those of
25 FGF-23 polypeptides. In contrast, substantial modifications in the functional and/or chemical characteristics of FGF-23 polypeptides may be accomplished by selecting substitutions in the amino acid sequence of SEQ ID NO: 2 that differ significantly in their effect on maintaining (a) the structure of the molecular backbone in the area of the substitution, for example, as a sheet or helical
30 conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain.

For example, a “conservative amino acid substitution” may involve a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. Furthermore, any native residue in the polypeptide may also be substituted with alanine, as has been previously described for “alanine scanning mutagenesis.”

Conservative amino acid substitutions also encompass non-naturally occurring amino acid residues that are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include peptidomimetics, and other reversed or inverted forms of amino acid moieties.

Naturally occurring residues may be divided into classes based on common side chain properties:

- 1) hydrophobic: norleucine, Met, Ala, Val, Leu, Ile;
- 2) neutral hydrophilic: Cys, Ser, Thr;
- 3) acidic: Asp, Glu;
- 4) basic: Asn, Gln, His, Lys, Arg;
- 5) residues that influence chain orientation: Gly, Pro; and
- 6) aromatic: Trp, Tyr, Phe.

For example, non-conservative substitutions may involve the exchange of a member of one of these classes for a member from another class. Such substituted residues may be introduced into regions of the human FGF-23 polypeptide that are homologous with non-human FGF-23 polypeptides, or into the non-homologous regions of the molecule.

In making such changes, the hydropathic index of amino acids may be considered. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics. The hydropathic indices are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

The importance of the hydropathic amino acid index in conferring interactive biological function on a protein is generally understood in the art (Kyte *et al.*, 1982, *J. Mol. Biol.* 157:105-31). It is known that certain amino acids may be substituted for other amino acids having a similar hydropathic index or score and still retain a similar biological activity. In making changes based upon the hydropathic index, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity, particularly where the biologically functionally equivalent protein or peptide thereby created is intended for use in immunological embodiments, as in the present case. The greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with its immunogenicity and antigenicity, *i.e.*, with a biological property of the protein.

The following hydrophilicity values have been assigned to these amino acid residues: arginine (+3.0); lysine (+3.0); aspartate ($+3.0 \pm 1$); glutamate ($+3.0 \pm 1$); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 ± 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); and tryptophan (-3.4). In making changes based upon similar hydrophilicity values, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. One may also identify epitopes from primary amino acid sequences on the basis of hydrophilicity. These regions are also referred to as "epitopic core regions."

Desired amino acid substitutions (whether conservative or non-conservative) can be determined by those skilled in the art at the time such substitutions are desired. For example, amino acid substitutions can be used to identify important residues of the FGF-23 polypeptide, or to increase or decrease

the affinity of the FGF-23 polypeptides described herein. Exemplary amino acid substitutions are set forth in Table I.

Table I

5

Amino Acid Substitutions

Original Residues	Exemplary Substitutions	Preferred Substitutions
Ala	Val, Leu, Ile	Val
Arg	Lys, Gln, Asn	Lys
Asn	Gln	Gln
Asp	Glu	Glu
Cys	Ser, Ala	Ser
Gln	Asn	Asn
Glu	Asp	Asp
Gly	Pro, Ala	Ala
His	Asn, Gln, Lys, Arg	Arg
Ile	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys	Arg, 1,4 Diamino-butyrlic Acid, Gln, Asn	Arg
Met	Leu, Phe, Ile	Leu
Phe	Leu, Val, Ile, Ala, Tyr	Leu
Pro	Ala	Gly
Ser	Thr, Ala, Cys	Thr
Thr	Ser	Ser
Trp	Tyr, Phe	Tyr
Tyr	Trp, Phe, Thr, Ser	Phe

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Val	Ile, Met, Leu, Phe, Ala, Norleucine	Leu
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A skilled artisan will be able to determine suitable variants of the polypeptide as set forth in SEQ ID NO: 2 using well-known techniques. For identifying suitable areas of the molecule that may be changed without destroying biological activity, one skilled in the art may target areas not believed to be important for activity. For example, when similar polypeptides with similar activities from the same species or from other species are known, one skilled in the art may compare the amino acid sequence of an FGF-23 polypeptide to such similar polypeptides. With such a comparison, one can identify residues and portions of the molecules that are conserved among similar polypeptides. It will be appreciated that changes in areas of the FGF-23 molecule that are not conserved relative to such similar polypeptides would be less likely to adversely affect the biological activity and/or structure of an FGF-23 polypeptide. One skilled in the art would also know that, even in relatively conserved regions, one may substitute chemically similar amino acids for the naturally occurring residues while retaining activity (conservative amino acid residue substitutions). Therefore, even areas that may be important for biological activity or for structure may be subject to conservative amino acid substitutions without destroying the biological activity or without adversely affecting the polypeptide structure.

Additionally, one skilled in the art can review structure-function studies identifying residues in similar polypeptides that are important for activity or structure. In view of such a comparison, one can predict the importance of amino acid residues in an FGF-23 polypeptide that correspond to amino acid residues that are important for activity or structure in similar polypeptides. One skilled in the art may opt for chemically similar amino acid substitutions for such predicted important amino acid residues of FGF-23 polypeptides.

One skilled in the art can also analyze the three-dimensional structure and amino acid sequence in relation to that structure in similar polypeptides. In view of such information, one skilled in the art may predict the alignment of amino acid

residues of FGF-23 polypeptide with respect to its three dimensional structure. One skilled in the art may choose not to make radical changes to amino acid residues predicted to be on the surface of the protein, since such residues may be involved in important interactions with other molecules. Moreover, one skilled in the art may generate test variants containing a single amino acid substitution at each amino acid residue. The variants could be screened using activity assays known to those with skill in the art. Such variants could be used to gather information about suitable variants. For example, if one discovered that a change to a particular amino acid residue resulted in destroyed, undesirably reduced, or unsuitable activity, variants with such a change would be avoided. In other words, based on information gathered from such routine experiments, one skilled in the art can readily determine the amino acids where further substitutions should be avoided either alone or in combination with other mutations.

A number of scientific publications have been devoted to the prediction of secondary structure. See Moulton, 1996, *Curr. Opin. Biotechnol.* 7:422-27; Chou *et al.*, 1974, *Biochemistry* 13:222-45; Chou *et al.*, 1974, *Biochemistry* 113:211-22; Chou *et al.*, 1978, *Adv. Enzymol. Relat. Areas Mol. Biol.* 47:45-48; Chou *et al.*, 1978, *Ann. Rev. Biochem.* 47:251-276; and Chou *et al.*, 1979, *Biophys. J.* 26:367-84. Moreover, computer programs are currently available to assist with predicting secondary structure. One method of predicting secondary structure is based upon homology modeling. For example, two polypeptides or proteins which have a sequence identity of greater than 30%, or similarity greater than 40%, often have similar structural topologies. The recent growth of the protein structural database (PDB) has provided enhanced predictability of secondary structure, including the potential number of folds within the structure of a polypeptide or protein. See Holm *et al.*, 1999, *Nucleic Acids Res.* 27:244-47. It has been suggested that there are a limited number of folds in a given polypeptide or protein and that once a critical number of structures have been resolved, structural prediction will become dramatically more accurate (Brenner *et al.*, 1997, *Curr. Opin. Struct. Biol.* 7:369-76).

Additional methods of predicting secondary structure include "threading" (Jones, 1997, *Curr. Opin. Struct. Biol.* 7:377-87; Sippl *et al.*, 1996, *Structure* 4:15-19), "profile analysis" (Bowie *et al.*, 1991, *Science*, 253:164-70; Gribskov *et al.*, 1990, *Methods Enzymol.* 183:146-59; Gribskov *et al.*, 1987, *Proc. Nat. Acad. Sci. U.S.A.* 84:4355-58), and "evolutionary linkage" (See Holm *et al.*, *supra*, and Brenner *et al.*, *supra*).

Preferred FGF-23 polypeptide variants include glycosylation variants wherein the number and/or type of glycosylation sites have been altered compared to the amino acid sequence set forth in SEQ ID NO: 2. In one embodiment, FGF-23 polypeptide variants comprise a greater or a lesser number of N-linked glycosylation sites than the amino acid sequence set forth in SEQ ID NO: 2. An N-linked glycosylation site is characterized by the sequence: Asn-X-Ser or Asn-X-Thr, wherein the amino acid residue designated as X may be any amino acid residue except proline. The substitution of amino acid residues to create this sequence provides a potential new site for the addition of an N-linked carbohydrate chain. Alternatively, substitutions that eliminate this sequence will remove an existing N-linked carbohydrate chain. Also provided is a rearrangement of N-linked carbohydrate chains wherein one or more N-linked glycosylation sites (typically those that are naturally occurring) are eliminated and one or more new N-linked sites are created. Additional preferred FGF-23 variants include cysteine variants, wherein one or more cysteine residues are deleted or substituted with another amino acid (*e.g.*, serine) as compared to the amino acid sequence set forth in SEQ ID NO: 2. Cysteine variants are useful when FGF-23 polypeptides must be refolded into a biologically active conformation such as after the isolation of insoluble inclusion bodies. Cysteine variants generally have fewer cysteine residues than the native protein, and typically have an even number to minimize interactions resulting from unpaired cysteines.

In other embodiments, related nucleic acid molecules comprise or consist of a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid insertion and wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2, or a nucleotide sequence encoding a

polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid deletion and wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2. Related nucleic acid molecules also comprise or consist of a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 wherein the polypeptide has a carboxyl- and/or amino-terminal truncation and further wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2. Related nucleic acid molecules also comprise or consist of a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one modification selected from the group consisting of amino acid substitutions, amino acid insertions, amino acid deletions, carboxyl-terminal truncations, and amino-terminal truncations and wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2.

In addition, the polypeptide comprising the amino acid sequence of SEQ ID NO: 2, or other FGF-23 polypeptide, may be fused to a homologous polypeptide to form a homodimer or to a heterologous polypeptide to form a heterodimer. Heterologous peptides and polypeptides include, but are not limited to: an epitope to allow for the detection and/or isolation of an FGF-23 fusion polypeptide; a transmembrane receptor protein or a portion thereof, such as an extracellular domain or a transmembrane and intracellular domain; a ligand or a portion thereof which binds to a transmembrane receptor protein; an enzyme or portion thereof which is catalytically active; a polypeptide or peptide which promotes oligomerization, such as a leucine zipper domain; a polypeptide or peptide which increases stability, such as an immunoglobulin constant region; and a polypeptide which has a therapeutic activity different from the polypeptide comprising the amino acid sequence as set forth in SEQ ID NO: 2, or other FGF-23 polypeptide.

Fusions can be made either at the amino-terminus or at the carboxyl-terminus of the polypeptide comprising the amino acid sequence set forth in SEQ ID NO: 2, or other FGF-23 polypeptide. Fusions may be direct with no linker or adapter molecule or may be through a linker or adapter molecule. A linker or adapter molecule may be one or more amino acid residues, typically from about

20 to about 50 amino acid residues. A linker or adapter molecule may also be designed with a cleavage site for a DNA restriction endonuclease or for a protease to allow for the separation of the fused moieties. It will be appreciated that once constructed, the fusion polypeptides can be derivatized according to the methods
5 described herein.

In a further embodiment of the invention, the polypeptide comprising the amino acid sequence of SEQ ID NO: 2, or other FGF-23 polypeptide, is fused to one or more domains of an Fc region of human IgG. Antibodies comprise two functionally independent parts, a variable domain known as "Fab," that binds an
10 antigen, and a constant domain known as "Fc," that is involved in effector functions such as complement activation and attack by phagocytic cells. An Fc has a long serum half-life, whereas an Fab is short-lived. Capon *et al.*, 1989, *Nature* 337:525-31. When constructed together with a therapeutic protein, an Fc domain can provide longer half-life or incorporate such functions as Fc receptor
15 binding, protein A binding, complement fixation, and perhaps even placental transfer. *Id.* Table II summarizes the use of certain Fc fusions known in the art.

Table II
Fc Fusion with Therapeutic Proteins

Form of Fc	Fusion partner	Therapeutic implications	Reference
IgG1	N-terminus of CD30-L	Hodgkin's disease; anaplastic lymphoma; T-cell leukemia	U.S. Patent No. 5,480,981
Murine Fcγ2a	IL-10	anti-inflammatory; transplant rejection	Zheng <i>et al.</i> , 1995, <i>J. Immunol.</i> 154:5590-600
IgG1	TNF receptor	septic shock	Fisher <i>et al.</i> , 1996, <i>N. Engl. J. Med.</i> 334:1697-1702; Van Zee <i>et al.</i> , 1996, <i>J. Immunol.</i> 156:2221-30
IgG, IgA, IgM, or IgE (excluding the first domain)	TNF receptor	inflammation, autoimmune disorders	U.S. Patent No. 5,808,029
IgG1	CD4 receptor	AIDS	Capon <i>et al.</i> , 1989, <i>Nature</i> 337: 525-31

IgG1, IgG3	N-terminus of IL-2	anti-cancer, antiviral	Harvill <i>et al.</i> , 1995, <i>Immunotech.</i> 1:95-105
IgG1	C-terminus of OPG	osteoarthritis; bone density	WO 97/23614
IgG1	N-terminus of leptin	anti-obesity	PCT/US 97/23183, filed December 11, 1997
Human Ig C γ 1	CTLA-4	autoimmune disorders	Linsley, 1991, <i>J. Exp. Med.</i> , 174:561-69

In one example, a human IgG hinge, CH2, and CH3 region may be fused at either the amino-terminus or carboxyl-terminus of the FGF-23 polypeptides using methods known to the skilled artisan. In another example, a human IgG
5 hinge, CH2, and CH3 region may be fused at either the amino-terminus or carboxyl-terminus of an FGF-23 polypeptide fragment (*e.g.*, the predicted extracellular portion of FGF-23 polypeptide).

The resulting FGF-23 fusion polypeptide may be purified by use of a Protein A affinity column. Peptides and proteins fused to an Fc region have been
10 found to exhibit a substantially greater half-life *in vivo* than the unfused counterpart. Also, a fusion to an Fc region allows for dimerization/multimerization of the fusion polypeptide. The Fc region may be a naturally occurring Fc region, or may be altered to improve certain qualities, such as therapeutic qualities, circulation time, or reduced aggregation.

15 Identity and similarity of related nucleic acid molecules and polypeptides are readily calculated by known methods. Such methods include, but are not limited to those described in *Computational Molecular Biology* (A.M. Lesk, ed., Oxford University Press 1988); *Biocomputing: Informatics and Genome Projects* (D.W. Smith, ed., Academic Press 1993); *Computer Analysis of Sequence Data*
20 (Part 1, A.M. Griffin and H.G. Griffin, eds., Humana Press 1994); G. von Heinle, *Sequence Analysis in Molecular Biology* (Academic Press 1987); *Sequence Analysis Primer* (M. Gribskov and J. Devereux, eds., M. Stockton Press 1991); and Carillo *et al.*, 1988, *SIAM J. Applied Math.*, 48:1073.

Preferred methods to determine identity and/or similarity are designed to
25 give the largest match between the sequences tested. Methods to determine

identity and similarity are described in publicly available computer programs. Preferred computer program methods to determine identity and similarity between two sequences include, but are not limited to, the GCG program package, including GAP (Devereux *et al.*, 1984, *Nucleic Acids Res.* 12:387; Genetics
5 Computer Group, University of Wisconsin, Madison, WI), BLASTP, BLASTN, and FASTA (Altschul *et al.*, 1990, *J. Mol. Biol.* 215:403-10). The BLASTX program is publicly available from the National Center for Biotechnology Information (NCBI) and other sources (Altschul *et al.*, *BLAST Manual* (NCB NLM NIH, Bethesda, MD); Altschul *et al.*, 1990, *supra*). The well-known Smith
10 Waterman algorithm may also be used to determine identity.

Certain alignment schemes for aligning two amino acid sequences may result in the matching of only a short region of the two sequences, and this small aligned region may have very high sequence identity even though there is no significant relationship between the two full-length sequences. Accordingly, in a
15 preferred embodiment, the selected alignment method (GAP program) will result in an alignment that spans at least 50 contiguous amino acids of the claimed polypeptide.

For example, using the computer algorithm GAP (Genetics Computer Group, University of Wisconsin, Madison, WI), two polypeptides for which the
20 percent sequence identity is to be determined are aligned for optimal matching of their respective amino acids (the "matched span," as determined by the algorithm). A gap opening penalty (which is calculated as 3X the average diagonal; the "average diagonal" is the average of the diagonal of the comparison matrix being used; the "diagonal" is the score or number assigned to each perfect
25 amino acid match by the particular comparison matrix) and a gap extension penalty (which is usually 0.1X the gap opening penalty), as well as a comparison matrix such as PAM 250 or BLOSUM 62 are used in conjunction with the algorithm. A standard comparison matrix is also used by the algorithm (*see* Dayhoff *et al.*, 5 *Atlas of Protein Sequence and Structure* (Supp. 3
30 1978)(PAM250 comparison matrix); Henikoff *et al.*, 1992, *Proc. Natl. Acad. Sci USA* 89:10915-19 (BLOSUM 62 comparison matrix)).

Preferred parameters for polypeptide sequence comparison include the following:

- 5 Algorithm: Needleman and Wunsch, 1970, *J. Mol. Biol.* 48:443-53;
Comparison matrix: BLOSUM 62 (Henikoff *et al.*, *supra*);
Gap Penalty: 12
Gap Length Penalty: 4
Threshold of Similarity: 0

- 10 The GAP program is useful with the above parameters. The aforementioned parameters are the default parameters for polypeptide comparisons (along with no penalty for end gaps) using the GAP algorithm.

Preferred parameters for nucleic acid molecule sequence comparison include the following:

- 15 Algorithm: Needleman and Wunsch, *supra*;
Comparison matrix: matches = +10, mismatch = 0
Gap Penalty: 50
Gap Length Penalty: 3

20

The GAP program is also useful with the above parameters. The aforementioned parameters are the default parameters for nucleic acid molecule comparisons.

- Other exemplary algorithms, gap opening penalties, gap extension penalties, comparison matrices, and thresholds of similarity may be used, including those set forth in the Program Manual, Wisconsin Package, Version 9, September, 1997. The particular choices to be made will be apparent to those of skill in the art and will depend on the specific comparison to be made, such as DNA-to-DNA, protein-to-protein, protein-to-DNA; and additionally, whether the comparison is between given pairs of sequences (in which case GAP or BestFit
25 are generally preferred) or between one sequence and a large database of sequences (in which case FASTA or BLASTA are preferred).
30

Nucleic Acid Molecules

The nucleic acid molecules encoding a polypeptide comprising the amino acid sequence of an FGF-23 polypeptide can readily be obtained in a variety of ways including, without limitation, chemical synthesis, cDNA or genomic library
5 screening, expression library screening, and/or PCR amplification of cDNA.

Recombinant DNA methods used herein are generally those set forth in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (Cold Spring Harbor Laboratory Press, 1989) and/or *Current Protocols in Molecular Biology* (Ausubel
10 *et al.*, eds., Green Publishers Inc. and Wiley and Sons 1994). The invention provides for nucleic acid molecules as described herein and methods for obtaining such molecules.

Where a gene encoding the amino acid sequence of an FGF-23 polypeptide has been identified from one species, all or a portion of that gene may
15 be used as a probe to identify orthologs or related genes from the same species. The probes or primers may be used to screen cDNA libraries from various tissue sources believed to express the FGF-23 polypeptide. In addition, part or all of a nucleic acid molecule having the sequence as set forth in SEQ ID NO: 1 may be used to screen a genomic library to identify and isolate a gene encoding the amino
20 acid sequence of an FGF-23 polypeptide. Typically, conditions of moderate or high stringency will be employed for screening to minimize the number of false positives obtained from the screening.

Nucleic acid molecules encoding the amino acid sequence of FGF-23 polypeptides may also be identified by expression cloning which employs the
25 detection of positive clones based upon a property of the expressed protein. Typically, nucleic acid libraries are screened by the binding an antibody or other binding partner (*e.g.*, receptor or ligand) to cloned proteins that are expressed and displayed on a host cell surface. The antibody or binding partner is modified with a detectable label to identify those cells expressing the desired clone.

30 Recombinant expression techniques conducted in accordance with the descriptions set forth below may be followed to produce these polynucleotides

and to express the encoded polypeptides. For example, by inserting a nucleic acid sequence that encodes the amino acid sequence of an FGF-23 polypeptide into an appropriate vector, one skilled in the art can readily produce large quantities of the desired nucleotide sequence. The sequences can then be used to generate
5 detection probes or amplification primers. Alternatively, a polynucleotide encoding the amino acid sequence of an FGF-23 polypeptide can be inserted into an expression vector. By introducing the expression vector into an appropriate host, the encoded FGF-23 polypeptide may be produced in large amounts.

Another method for obtaining a suitable nucleic acid sequence is the
10 polymerase chain reaction (PCR). In this method, cDNA is prepared from poly(A)+RNA or total RNA using the enzyme reverse transcriptase. Two primers, typically complementary to two separate regions of cDNA encoding the amino acid sequence of an FGF-23 polypeptide, are then added to the cDNA along with a polymerase such as *Taq* polymerase, and the polymerase amplifies
15 the cDNA region between the two primers.

Another means of preparing a nucleic acid molecule encoding the amino acid sequence of an FGF-23 polypeptide is chemical synthesis using methods well known to the skilled artisan such as those described by Engels *et al.*, 1989, *Angew. Chem. Intl. Ed.* 28:716-34. These methods include, *inter alia*, the
20 phosphotriester, phosphoramidite, and H-phosphonate methods for nucleic acid synthesis. A preferred method for such chemical synthesis is polymer-supported synthesis using standard phosphoramidite chemistry. Typically, the DNA encoding the amino acid sequence of an FGF-23 polypeptide will be several hundred nucleotides in length. Nucleic acids larger than about 100 nucleotides
25 can be synthesized as several fragments using these methods. The fragments can then be ligated together to form the full-length nucleotide sequence of an FGF-23 gene. Usually, the DNA fragment encoding the amino-terminus of the polypeptide will have an ATG, which encodes a methionine residue. This methionine may or may not be present on the mature form of the FGF-23
30 polypeptide, depending on whether the polypeptide produced in the host cell is

designed to be secreted from that cell. Other methods known to the skilled artisan may be used as well.

In certain embodiments, nucleic acid variants contain codons which have been altered for optimal expression of an FGF-23 polypeptide in a given host cell. Particular codon alterations will depend upon the FGF-23 polypeptide and host cell selected for expression. Such "codon optimization" can be carried out by a variety of methods, for example, by selecting codons which are preferred for use in highly expressed genes in a given host cell. Computer algorithms which incorporate codon frequency tables such as "Eco_high.Cod" for codon preference of highly expressed bacterial genes may be used and are provided by the University of Wisconsin Package Version 9.0 (Genetics Computer Group, Madison, WI). Other useful codon frequency tables include "Celegans_high.cod," "Celegans_low.cod," "Drosophila_high.cod," "Human_high.cod," "Maize_high.cod," and "Yeast_high.cod."

In some cases, it may be desirable to prepare nucleic acid molecules encoding FGF-23 polypeptide variants. Nucleic acid molecules encoding variants may be produced using site directed mutagenesis, PCR amplification, or other appropriate methods, where the primer(s) have the desired point mutations (*see* Sambrook *et al.*, *supra*, and Ausubel *et al.*, *supra*, for descriptions of mutagenesis techniques). Chemical synthesis using methods described by Engels *et al.*, *supra*, may also be used to prepare such variants. Other methods known to the skilled artisan may be used as well.

Vectors and Host Cells

A nucleic acid molecule encoding the amino acid sequence of an FGF-23 polypeptide is inserted into an appropriate expression vector using standard ligation techniques. The vector is typically selected to be functional in the particular host cell employed (*i.e.*, the vector is compatible with the host cell machinery such that amplification of the gene and/or expression of the gene can occur). A nucleic acid molecule encoding the amino acid sequence of an FGF-23 polypeptide may be amplified/expressed in prokaryotic, yeast, insect (baculovirus

systems) and/or eukaryotic host cells. Selection of the host cell will depend in part on whether an FGF-23 polypeptide is to be post-translationally modified (e.g., glycosylated and/or phosphorylated). If so, yeast, insect, or mammalian host cells are preferable. For a review of expression vectors, see *Meth. Enz.*, vol. 5 185 (D.V. Goeddel, ed., Academic Press 1990).

Typically, expression vectors used in any of the host cells will contain sequences for plasmid maintenance and for cloning and expression of exogenous nucleotide sequences. Such sequences, collectively referred to as "flanking sequences" in certain embodiments will typically include one or more of the 10 following nucleotide sequences: a promoter, one or more enhancer sequences, an origin of replication, a transcriptional termination sequence, a complete intron sequence containing a donor and acceptor splice site, a sequence encoding a leader sequence for polypeptide secretion, a ribosome binding site, a polyadenylation sequence, a polylinker region for inserting the nucleic acid 15 encoding the polypeptide to be expressed, and a selectable marker element. Each of these sequences is discussed below.

Optionally, the vector may contain a "tag"-encoding sequence, i.e., an oligonucleotide molecule located at the 5' or 3' end of the FGF-23 polypeptide coding sequence; the oligonucleotide sequence encodes polyHis (such as 20 hexaHis), or another "tag" such as FLAG, HA (hemagglutinin influenza virus), or *myc* for which commercially available antibodies exist. This tag is typically fused to the polypeptide upon expression of the polypeptide, and can serve as a means for affinity purification of the FGF-23 polypeptide from the host cell. Affinity purification can be accomplished, for example, by column chromatography using 25 antibodies against the tag as an affinity matrix. Optionally, the tag can subsequently be removed from the purified FGF-23 polypeptide by various means such as using certain peptidases for cleavage.

Flanking sequences may be homologous (i.e., from the same species and/or strain as the host cell), heterologous (i.e., from a species other than the host 30 cell species or strain), hybrid (i.e., a combination of flanking sequences from more than one source), or synthetic, or the flanking sequences may be native

sequences which normally function to regulate FGF-23 polypeptide expression. As such, the source of a flanking sequence may be any prokaryotic or eukaryotic organism, any vertebrate or invertebrate organism, or any plant, provided that the flanking sequence is functional in, and can be activated by, the host cell
5 machinery.

Flanking sequences useful in the vectors of this invention may be obtained by any of several methods well known in the art. Typically, flanking sequences useful herein – other than the FGF-23 gene flanking sequences – will have been previously identified by mapping and/or by restriction endonuclease digestion and
10 can thus be isolated from the proper tissue source using the appropriate restriction endonucleases. In some cases, the full nucleotide sequence of a flanking sequence may be known. Here, the flanking sequence may be synthesized using the methods described herein for nucleic acid synthesis or cloning.

Where all or only a portion of the flanking sequence is known, it may be
15 obtained using PCR and/or by screening a genomic library with a suitable oligonucleotide and/or flanking sequence fragment from the same or another species. Where the flanking sequence is not known, a fragment of DNA containing a flanking sequence may be isolated from a larger piece of DNA that may contain, for example, a coding sequence or even another gene or genes.
20 Isolation may be accomplished by restriction endonuclease digestion to produce the proper DNA fragment followed by isolation using agarose gel purification, Qiagen® column chromatography (Chatsworth, CA), or other methods known to the skilled artisan. The selection of suitable enzymes to accomplish this purpose will be readily apparent to one of ordinary skill in the art.

25 An origin of replication is typically a part of those prokaryotic expression vectors purchased commercially, and the origin aids in the amplification of the vector in a host cell. Amplification of the vector to a certain copy number can, in some cases, be important for the optimal expression of an FGF-23 polypeptide. If the vector of choice does not contain an origin of replication site, one may be
30 chemically synthesized based on a known sequence, and ligated into the vector. For example, the origin of replication from the plasmid pBR322 (New England

Biolabs, Beverly, MA) is suitable for most gram-negative bacteria and various origins (*e.g.*, SV40, polyoma, adenovirus, vesicular stomatitis virus (VSV), or papillomaviruses such as HPV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed
5 for mammalian expression vectors (for example, the SV40 origin is often used only because it contains the early promoter).

A transcription termination sequence is typically located 3' of the end of a polypeptide coding region and serves to terminate transcription. Usually, a transcription termination sequence in prokaryotic cells is a G-C rich fragment
10 followed by a poly-T sequence. While the sequence is easily cloned from a library or even purchased commercially as part of a vector, it can also be readily synthesized using methods for nucleic acid synthesis such as those described herein.

A selectable marker gene element encodes a protein necessary for the
15 survival and growth of a host cell grown in a selective culture medium. Typical selection marker genes encode proteins that (a) confer resistance to antibiotics or other toxins, *e.g.*, ampicillin, tetracycline, or kanamycin for prokaryotic host cells; (b) complement auxotrophic deficiencies of the cell; or (c) supply critical nutrients not available from complex media. Preferred selectable markers are the
20 kanamycin resistance gene, the ampicillin resistance gene, and the tetracycline resistance gene. A neomycin resistance gene may also be used for selection in prokaryotic and eukaryotic host cells.

Other selection genes may be used to amplify the gene that will be expressed. Amplification is the process wherein genes that are in greater demand
25 for the production of a protein critical for growth are reiterated in tandem within the chromosomes of successive generations of recombinant cells. Examples of suitable selectable markers for mammalian cells include dihydrofolate reductase (DHFR) and thymidine kinase. The mammalian cell transformants are placed under selection pressure wherein only the transformants are uniquely adapted to
30 survive by virtue of the selection gene present in the vector. Selection pressure is imposed by culturing the transformed cells under conditions in which the

concentration of selection agent in the medium is successively changed, thereby leading to the amplification of both the selection gene and the DNA that encodes an FGF-23 polypeptide. As a result, increased quantities of FGF-23 polypeptide are synthesized from the amplified DNA.

5 A ribosome binding site is usually necessary for translation initiation of mRNA and is characterized by a Shine-Dalgarno sequence (prokaryotes) or a Kozak sequence (eukaryotes). The element is typically located 3' to the promoter and 5' to the coding sequence of an FGF-23 polypeptide to be expressed. The Shine-Dalgarno sequence is varied but is typically a polypurine (*i.e.*, having a
10 high A-G content). Many Shine-Dalgarno sequences have been identified, each of which can be readily synthesized using methods set forth herein and used in a prokaryotic vector.

 A leader, or signal, sequence may be used to direct an FGF-23 polypeptide out of the host cell. Typically, a nucleotide sequence encoding the signal
15 sequence is positioned in the coding region of an FGF-23 nucleic acid molecule, or directly at the 5' end of an FGF-23 polypeptide coding region. Many signal sequences have been identified, and any of those that are functional in the selected host cell may be used in conjunction with an FGF-23 nucleic acid molecule. Therefore, a signal sequence may be homologous (naturally occurring) or
20 heterologous to the FGF-23 nucleic acid molecule. Additionally, a signal sequence may be chemically synthesized using methods described herein. In most cases, the secretion of an FGF-23 polypeptide from the host cell via the presence of a signal peptide will result in the removal of the signal peptide from the secreted FGF-23 polypeptide. The signal sequence may be a component of
25 the vector, or it may be a part of an FGF-23 nucleic acid molecule that is inserted into the vector.

 Included within the scope of this invention is the use of either a nucleotide sequence encoding a native FGF-23 polypeptide signal sequence joined to an FGF-23 polypeptide coding region or a nucleotide sequence encoding a
30 heterologous signal sequence joined to an FGF-23 polypeptide coding region. The heterologous signal sequence selected should be one that is recognized and

processed, *i.e.*, cleaved by a signal peptidase, by the host cell. For prokaryotic host cells that do not recognize and process the native FGF-23 polypeptide signal sequence, the signal sequence is substituted by a prokaryotic signal sequence selected, for example, from the group of the alkaline phosphatase, penicillinase, or heat-stable enterotoxin II leaders. For yeast secretion, the native FGF-23 polypeptide signal sequence may be substituted by the yeast invertase, alpha factor, or acid phosphatase leaders. In mammalian cell expression the native signal sequence is satisfactory, although other mammalian signal sequences may be suitable.

10 In some cases, such as where glycosylation is desired in a eukaryotic host cell expression system, one may manipulate the various presequences to improve glycosylation or yield. For example, one may alter the peptidase cleavage site of a particular signal peptide, or add pro-sequences, which also may affect glycosylation. The final protein product may have, in the -1 position (relative to
15 the first amino acid of the mature protein) one or more additional amino acids incident to expression, which may not have been totally removed. For example, the final protein product may have one or two amino acid residues found in the peptidase cleavage site, attached to the amino-terminus. Alternatively, use of some enzyme cleavage sites may result in a slightly truncated form of the desired
20 FGF-23 polypeptide, if the enzyme cuts at such area within the mature polypeptide.

In many cases, transcription of a nucleic acid molecule is increased by the presence of one or more introns in the vector; this is particularly true where a polypeptide is produced in eukaryotic host cells, especially mammalian host cells.
25 The introns used may be naturally occurring within the FGF-23 gene especially where the gene used is a full-length genomic sequence or a fragment thereof. Where the intron is not naturally occurring within the gene (as for most cDNAs), the intron may be obtained from another source. The position of the intron with respect to flanking sequences and the FGF-23 gene is generally important, as the
30 intron must be transcribed to be effective. Thus, when an FGF-23 cDNA molecule is being transcribed, the preferred position for the intron is 3' to the

transcription start site and 5' to the poly-A transcription termination sequence. Preferably, the intron or introns will be located on one side or the other (*i.e.*, 5' or 3') of the cDNA such that it does not interrupt the coding sequence. Any intron from any source, including viral, prokaryotic and eukaryotic (plant or animal) organisms, may be used to practice this invention, provided that it is compatible with the host cell into which it is inserted. Also included herein are synthetic introns. Optionally, more than one intron may be used in the vector.

The expression and cloning vectors of the present invention will typically contain a promoter that is recognized by the host organism and operably linked to the molecule encoding the FGF-23 polypeptide. Promoters are untranscribed sequences located upstream (*i.e.*, 5') to the start codon of a structural gene (generally within about 100 to 1000 bp) that control the transcription of the structural gene. Promoters are conventionally grouped into one of two classes: inducible promoters and constitutive promoters. Inducible promoters initiate increased levels of transcription from DNA under their control in response to some change in culture conditions, such as the presence or absence of a nutrient or a change in temperature. Constitutive promoters, on the other hand, initiate continual gene product production; that is, there is little or no control over gene expression. A large number of promoters, recognized by a variety of potential host cells, are well known. A suitable promoter is operably linked to the DNA encoding FGF-23 polypeptide by removing the promoter from the source DNA by restriction enzyme digestion and inserting the desired promoter sequence into the vector. The native FGF-23 promoter sequence may be used to direct amplification and/or expression of an FGF-23 nucleic acid molecule. A heterologous promoter is preferred, however, if it permits greater transcription and higher yields of the expressed protein as compared to the native promoter, and if it is compatible with the host cell system that has been selected for use.

Promoters suitable for use with prokaryotic hosts include the beta-lactamase and lactose promoter systems; alkaline phosphatase; a tryptophan (trp) promoter system; and hybrid promoters such as the tac promoter. Other known bacterial promoters are also suitable. Their sequences have been published,

thereby enabling one skilled in the art to ligate them to the desired DNA sequence, using linkers or adapters as needed to supply any useful restriction sites.

Suitable promoters for use with yeast hosts are also well known in the art. Yeast enhancers are advantageously used with yeast promoters. Suitable promoters for use with mammalian host cells are well known and include, but are not limited to, those obtained from the genomes of viruses such as polyoma virus, fowlpox virus, adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, retroviruses, hepatitis-B virus and most preferably Simian Virus 40 (SV40). Other suitable mammalian promoters include heterologous mammalian promoters, for example, heat-shock promoters and the actin promoter.

Additional promoters which may be of interest in controlling FGF-23 gene expression include, but are not limited to: the SV40 early promoter region (Bernoist and Chambon, 1981, *Nature* 290:304-10); the CMV promoter; the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, *et al.*, 1980, *Cell* 22:787-97); the herpes thymidine kinase promoter (Wagner *et al.*, 1981, *Proc. Natl. Acad. Sci. U.S.A.* 78:1444-45); the regulatory sequences of the metallothionein gene (Brinster *et al.*, 1982, *Nature* 296:39-42); prokaryotic expression vectors such as the beta-lactamase promoter (Villa-Kamaroff *et al.*, 1978, *Proc. Natl. Acad. Sci. U.S.A.*, 75:3727-31); or the tac promoter (DeBoer *et al.*, 1983, *Proc. Natl. Acad. Sci. U.S.A.*, 80:21-25). Also of interest are the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: the elastase I gene control region which is active in pancreatic acinar cells (Swift *et al.*, 1984, *Cell* 38:639-46; Ornitz *et al.*, 1986, *Cold Spring Harbor Symp. Quant. Biol.* 50:399-409 (1986); MacDonald, 1987, *Hepatology* 7:425-515); the insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, *Nature* 315:115-22); the immunoglobulin gene control region which is active in lymphoid cells (Grosschedl *et al.*, 1984, *Cell* 38:647-58; Adames *et al.*, 1985, *Nature* 318:533-38; Alexander *et al.*, 1987, *Mol. Cell. Biol.*, 7:1436-44); the mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells

(Leder *et al.*, 1986, *Cell* 45:485-95); the albumin gene control region which is active in liver (Pinkert *et al.*, 1987, *Genes and Devel.* 1:268-76); the alpha-feto-protein gene control region which is active in liver (Krumlauf *et al.*, 1985, *Mol. Cell. Biol.*, 5:1639-48; Hammer *et al.*, 1987, *Science* 235:53-58); the alpha 1-
5 antitrypsin gene control region which is active in the liver (Kelsey *et al.*, 1987, *Genes and Devel.* 1:161-71); the beta-globin gene control region which is active in myeloid cells (Mogram *et al.*, 1985, *Nature* 315:338-40; Kollias *et al.*, 1986, *Cell* 46:89-94); the myelin basic protein gene control region which is active in oligodendrocyte cells in the brain (Readhead *et al.*, 1987, *Cell* 48:703-12); the
10 myosin light chain-2 gene control region which is active in skeletal muscle (Sani, 1985, *Nature* 314:283-86); and the gonadotropic releasing hormone gene control region which is active in the hypothalamus (Mason *et al.*, 1986, *Science* 234:1372-78).

An enhancer sequence may be inserted into the vector to increase the
15 transcription of a DNA encoding an FGF-23 polypeptide of the present invention by higher eukaryotes. Enhancers are cis-acting elements of DNA, usually about 10-300 bp in length, that act on the promoter to increase transcription. Enhancers are relatively orientation and position independent. They have been found 5' and 3' to the transcription unit. Several enhancer sequences available from
20 mammalian genes are known (e.g., globin, elastase, albumin, alpha-feto-protein and insulin). Typically, however, an enhancer from a virus will be used. The SV40 enhancer, the cytomegalovirus early promoter enhancer, the polyoma enhancer, and adenovirus enhancers are exemplary enhancing elements for the activation of eukaryotic promoters. While an enhancer may be spliced into the
25 vector at a position 5' or 3' to an FGF-23 nucleic acid molecule, it is typically located at a site 5' from the promoter.

Expression vectors of the invention may be constructed from a starting vector such as a commercially available vector. Such vectors may or may not contain all of the desired flanking sequences. Where one or more of the flanking
30 sequences described herein are not already present in the vector, they may be

individually obtained and ligated into the vector. Methods used for obtaining each of the flanking sequences are well known to one skilled in the art.

Preferred vectors for practicing this invention are those which are compatible with bacterial, insect, and mammalian host cells. Such vectors include, *inter alia*, pCRII, pCR3, and pcDNA3.1 (Invitrogen, San Diego, CA), pBSII (Stratagene, La Jolla, CA), pET15 (Novagen, Madison, WI), pGEX (Pharmacia Biotech, Piscataway, NJ), pEGFP-N2 (Clontech, Palo Alto, CA), pETL (BlueBacII, Invitrogen), pDSR-alpha (PCT Pub. No. WO 90/14363) and pFastBacDual (Gibco-BRL, Grand Island, NY).

Additional suitable vectors include, but are not limited to, cosmids, plasmids, or modified viruses, but it will be appreciated that the vector system must be compatible with the selected host cell. Such vectors include, but are not limited to plasmids such as Bluescript[®] plasmid derivatives (a high copy number ColE1-based phagemid, Stratagene Cloning Systems, La Jolla CA), PCR cloning plasmids designed for cloning Taq-amplified PCR products (*e.g.*, TOPO[™] TA Cloning[®] Kit, PCR2.1[®] plasmid derivatives, Invitrogen, Carlsbad, CA), and mammalian, yeast or virus vectors such as a baculovirus expression system (pBacPAK plasmid derivatives, Clontech, Palo Alto, CA).

After the vector has been constructed and a nucleic acid molecule encoding an FGF-23 polypeptide has been inserted into the proper site of the vector, the completed vector may be inserted into a suitable host cell for amplification and/or polypeptide expression. The transformation of an expression vector for an FGF-23 polypeptide into a selected host cell may be accomplished by well known methods including methods such as transfection, infection, calcium chloride, electroporation, microinjection, lipofection, DEAE-dextran method, or other known techniques. The method selected will in part be a function of the type of host cell to be used. These methods and other suitable methods are well known to the skilled artisan, and are set forth, for example, in Sambrook *et al.*, *supra*.

Host cells may be prokaryotic host cells (such as *E. coli*) or eukaryotic host cells (such as a yeast, insect, or vertebrate cell). The host cell, when cultured

under appropriate conditions, synthesizes an FGF-23 polypeptide which can subsequently be collected from the culture medium (if the host cell secretes it into the medium) or directly from the host cell producing it (if it is not secreted). The selection of an appropriate host cell will depend upon various factors, such as
5 desired expression levels, polypeptide modifications that are desirable or necessary for activity (such as glycosylation or phosphorylation) and ease of folding into a biologically active molecule.

A number of suitable host cells are known in the art and many are available from the American Type Culture Collection (ATCC), Manassas, VA.
10 Examples include, but are not limited to, mammalian cells, such as Chinese hamster ovary cells (CHO), CHO DHFR(-) cells (Urlaub *et al.*, 1980, *Proc. Natl. Acad. Sci. U.S.A.* 97:4216-20), human embryonic kidney (HEK) 293 or 293T cells, or 3T3 cells. The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production, and
15 purification are known in the art. Other suitable mammalian cell lines, are the monkey COS-1 and COS-7 cell lines, and the CV-1 cell line. Further exemplary mammalian host cells include primate cell lines and rodent cell lines, including transformed cell lines. Normal diploid cells, cell strains derived from *in vitro* culture of primary tissue, as well as primary explants, are also suitable. Candidate
20 cells may be genotypically deficient in the selection gene, or may contain a dominantly acting selection gene. Other suitable mammalian cell lines include but are not limited to, mouse neuroblastoma N2A cells, HeLa, mouse L-929 cells, 3T3 lines derived from Swiss, Balb-c or NIH mice, BHK or HaK hamster cell lines. Each of these cell lines is known by and available to those skilled in the art
25 of protein expression.

Similarly useful as host cells suitable for the present invention are bacterial cells. For example, the various strains of *E. coli* (*e.g.*, HB101, DH5 α , DH10, and MC1061) are well-known as host cells in the field of biotechnology. Various strains of *B. subtilis*, *Pseudomonas spp.*, other *Bacillus spp.*,
30 *Streptomyces spp.*, and the like may also be employed in this method.

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Many strains of yeast cells known to those skilled in the art are also available as host cells for the expression of the polypeptides of the present invention. Preferred yeast cells include, for example, *Saccharomyces cerevisiae* and *Pichia pastoris*.

5 Additionally, where desired, insect cell systems may be utilized in the methods of the present invention. Such systems are described, for example, in Kitts *et al.*, 1993, *Biotechniques*, 14:810-17; Lucklow, 1993, *Curr. Opin. Biotechnol.* 4:564-72; and Lucklow *et al.*, 1993, *J. Virol.*, 67:4566-79. Preferred insect cells are Sf-9 and Hi5 (Invitrogen).

10 One may also use transgenic animals to express glycosylated FGF-23 polypeptides. For example, one may use a transgenic milk-producing animal (a cow or goat, for example) and obtain the present glycosylated polypeptide in the animal milk. One may also use plants to produce FGF-23 polypeptides, however, in general, the glycosylation occurring in plants is different from that produced in
15 mammalian cells, and may result in a glycosylated product which is not suitable for human therapeutic use.

Polypeptide Production

Host cells comprising an FGF-23 polypeptide expression vector may be
20 cultured using standard media well known to the skilled artisan. The media will usually contain all nutrients necessary for the growth and survival of the cells. Suitable media for culturing *E. coli* cells include, for example, Luria Broth (LB) and/or Terrific Broth (TB). Suitable media for culturing eukaryotic cells include Roswell Park Memorial Institute medium 1640 (RPMI 1640), Minimal Essential
25 Medium (MEM) and/or Dulbecco's Modified Eagle Medium (DMEM), all of which may be supplemented with serum and/or growth factors as necessary for the particular cell line being cultured. A suitable medium for insect cultures is Grace's medium supplemented with yeastolate, lactalbumin hydrolysate, and/or fetal calf serum as necessary.

30 Typically, an antibiotic or other compound useful for selective growth of transfected or transformed cells is added as a supplement to the media. The

compound to be used will be dictated by the selectable marker element present on the plasmid with which the host cell was transformed. For example, where the selectable marker element is kanamycin resistance, the compound added to the culture medium will be kanamycin. Other compounds for selective growth
5 include ampicillin, tetracycline, and neomycin.

The amount of an FGF-23 polypeptide produced by a host cell can be evaluated using standard methods known in the art. Such methods include, without limitation, Western blot analysis, SDS-polyacrylamide gel electrophoresis, non-denaturing gel electrophoresis, High Performance Liquid
10 Chromatography (HPLC) separation, immunoprecipitation, and/or activity assays such as DNA binding gel shift assays.

If an FGF-23 polypeptide has been designed to be secreted from the host cells, the majority of polypeptide may be found in the cell culture medium. If however, the FGF-23 polypeptide is not secreted from the host cells, it will be
15 present in the cytoplasm and/or the nucleus (for eukaryotic host cells) or in the cytosol (for gram-negative bacteria host cells).

For an FGF-23 polypeptide situated in the host cell cytoplasm and/or nucleus (for eukaryotic host cells) or in the cytosol (for bacterial host cells), the intracellular material (including inclusion bodies for gram-negative bacteria) can
20 be extracted from the host cell using any standard technique known to the skilled artisan. For example, the host cells can be lysed to release the contents of the periplasm/cytoplasm by French press, homogenization, and/or sonication followed by centrifugation.

If an FGF-23 polypeptide has formed inclusion bodies in the cytosol, the
25 inclusion bodies can often bind to the inner and/or outer cellular membranes and thus will be found primarily in the pellet material after centrifugation. The pellet material can then be treated at pH extremes or with a chaotropic agent such as a detergent, guanidine, guanidine derivatives, urea, or urea derivatives in the presence of a reducing agent such as dithiothreitol at alkaline pH or tris
30 carboxyethyl phosphine at acid pH to release, break apart, and solubilize the inclusion bodies. The solubilized FGF-23 polypeptide can then be analyzed using

gel electrophoresis, immunoprecipitation, or the like. If it is desired to isolate the FGF-23 polypeptide, isolation may be accomplished using standard methods such as those described herein and in Marston *et al.*, 1990, *Meth. Enz.*, 182:264-75.

In some cases, an FGF-23 polypeptide may not be biologically active upon
5 isolation. Various methods for "refolding" or converting the polypeptide to its tertiary structure and generating disulfide linkages can be used to restore biological activity. Such methods include exposing the solubilized polypeptide to a pH usually above 7 and in the presence of a particular concentration of a chaotrope. The selection of chaotrope is very similar to the choices used for
10 inclusion body solubilization, but usually the chaotrope is used at a lower concentration and is not necessarily the same as chaotropes used for the solubilization. In most cases the refolding/oxidation solution will also contain a reducing agent or the reducing agent plus its oxidized form in a specific ratio to generate a particular redox potential allowing for disulfide shuffling to occur in
15 the formation of the protein's cysteine bridges. Some of the commonly used redox couples include cysteine/cystamine, glutathione (GSH)/dithiobis GSH, cupric chloride, dithiothreitol(DTT)/dithiane DTT, and 2-2-mercaptoethanol(bME)/dithio-b(ME). In many instances, a cosolvent may be used or may be needed to increase the efficiency of the refolding, and the more
20 common reagents used for this purpose include glycerol, polyethylene glycol of various molecular weights, arginine and the like.

If inclusion bodies are not formed to a significant degree upon expression of an FGF-23 polypeptide, then the polypeptide will be found primarily in the supernatant after centrifugation of the cell homogenate. The polypeptide may be
25 further isolated from the supernatant using methods such as those described herein.

The purification of an FGF-23 polypeptide from solution can be accomplished using a variety of techniques. If the polypeptide has been synthesized such that it contains a tag such as Hexahistidine (FGF-23
30 polypeptide/hexaHis) or other small peptide such as FLAG (Eastman Kodak Co., New Haven, CT) or *myc* (Invitrogen, Carlsbad, CA) at either its carboxyl- or

amino-terminus, it may be purified in a one-step process by passing the solution through an affinity column where the column matrix has a high affinity for the tag.

For example, polyhistidine binds with great affinity and specificity to nickel. Thus, an affinity column of nickel (such as the Qiagen[®] nickel columns) can be used for purification of FGF-23 polypeptide/polyHis. See, e.g., *Current Protocols in Molecular Biology* § 10.11.8 (Ausubel *et al.*, eds., Green Publishers Inc. and Wiley and Sons 1993).

Additionally, FGF-23 polypeptides may be purified through the use of a monoclonal antibody that is capable of specifically recognizing and binding to an FGF-23 polypeptide.

Other suitable procedures for purification include, without limitation, affinity chromatography, immunoaffinity chromatography, ion exchange chromatography, molecular sieve chromatography, HPLC, electrophoresis (including native gel electrophoresis) followed by gel elution, and preparative isoelectric focusing ("Isoprime" machine/technique, Hoefer Scientific, San Francisco, CA). In some cases, two or more purification techniques may be combined to achieve increased purity.

FGF-23 polypeptides may also be prepared by chemical synthesis methods (such as solid phase peptide synthesis) using techniques known in the art such as those set forth by Merrifield *et al.*, 1963, *J. Am. Chem. Soc.* 85:2149; Houghten *et al.*, 1985, *Proc Natl Acad. Sci. USA* 82:5132; and Stewart and Young, *Solid Phase Peptide Synthesis* (Pierce Chemical Co. 1984). Such polypeptides may be synthesized with or without a methionine on the amino-terminus. Chemically synthesized FGF-23 polypeptides may be oxidized using methods set forth in these references to form disulfide bridges. Chemically synthesized FGF-23 polypeptides are expected to have comparable biological activity to the corresponding FGF-23 polypeptides produced recombinantly or purified from natural sources, and thus may be used interchangeably with a recombinant or natural FGF-23 polypeptide.

Another means of obtaining FGF-23 polypeptide is via purification from biological samples such as source tissues and/or fluids in which the FGF-23 polypeptide is naturally found. Such purification can be conducted using methods for protein purification as described herein. The presence of the FGF-23 polypeptide during purification may be monitored, for example, using an antibody prepared against recombinantly produced FGF-23 polypeptide or peptide fragments thereof.

A number of additional methods for producing nucleic acids and polypeptides are known in the art, and the methods can be used to produce polypeptides having specificity for FGF-23 polypeptide. *See, e.g., Roberts et al., 1997, Proc. Natl. Acad. Sci. U.S.A. 94:12297-303*, which describes the production of fusion proteins between an mRNA and its encoded peptide. *See also, Roberts, 1999, Curr. Opin. Chem. Biol. 3:268-73*. Additionally, U.S. Patent No. 5,824,469 describes methods for obtaining oligonucleotides capable of carrying out a specific biological function. The procedure involves generating a heterogeneous pool of oligonucleotides, each having a 5' randomized sequence, a central preselected sequence, and a 3' randomized sequence. The resulting heterogeneous pool is introduced into a population of cells that do not exhibit the desired biological function. Subpopulations of the cells are then screened for those that exhibit a predetermined biological function. From that subpopulation, oligonucleotides capable of carrying out the desired biological function are isolated.

U.S. Patent Nos. 5,763,192; 5,814,476; 5,723,323; and 5,817,483 describe processes for producing peptides or polypeptides. This is done by producing stochastic genes or fragments thereof, and then introducing these genes into host cells which produce one or more proteins encoded by the stochastic genes. The host cells are then screened to identify those clones producing peptides or polypeptides having the desired activity.

Another method for producing peptides or polypeptides is described in PCT/US98/20094 (WO99/15650) filed by Athersys, Inc. Known as "Random Activation of Gene Expression for Gene Discovery" (RAGE-GD), the process

involves the activation of endogenous gene expression or over-expression of a gene by *in situ* recombination methods. For example, expression of an endogenous gene is activated or increased by integrating a regulatory sequence into the target cell which is capable of activating expression of the gene by non-
5 homologous or illegitimate recombination. The target DNA is first subjected to radiation, and a genetic promoter inserted. The promoter eventually locates a break at the front of a gene, initiating transcription of the gene. This results in expression of the desired peptide or polypeptide.

It will be appreciated that these methods can also be used to create
10 comprehensive FGF-23 polypeptide expression libraries, which can subsequently be used for high throughput phenotypic screening in a variety of assays, such as biochemical assays, cellular assays, and whole organism assays (e.g., plant, mouse, etc.).

15 Synthesis

It will be appreciated by those skilled in the art that the nucleic acid and polypeptide molecules described herein may be produced by recombinant and other means.

20 Selective Binding Agents

The term "selective binding agent" refers to a molecule that has specificity for one or more FGF-23 polypeptides. Suitable selective binding agents include, but are not limited to, antibodies and derivatives thereof, polypeptides, and small molecules. Suitable selective binding agents may be prepared using methods
25 known in the art. An exemplary FGF-23 polypeptide selective binding agent of the present invention is capable of binding a certain portion of the FGF-23 polypeptide thereby inhibiting the binding of the polypeptide to an FGF-23 polypeptide receptor.

Selective binding agents such as antibodies and antibody fragments that
30 bind FGF-23 polypeptides are within the scope of the present invention. The antibodies may be polyclonal including monospecific polyclonal; monoclonal

(MAbs); recombinant; chimeric; humanized, such as CDR-grafted; human; single chain; and/or bispecific; as well as fragments; variants; or derivatives thereof. Antibody fragments include those portions of the antibody that bind to an epitope on the FGF-23 polypeptide. Examples of such fragments include Fab and F(ab')
5 fragments generated by enzymatic cleavage of full-length antibodies. Other binding fragments include those generated by recombinant DNA techniques, such as the expression of recombinant plasmids containing nucleic acid sequences encoding antibody variable regions.

Polyclonal antibodies directed toward an FGF-23 polypeptide generally
10 are produced in animals (e.g., rabbits or mice) by means of multiple subcutaneous or intraperitoneal injections of FGF-23 polypeptide and an adjuvant. It may be useful to conjugate an FGF-23 polypeptide to a carrier protein that is immunogenic in the species to be immunized, such as keyhole limpet hemocyanin, serum, albumin, bovine thyroglobulin, or soybean trypsin inhibitor.
15 Also, aggregating agents such as alum are used to enhance the immune response. After immunization, the animals are bled and the serum is assayed for anti-FGF-23 antibody titer.

Monoclonal antibodies directed toward FGF-23 polypeptides are produced using any method that provides for the production of antibody molecules by
20 continuous cell lines in culture. Examples of suitable methods for preparing monoclonal antibodies include the hybridoma methods of Kohler *et al.*, 1975, *Nature* 256:495-97 and the human B-cell hybridoma method (Kozbor, 1984, *J. Immunol.* 133:3001; Brodeur *et al.*, *Monoclonal Antibody Production Techniques and Applications* 51-63 (Marcel Dekker, Inc., 1987). Also provided by the
25 invention are hybridoma cell lines that produce monoclonal antibodies reactive with FGF-23 polypeptides.

Monoclonal antibodies of the invention may be modified for use as therapeutics. One embodiment is a "chimeric" antibody in which a portion of the heavy (H) and/or light (L) chain is identical with or homologous to a
30 corresponding sequence in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the

chain(s) is/are identical with or homologous to a corresponding sequence in antibodies derived from another species or belonging to another antibody class or subclass. Also included are fragments of such antibodies, so long as they exhibit the desired biological activity. See U.S. Patent No. 4,816,567; Morrison *et al.*,
5 1985, *Proc. Natl. Acad. Sci.* 81:6851-55.

In another embodiment, a monoclonal antibody of the invention is a "humanized" antibody. Methods for humanizing non-human antibodies are well known in the art. See U.S. Patent Nos. 5,585,089 and 5,693,762. Generally, a humanized antibody has one or more amino acid residues introduced into it from
10 a source that is non-human. Humanization can be performed, for example, using methods described in the art (Jones *et al.*, 1986, *Nature* 321:522-25; Riechmann *et al.*, 1998, *Nature* 332:323-27; Verhoeven *et al.*, 1988, *Science* 239:1534-36), by substituting at least a portion of a rodent complementarity-determining region (CDR) for the corresponding regions of a human antibody.

Also encompassed by the invention are human antibodies that bind FGF-23 polypeptides. Using transgenic animals (*e.g.*, mice) that are capable of producing a repertoire of human antibodies in the absence of endogenous immunoglobulin production such antibodies are produced by immunization with an FGF-23 polypeptide antigen (*i.e.*, having at least 6 contiguous amino acids),
20 optionally conjugated to a carrier. See, *e.g.*, Jakobovits *et al.*, 1993, *Proc. Natl. Acad. Sci.* 90:2551-55; Jakobovits *et al.*, 1993, *Nature* 362:255-58; Bruggermann *et al.*, 1993, *Year in Immuno.* 7:33. In one method, such transgenic animals are produced by incapacitating the endogenous loci encoding the heavy and light immunoglobulin chains therein, and inserting loci encoding human heavy and
25 light chain proteins into the genome thereof. Partially modified animals, that is those having less than the full complement of modifications, are then cross-bred to obtain an animal having all of the desired immune system modifications. When administered an immunogen, these transgenic animals produce antibodies with human (rather than, *e.g.*, murine) amino acid sequences, including variable
30 regions which are immunospecific for these antigens. See PCT App. Nos. PCT/US96/05928 and PCT/US93/06926. Additional methods are described in

U.S. Patent No. 5,545,807, PCT App. Nos. PCT/US91/245 and PCT/GB89/01207, and in European Patent Nos. 546073B1 and 546073A1. Human antibodies can also be produced by the expression of recombinant DNA in host cells or by expression in hybridoma cells as described herein.

5 In an alternative embodiment, human antibodies can also be produced from phage-display libraries (Hoogenboom *et al.*, 1991, *J. Mol. Biol.* 227:381; Marks *et al.*, 1991, *J. Mol. Biol.* 222:581). These processes mimic immune selection through the display of antibody repertoires on the surface of filamentous bacteriophage, and subsequent selection of phage by their binding to an antigen of
10 choice. One such technique is described in PCT App. No. PCT/US98/17364, which describes the isolation of high affinity and functional agonistic antibodies for MPL- and msk- receptors using such an approach.

Chimeric, CDR grafted, and humanized antibodies are typically produced by recombinant methods. Nucleic acids encoding the antibodies are introduced
15 into host cells and expressed using materials and procedures described herein. In a preferred embodiment, the antibodies are produced in mammalian host cells, such as CHO cells. Monoclonal (*e.g.*, human) antibodies may be produced by the expression of recombinant DNA in host cells or by expression in hybridoma cells as described herein.

20 The anti-FGF-23 antibodies of the invention may be employed in any known assay method, such as competitive binding assays, direct and indirect sandwich assays, and immunoprecipitation assays (Sola, *Monoclonal Antibodies: A Manual of Techniques* 147-158 (CRC Press, Inc., 1987)) for the detection and quantitation of FGF-23 polypeptides. The antibodies will bind FGF-23
25 polypeptides with an affinity that is appropriate for the assay method being employed.

For diagnostic applications, in certain embodiments, anti-FGF-23 antibodies may be labeled with a detectable moiety. The detectable moiety can be any one that is capable of producing, either directly or indirectly, a detectable
30 signal. For example, the detectable moiety may be a radioisotope, such as ³H, ¹⁴C, ³²P, ³⁵S, ¹²⁵I, ⁹⁹Tc, ¹¹¹In, or ⁶⁷Ga; a fluorescent or chemiluminescent

compound, such as fluorescein isothiocyanate, rhodamine, or luciferin; or an enzyme, such as alkaline phosphatase, β -galactosidase, or horseradish peroxidase (Bayer, *et al.*, 1990, *Meth. Enz.* 184:138-63).

Competitive binding assays rely on the ability of a labeled standard (*e.g.*,
5 an FGF-23 polypeptide, or an immunologically reactive portion thereof) to compete with the test sample analyte (an FGF-23 polypeptide) for binding with a limited amount of anti-FGF-23 antibody. The amount of an FGF-23 polypeptide in the test sample is inversely proportional to the amount of standard that becomes bound to the antibodies. To facilitate determining the amount of standard that
10 becomes bound, the antibodies typically are insolubilized before or after the competition, so that the standard and analyte that are bound to the antibodies may conveniently be separated from the standard and analyte which remain unbound.

Sandwich assays typically involve the use of two antibodies, each capable of binding to a different immunogenic portion, or epitope, of the protein to be
15 detected and/or quantitated. In a sandwich assay, the test sample analyte is typically bound by a first antibody which is immobilized on a solid support, and thereafter a second antibody binds to the analyte, thus forming an insoluble three-part complex. *See, e.g.*, U.S. Patent No. 4,376,110. The second antibody may itself be labeled with a detectable moiety (direct sandwich assays) or may be
20 measured using an anti-immunoglobulin antibody that is labeled with a detectable moiety (indirect sandwich assays). For example, one type of sandwich assay is an enzyme-linked immunosorbent assay (ELISA), in which case the detectable moiety is an enzyme.

The selective binding agents, including anti-FGF-23 antibodies, are also
25 useful for *in vivo* imaging. An antibody labeled with a detectable moiety may be administered to an animal, preferably into the bloodstream, and the presence and location of the labeled antibody in the host assayed. The antibody may be labeled with any moiety that is detectable in an animal, whether by nuclear magnetic resonance, radiology, or other detection means known in the art.

30 Selective binding agents of the invention, including antibodies, may be used as therapeutics. These therapeutic agents are generally agonists or

antagonists, in that they either enhance or reduce, respectively, at least one of the biological activities of an FGF-23 polypeptide. In one embodiment, antagonist antibodies of the invention are antibodies or binding fragments thereof which are capable of specifically binding to an FGF-23 polypeptide and which are capable of inhibiting or eliminating the functional activity of an FGF-23 polypeptide *in vivo* or *in vitro*. In preferred embodiments, the selective binding agent, *e.g.*, an antagonist antibody, will inhibit the functional activity of an FGF-23 polypeptide by at least about 50%, and preferably by at least about 80%. In another embodiment, the selective binding agent may be an anti-FGF-23 polypeptide antibody that is capable of interacting with an FGF-23 polypeptide binding partner (a ligand or receptor) thereby inhibiting or eliminating FGF-23 polypeptide activity *in vitro* or *in vivo*. Selective binding agents, including agonist and antagonist anti-FGF-23 polypeptide antibodies, are identified by screening assays that are well known in the art.

The invention also relates to a kit comprising FGF-23 selective binding agents (such as antibodies) and other reagents useful for detecting FGF-23 polypeptide levels in biological samples. Such reagents may include a detectable label, blocking serum, positive and negative control samples, and detection reagents.

20

Microarrays

It will be appreciated that DNA microarray technology can be utilized in accordance with the present invention. DNA microarrays are miniature, high-density arrays of nucleic acids positioned on a solid support, such as glass. Each cell or element within the array contains numerous copies of a single nucleic acid species that acts as a target for hybridization with a complementary nucleic acid sequence (*e.g.*, mRNA). In expression profiling using DNA microarray technology, mRNA is first extracted from a cell or tissue sample and then converted enzymatically to fluorescently labeled cDNA. This material is hybridized to the microarray and unbound cDNA is removed by washing. The expression of discrete genes represented on the array is then visualized by

30

quantitating the amount of labeled cDNA that is specifically bound to each target nucleic acid molecule. In this way, the expression of thousands of genes can be quantitated in a high throughput, parallel manner from a single sample of biological material.

5 This high throughput expression profiling has a broad range of applications with respect to the FGF-23 molecules of the invention, including, but not limited to: the identification and validation of FGF-23 disease-related genes as targets for therapeutics; molecular toxicology of related FGF-23 molecules and inhibitors thereof; stratification of populations and generation of surrogate
10 markers for clinical trials; and enhancing related FGF-23 polypeptide small molecule drug discovery by aiding in the identification of selective compounds in high throughput screens.

Chemical Derivatives

15 Chemically modified derivatives of FGF-23 polypeptides may be prepared by one skilled in the art, given the disclosures described herein. FGF-23 polypeptide derivatives are modified in a manner that is different – either in the type or location of the molecules naturally attached to the polypeptide. Derivatives may include molecules formed by the deletion of one or more
20 naturally-attached chemical groups. The polypeptide comprising the amino acid sequence of SEQ ID NO: 2, or other FGF-23 polypeptide, may be modified by the covalent attachment of one or more polymers. For example, the polymer selected is typically water-soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a physiological environment.
25 Included within the scope of suitable polymers is a mixture of polymers. Preferably, for therapeutic use of the end-product preparation, the polymer will be pharmaceutically acceptable.

 The polymers each may be of any molecular weight and may be branched or unbranched. The polymers each typically have an average molecular weight of
30 between about 2 kDa to about 100 kDa (the term “about” indicating that in preparations of a water-soluble polymer, some molecules will weigh more, some

less, than the stated molecular weight). The average molecular weight of each polymer is preferably between about 5 kDa and about 50 kDa, more preferably between about 12 kDa and about 40 kDa and most preferably between about 20 kDa and about 35 kDa.

5 Suitable water-soluble polymers or mixtures thereof include, but are not limited to, N-linked or O-linked carbohydrates, sugars, phosphates, polyethylene glycol (PEG) (including the forms of PEG that have been used to derivatize proteins, including mono-(C₁-C₁₀), alkoxy-, or aryloxy-polyethylene glycol), monomethoxy-polyethylene glycol, dextran (such as low molecular weight
10 dextran of, for example, about 6 kD), cellulose, or other carbohydrate based polymers, poly-(N-vinyl pyrrolidone) polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide co-polymers, polyoxyethylated polyols (*e.g.*, glycerol), and polyvinyl alcohol. Also encompassed by the present invention are bifunctional crosslinking molecules
15 which may be used to prepare covalently attached FGF-23 polypeptide multimers.

 In general, chemical derivatization may be performed under any suitable condition used to react a protein with an activated polymer molecule. Methods for preparing chemical derivatives of polypeptides will generally comprise the steps of: (a) reacting the polypeptide with the activated polymer molecule (such as
20 a reactive ester or aldehyde derivative of the polymer molecule) under conditions whereby the polypeptide comprising the amino acid sequence of SEQ ID NO: 2, or other FGF-23 polypeptide, becomes attached to one or more polymer molecules, and (b) obtaining the reaction products. The optimal reaction conditions will be determined based on known parameters and the desired result.
25 For example, the larger the ratio of polymer molecules to protein, the greater the percentage of attached polymer molecule. In one embodiment, the FGF-23 polypeptide derivative may have a single polymer molecule moiety at the amino-terminus. *See, e.g.*, U.S. Patent No. 5,234,784.

 The pegylation of a polypeptide may be specifically carried out using any
30 of the pegylation reactions known in the art. Such reactions are described, for example, in the following references: Francis *et al.*, 1992, *Focus on Growth*

Factors 3:4-10; European Patent Nos. 0154316 and 0401384; and U.S. Patent No. 4,179,337. For example, pegylation may be carried out via an acylation reaction or an alkylation reaction with a reactive polyethylene glycol molecule (or an analogous reactive water-soluble polymer) as described herein. For the acylation reactions, a selected polymer should have a single reactive ester group. For reductive alkylation, a selected polymer should have a single reactive aldehyde group. A reactive aldehyde is, for example, polyethylene glycol propionaldehyde, which is water stable, or mono C₁-C₁₀ alkoxy or aryloxy derivatives thereof (*see* U.S. Patent No. 5,252,714).

10 In another embodiment, FGF-23 polypeptides may be chemically coupled to biotin. The biotin/FGF-23 polypeptide molecules are then allowed to bind to avidin, resulting in tetravalent avidin/biotin/FGF-23 polypeptide molecules. FGF-23 polypeptides may also be covalently coupled to dinitrophenol (DNP) or trinitrophenol (TNP) and the resulting conjugates precipitated with anti-DNP or
15 anti-TNP-IgM to form decameric conjugates with a valency of 10.

Generally, conditions that may be alleviated or modulated by the administration of the present FGF-23 polypeptide derivatives include those described herein for FGF-23 polypeptides. However, the FGF-23 polypeptide derivatives disclosed herein may have additional activities, enhanced or reduced
20 biological activity, or other characteristics, such as increased or decreased half-life, as compared to the non-derivatized molecules.

Genetically Engineered Non-Human Animals

Additionally included within the scope of the present invention are non-
25 human animals such as mice, rats, or other rodents; rabbits, goats, sheep, or other farm animals, in which the genes encoding native FGF-23 polypeptide have been disrupted (*i.e.*, "knocked out") such that the level of expression of FGF-23 polypeptide is significantly decreased or completely abolished. Such animals may be prepared using techniques and methods such as those described in U.S.
30 Patent No. 5,557,032.

The present invention further includes non-human animals such as mice, rats, or other rodents; rabbits, goats, sheep, or other farm animals, in which either the native form of an FGF-23 gene for that animal or a heterologous FGF-23 gene is over-expressed by the animal, thereby creating a "transgenic" animal. Such transgenic animals may be prepared using well known methods such as those described in U.S. Patent No 5,489,743 and PCT Pub. No. WO 94/28122.

The present invention further includes non-human animals in which the promoter for one or more of the FGF-23 polypeptides of the present invention is either activated or inactivated (*e.g.*, by using homologous recombination methods) to alter the level of expression of one or more of the native FGF-23 polypeptides.

These non-human animals may be used for drug candidate screening. In such screening, the impact of a drug candidate on the animal may be measured. For example, drug candidates may decrease or increase the expression of the FGF-23 gene. In certain embodiments, the amount of FGF-23 polypeptide that is produced may be measured after the exposure of the animal to the drug candidate. Additionally, in certain embodiments, one may detect the actual impact of the drug candidate on the animal. For example, over-expression of a particular gene may result in, or be associated with, a disease or pathological condition. In such cases, one may test a drug candidate's ability to decrease expression of the gene or its ability to prevent or inhibit a pathological condition. In other examples, the production of a particular metabolic product such as a fragment of a polypeptide, may result in, or be associated with, a disease or pathological condition. In such cases, one may test a drug candidate's ability to decrease the production of such a metabolic product or its ability to prevent or inhibit a pathological condition.

25

Assaying for Other Modulators of FGF-23 Polypeptide Activity

In some situations, it may be desirable to identify molecules that are modulators, *i.e.*, agonists or antagonists, of the activity of FGF-23 polypeptide. Natural or synthetic molecules that modulate FGF-23 polypeptide may be identified using one or more screening assays, such as those described herein.

30

Such molecules may be administered either in an *ex vivo* manner or in an *in vivo* manner by injection, or by oral delivery, implantation device, or the like.

“Test molecule” refers to a molecule that is under evaluation for the ability to modulate (*i.e.*, increase or decrease) the activity of an FGF-23 polypeptide.

5 Most commonly, a test molecule will interact directly with an FGF-23 polypeptide. However, it is also contemplated that a test molecule may also modulate FGF-23 polypeptide activity indirectly, such as by affecting FGF-23 gene expression, or by binding to an FGF-23 polypeptide binding partner (*e.g.*, receptor or ligand). In one embodiment, a test molecule will bind to an FGF-23
10 polypeptide with an affinity constant of at least about 10^{-6} M, preferably about 10^{-8} M, more preferably about 10^{-9} M, and even more preferably about 10^{-10} M.

Methods for identifying compounds that interact with FGF-23 polypeptides are encompassed by the present invention. In certain embodiments, an FGF-23 polypeptide is incubated with a test molecule under conditions that
15 permit the interaction of the test molecule with an FGF-23 polypeptide, and the extent of the interaction is measured. The test molecule can be screened in a substantially purified form or in a crude mixture.

In certain embodiments, an FGF-23 polypeptide agonist or antagonist may be a protein, peptide, carbohydrate, lipid, or small molecular weight molecule that
20 interacts with FGF-23 polypeptide to regulate its activity. Molecules which regulate FGF-23 polypeptide expression include nucleic acids which are complementary to nucleic acids encoding an FGF-23 polypeptide, or are complementary to nucleic acids sequences which direct or control the expression of FGF-23 polypeptide, and which act as anti-sense regulators of expression.

25 Once a test molecule has been identified as interacting with an FGF-23 polypeptide, the molecule may be further evaluated for its ability to increase or decrease FGF-23 polypeptide activity. The measurement of the interaction of a test molecule with FGF-23 polypeptide may be carried out in several formats, including cell-based binding assays, membrane binding assays, solution-phase
30 assays, and immunoassays. In general, a test molecule is incubated with an FGF-

23 polypeptide for a specified period of time, and FGF-23 polypeptide activity is determined by one or more assays for measuring biological activity.

The interaction of test molecules with FGF-23 polypeptides may also be assayed directly using polyclonal or monoclonal antibodies in an immunoassay.

5 Alternatively, modified forms of FGF-23 polypeptides containing epitope tags as described herein may be used in solution and immunoassays.

In the event that FGF-23 polypeptides display biological activity through an interaction with a binding partner (*e.g.*, a receptor or a ligand), a variety of *in vitro* assays may be used to measure the binding of an FGF-23 polypeptide to the
10 corresponding binding partner (such as a selective binding agent, receptor, or ligand). These assays may be used to screen test molecules for their ability to increase or decrease the rate and/or the extent of binding of an FGF-23 polypeptide to its binding partner. In one assay, an FGF-23 polypeptide is immobilized in the wells of a microtiter plate. Radiolabeled FGF-23 polypeptide
15 binding partner (for example, iodinated FGF-23 polypeptide binding partner) and a test molecule can then be added either one at a time (in either order) or simultaneously to the wells. After incubation, the wells can be washed and counted for radioactivity, using a scintillation counter, to determine the extent to which the binding partner bound to the FGF-23 polypeptide. Typically, a
20 molecule will be tested over a range of concentrations, and a series of control wells lacking one or more elements of the test assays can be used for accuracy in the evaluation of the results. An alternative to this method involves reversing the "positions" of the proteins, *i.e.*, immobilizing FGF-23 polypeptide binding partner to the microtiter plate wells, incubating with the test molecule and radiolabeled
25 FGF-23 polypeptide, and determining the extent of FGF-23 polypeptide binding. *See, e.g., Current Protocols in Molecular Biology*, chap. 18 (Ausubel *et al.*, eds., Green Publishers Inc. and Wiley and Sons 1995).

As an alternative to radiolabeling, an FGF-23 polypeptide or its binding partner may be conjugated to biotin, and the presence of biotinylated protein can
30 then be detected using streptavidin linked to an enzyme, such as horse radish peroxidase (HRP) or alkaline phosphatase (AP), which can be detected

colorometrically, or by fluorescent tagging of streptavidin. An antibody directed to an FGF-23 polypeptide or to an FGF-23 polypeptide binding partner, and which is conjugated to biotin, may also be used for purposes of detection following incubation of the complex with enzyme-linked streptavidin linked to
5 AP or HRP.

A FGF-23 polypeptide or an FGF-23 polypeptide binding partner can also be immobilized by attachment to agarose beads, acrylic beads, or other types of such inert solid phase substrates. The substrate-protein complex can be placed in a solution containing the complementary protein and the test compound. After
10 incubation, the beads can be precipitated by centrifugation, and the amount of binding between an FGF-23 polypeptide and its binding partner can be assessed using the methods described herein. Alternatively, the substrate-protein complex can be immobilized in a column with the test molecule and complementary protein passing through the column. The formation of a complex between an
15 FGF-23 polypeptide and its binding partner can then be assessed using any of the techniques described herein (*e.g.*, radiolabelling or antibody binding).

Another *in vitro* assay that is useful for identifying a test molecule which increases or decreases the formation of a complex between an FGF-23 polypeptide binding protein and an FGF-23 polypeptide binding partner is a
20 surface plasmon resonance detector system such as the BIAcore assay system (Pharmacia, Piscataway, NJ). The BIAcore system is utilized as specified by the manufacturer. This assay essentially involves the covalent binding of either FGF-23 polypeptide or an FGF-23 polypeptide binding partner to a dextran-coated sensor chip that is located in a detector. The test compound and the other
25 complementary protein can then be injected, either simultaneously or sequentially, into the chamber containing the sensor chip. The amount of complementary protein that binds can be assessed based on the change in molecular mass that is physically associated with the dextran-coated side of the sensor chip, with the change in molecular mass being measured by the detector system.

30 In some cases, it may be desirable to evaluate two or more test compounds together for their ability to increase or decrease the formation of a complex

between an FGF-23 polypeptide and an FGF-23 polypeptide binding partner. In these cases, the assays set forth herein can be readily modified by adding such additional test compound(s) either simultaneously with, or subsequent to, the first test compound. The remainder of the steps in the assay are as set forth herein.

5 *In vitro* assays such as those described herein may be used advantageously to screen large numbers of compounds for an effect on the formation of a complex between an FGF-23 polypeptide and FGF-23 polypeptide binding partner. The assays may be automated to screen compounds generated in phage display, synthetic peptide, and chemical synthesis libraries.

10 Compounds which increase or decrease the formation of a complex between an FGF-23 polypeptide and an FGF-23 polypeptide binding partner may also be screened in cell culture using cells and cell lines expressing either FGF-23 polypeptide or FGF-23 polypeptide binding partner. Cells and cell lines may be obtained from any mammal, but preferably will be from human or other primate,
15 canine, or rodent sources. The binding of an FGF-23 polypeptide to cells expressing FGF-23 polypeptide binding partner at the surface is evaluated in the presence or absence of test molecules, and the extent of binding may be determined by, for example, flow cytometry using a biotinylated antibody to an FGF-23 polypeptide binding partner. Cell culture assays can be used
20 advantageously to further evaluate compounds that score positive in protein binding assays described herein.

 Cell cultures can also be used to screen the impact of a drug candidate. For example, drug candidates may decrease or increase the expression of the FGF-23 gene. In certain embodiments, the amount of FGF-23 polypeptide or an
25 FGF-23 polypeptide fragment that is produced may be measured after exposure of the cell culture to the drug candidate. In certain embodiments, one may detect the actual impact of the drug candidate on the cell culture. For example, the over-expression of a particular gene may have a particular impact on the cell culture. In such cases, one may test a drug candidate's ability to increase or decrease the
30 expression of the gene or its ability to prevent or inhibit a particular impact on the cell culture. In other examples, the production of a particular metabolic product

such as a fragment of a polypeptide, may result in, or be associated with, a disease or pathological condition. In such cases, one may test a drug candidate's ability to decrease the production of such a metabolic product in a cell culture.

5 Internalizing Proteins

The *tat* protein sequence (from HIV) can be used to internalize proteins into a cell. See, e.g., Falwell *et al.*, 1994, *Proc. Natl. Acad. Sci. U.S.A.* 91:664-68. For example, an 11 amino acid sequence (Y-G-R-K-K-R-R-Q-R-R-R; SEQ ID NO: 40) of the HIV *tat* protein (termed the "protein transduction domain," or TAT
10 PDT) has been described as mediating delivery across the cytoplasmic membrane and the nuclear membrane of a cell. See Schwarze *et al.*, 1999, *Science* 285:1569-72; and Nagahara *et al.*, 1998, *Nat. Med.* 4:1449-52. In these procedures, FITC-constructs (FITC-labeled G-G-G-G-Y-G-R-K-K-R-R-Q-R-R-R; SEQ ID NO: 41), which penetrate tissues following intraperitoneal administration, are prepared, and
15 the binding of such constructs to cells is detected by fluorescence-activated cell sorting (FACS) analysis. Cells treated with a *tat*- β -gal fusion protein will demonstrate β -gal activity. Following injection, expression of such a construct can be detected in a number of tissues, including liver, kidney, lung, heart, and brain tissue. It is believed that such constructs undergo some degree of unfolding
20 in order to enter the cell, and as such, may require a refolding following entry into the cell.

It will thus be appreciated that the *tat* protein sequence may be used to internalize a desired polypeptide into a cell. For example, using the *tat* protein sequence, an FGF-23 antagonist (such as an anti-FGF-23 selective binding agent,
25 small molecule, soluble receptor, or antisense oligonucleotide) can be administered intracellularly to inhibit the activity of an FGF-23 molecule. As used herein, the term "FGF-23 molecule" refers to both FGF-23 nucleic acid molecules and FGF-23 polypeptides as defined herein. Where desired, the FGF-23 protein itself may also be internally administered to a cell using these
30 procedures. See also, Straus, 1999, *Science* 285:1466-67.

Cell Source Identification Using FGF-23 Polypeptide

In accordance with certain embodiments of the invention, it may be useful to be able to determine the source of a certain cell type associated with an FGF-23 polypeptide. For example, it may be useful to determine the origin of a disease or pathological condition as an aid in selecting an appropriate therapy. In certain
5 embodiments, nucleic acids encoding an FGF-23 polypeptide can be used as a probe to identify cells described herein by screening the nucleic acids of the cells with such a probe. In other embodiments, one may use anti-FGF-23 polypeptide antibodies to test for the presence of FGF-23 polypeptide in cells, and thus,
10 determine if such cells are of the types described herein.

FGF-23 Polypeptide Compositions and Administration

Therapeutic compositions are within the scope of the present invention. Such FGF-23 polypeptide pharmaceutical compositions may comprise a
15 therapeutically effective amount of an FGF-23 polypeptide or an FGF-23 nucleic acid molecule in admixture with a pharmaceutically or physiologically acceptable formulation agent selected for suitability with the mode of administration. Pharmaceutical compositions may comprise a therapeutically effective amount of one or more FGF-23 polypeptide selective binding agents in admixture with a
20 pharmaceutically or physiologically acceptable formulation agent selected for suitability with the mode of administration.

Acceptable formulation materials preferably are nontoxic to recipients at the dosages and concentrations employed.

The pharmaceutical composition may contain formulation materials for
25 modifying, maintaining, or preserving, for example, the pH, osmolarity, viscosity, clarity, color, isotonicity, odor, sterility, stability, rate of dissolution or release, adsorption, or penetration of the composition. Suitable formulation materials include, but are not limited to, amino acids (such as glycine, glutamine, asparagine, arginine, or lysine), antimicrobials, antioxidants (such as ascorbic
30 acid, sodium sulfite, or sodium hydrogen-sulfite), buffers (such as borate, bicarbonate, Tris-HCl, citrates, phosphates, or other organic acids), bulking agents

(such as mannitol or glycine), chelating agents (such as ethylenediamine tetraacetic acid (EDTA)), complexing agents (such as caffeine, polyvinylpyrrolidone, beta-cyclodextrin, or hydroxypropyl-beta-cyclodextrin), fillers, monosaccharides, disaccharides, and other carbohydrates (such as glucose, mannose, or dextrans), proteins (such as serum albumin, gelatin, or immunoglobulins), coloring, flavoring and diluting agents, emulsifying agents, hydrophilic polymers (such as polyvinylpyrrolidone), low molecular weight polypeptides, salt-forming counterions (such as sodium), preservatives (such as benzalkonium chloride, benzoic acid, salicylic acid, thimerosal, phenethyl alcohol, methylparaben, propylparaben, chlorhexidine, sorbic acid, or hydrogen peroxide), solvents (such as glycerin, propylene glycol, or polyethylene glycol), sugar alcohols (such as mannitol or sorbitol), suspending agents, surfactants or wetting agents (such as pluronics; PEG; sorbitan esters; polysorbates such as polysorbate 20 or polysorbate 80; triton; tromethamine; lecithin; cholesterol or tyloxapal), stability enhancing agents (such as sucrose or sorbitol), tonicity enhancing agents (such as alkali metal halides – preferably sodium or potassium chloride – or mannitol sorbitol), delivery vehicles, diluents, excipients and/or pharmaceutical adjuvants. *See Remington's Pharmaceutical Sciences* (18th Ed., A.R. Gennaro, ed., Mack Publishing Company 1990.

The optimal pharmaceutical composition will be determined by a skilled artisan depending upon, for example, the intended route of administration, delivery format, and desired dosage. *See, e.g., Remington's Pharmaceutical Sciences, supra.* Such compositions may influence the physical state, stability, rate of *in vivo* release, and rate of *in vivo* clearance of the FGF-23 molecule.

The primary vehicle or carrier in a pharmaceutical composition may be either aqueous or non-aqueous in nature. For example, a suitable vehicle or carrier for injection may be water, physiological saline solution, or artificial cerebrospinal fluid, possibly supplemented with other materials common in compositions for parenteral administration. Neutral buffered saline or saline mixed with serum albumin are further exemplary vehicles. Other exemplary pharmaceutical compositions comprise Tris buffer of about pH 7.0-8.5, or acetate

buffer of about pH 4.0-5.5, which may further include sorbitol or a suitable substitute. In one embodiment of the present invention, FGF-23 polypeptide compositions may be prepared for storage by mixing the selected composition having the desired degree of purity with optional formulation agents (*Remington's*
5 *Pharmaceutical Sciences, supra*) in the form of a lyophilized cake or an aqueous solution. Further, the FGF-23 polypeptide product may be formulated as a lyophilizate using appropriate excipients such as sucrose.

The FGF-23 polypeptide pharmaceutical compositions can be selected for parenteral delivery. Alternatively, the compositions may be selected for
10 inhalation or for delivery through the digestive tract, such as orally. The preparation of such pharmaceutically acceptable compositions is within the skill of the art.

The formulation components are present in concentrations that are acceptable to the site of administration. For example, buffers are used to maintain
15 the composition at physiological pH or at a slightly lower pH, typically within a pH range of from about 5 to about 8.

When parenteral administration is contemplated, the therapeutic compositions for use in this invention may be in the form of a pyrogen-free, parenterally acceptable, aqueous solution comprising the desired FGF-23
20 molecule in a pharmaceutically acceptable vehicle. A particularly suitable vehicle for parenteral injection is sterile distilled water in which an FGF-23 molecule is formulated as a sterile, isotonic solution, properly preserved. Yet another preparation can involve the formulation of the desired molecule with an agent, such as injectable microspheres, bio-erodible particles, polymeric compounds
25 (such as polylactic acid or polyglycolic acid), beads, or liposomes, that provides for the controlled or sustained release of the product which may then be delivered via a depot injection. Hyaluronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Other suitable means for the introduction of the desired molecule include implantable drug delivery
30 devices.

In one embodiment, a pharmaceutical composition may be formulated for inhalation. For example, FGF-23 polypeptide may be formulated as a dry powder for inhalation. FGF-23 polypeptide or nucleic acid molecule inhalation solutions may also be formulated with a propellant for aerosol delivery. In yet another
5 embodiment, solutions may be nebulized. Pulmonary administration is further described in PCT Pub. No. WO 94/20069, which describes the pulmonary delivery of chemically modified proteins.

It is also contemplated that certain formulations may be administered orally. In one embodiment of the present invention, FGF-23 polypeptides that are
10 administered in this fashion can be formulated with or without those carriers customarily used in the compounding of solid dosage forms such as tablets and capsules. For example, a capsule may be designed to release the active portion of the formulation at the point in the gastrointestinal tract when bioavailability is maximized and pre-systemic degradation is minimized. Additional agents can be
15 included to facilitate absorption of the FGF-23 polypeptide. Diluents, flavorings, low melting point waxes, vegetable oils, lubricants, suspending agents, tablet disintegrating agents, and binders may also be employed.

Another pharmaceutical composition may involve an effective quantity of FGF-23 polypeptides in a mixture with non-toxic excipients that are suitable for
20 the manufacture of tablets. By dissolving the tablets in sterile water, or another appropriate vehicle, solutions can be prepared in unit-dose form. Suitable excipients include, but are not limited to, inert diluents, such as calcium carbonate, sodium carbonate or bicarbonate, lactose, or calcium phosphate; or binding agents, such as starch, gelatin, or acacia; or lubricating agents such as
25 magnesium stearate, stearic acid, or talc.

Additional FGF-23 polypeptide pharmaceutical compositions will be evident to those skilled in the art, including formulations involving FGF-23 polypeptides in sustained- or controlled-delivery formulations. Techniques for formulating a variety of other sustained- or controlled-delivery means, such as
30 liposome carriers, bio-erodible microparticles or porous beads and depot injections, are also known to those skilled in the art. *See, e.g.*, PCT/US93/00829,

which describes the controlled release of porous polymeric microparticles for the delivery of pharmaceutical compositions.

Additional examples of sustained-release preparations include semipermeable polymer matrices in the form of shaped articles, *e.g.* films, or microcapsules. Sustained release matrices may include polyesters, hydrogels, 5 polylactides (U.S. Patent No. 3,773,919 and European Patent No. 058481), copolymers of L-glutamic acid and gamma ethyl-L-glutamate (Sidman *et al.*, 1983, *Biopolymers* 22:547-56), poly(2-hydroxyethyl-methacrylate) (Langer *et al.*, 1981, *J. Biomed. Mater. Res.* 15:167-277 and Langer, 1982, *Chem. Tech.* 12:98-10 105), ethylene vinyl acetate (Langer *et al.*, *supra*) or poly-D(-)-3-hydroxybutyric acid (European Patent No. 133988). Sustained-release compositions may also include liposomes, which can be prepared by any of several methods known in the art. *See, e.g.*, Eppstein *et al.*, 1985, *Proc. Natl. Acad. Sci. USA* 82:3688-92; and European Patent Nos. 036676, 088046, and 143949.

15 The FGF-23 pharmaceutical composition to be used for *in vivo* administration typically must be sterile. This may be accomplished by filtration through sterile filtration membranes. Where the composition is lyophilized, sterilization using this method may be conducted either prior to, or following, lyophilization and reconstitution. The composition for parenteral administration 20 may be stored in lyophilized form or in a solution. In addition, parenteral compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle.

Once the pharmaceutical composition has been formulated, it may be 25 stored in sterile vials as a solution, suspension, gel, emulsion, solid, or as a dehydrated or lyophilized powder. Such formulations may be stored either in a ready-to-use form or in a form (*e.g.*, lyophilized) requiring reconstitution prior to administration.

In a specific embodiment, the present invention is directed to kits for 30 producing a single-dose administration unit. The kits may each contain both a first container having a dried protein and a second container having an aqueous

formulation. Also included within the scope of this invention are kits containing single and multi-chambered pre-filled syringes (e.g., liquid syringes and lyosyringes).

The effective amount of an FGF-23 pharmaceutical composition to be employed therapeutically will depend, for example, upon the therapeutic context and objectives. One skilled in the art will appreciate that the appropriate dosage levels for treatment will thus vary depending, in part, upon the molecule delivered, the indication for which the FGF-23 molecule is being used, the route of administration, and the size (body weight, body surface, or organ size) and condition (the age and general health) of the patient. Accordingly, the clinician may titer the dosage and modify the route of administration to obtain the optimal therapeutic effect. A typical dosage may range from about 0.1 $\mu\text{g/kg}$ to up to about 100 mg/kg or more, depending on the factors mentioned above. In other embodiments, the dosage may range from 0.1 $\mu\text{g/kg}$ up to about 100 mg/kg; or 1 $\mu\text{g/kg}$ up to about 100 mg/kg; or 5 $\mu\text{g/kg}$ up to about 100 mg/kg.

The frequency of dosing will depend upon the pharmacokinetic parameters of the FGF-23 molecule in the formulation being used. Typically, a clinician will administer the composition until a dosage is reached that achieves the desired effect. The composition may therefore be administered as a single dose, as two or more doses (which may or may not contain the same amount of the desired molecule) over time, or as a continuous infusion via an implantation device or catheter. Further refinement of the appropriate dosage is routinely made by those of ordinary skill in the art and is within the ambit of tasks routinely performed by them. Appropriate dosages may be ascertained through use of appropriate dose-response data.

The route of administration of the pharmaceutical composition is in accord with known methods, e.g., orally; through injection by intravenous, intraperitoneal, intracerebral (intraparenchymal), intracerebroventricular, intramuscular, intraocular, intraarterial, intraportal, or intralesional routes; by sustained release systems; or by implantation devices. Where desired, the

compositions may be administered by bolus injection or continuously by infusion, or by implantation device.

Alternatively or additionally, the composition may be administered locally via implantation of a membrane, sponge, or other appropriate material onto which the desired molecule has been absorbed or encapsulated. Where an implantation device is used, the device may be implanted into any suitable tissue or organ, and delivery of the desired molecule may be via diffusion, timed-release bolus, or continuous administration.

In some cases, it may be desirable to use FGF-23 polypeptide pharmaceutical compositions in an *ex vivo* manner. In such instances, cells, tissues, or organs that have been removed from the patient are exposed to FGF-23 polypeptide pharmaceutical compositions after which the cells, tissues, or organs are subsequently implanted back into the patient.

In other cases, an FGF-23 polypeptide can be delivered by implanting certain cells that have been genetically engineered, using methods such as those described herein, to express and secrete the FGF-23 polypeptide. Such cells may be animal or human cells, and may be autologous, heterologous, or xenogeneic. Optionally, the cells may be immortalized. In order to decrease the chance of an immunological response, the cells may be encapsulated to avoid infiltration of surrounding tissues. The encapsulation materials are typically biocompatible, semi-permeable polymeric enclosures or membranes that allow the release of the protein product(s) but prevent the destruction of the cells by the patient's immune system or by other detrimental factors from the surrounding tissues.

As discussed herein, it may be desirable to treat isolated cell populations (such as stem cells, lymphocytes, red blood cells, chondrocytes, neurons, and the like) with one or more FGF-23 polypeptides. This can be accomplished by exposing the isolated cells to the polypeptide directly, where it is in a form that is permeable to the cell membrane.

Additional embodiments of the present invention relate to cells and methods (e.g., homologous recombination and/or other recombinant production methods) for both the *in vitro* production of therapeutic polypeptides and for the

production and delivery of therapeutic polypeptides by gene therapy or cell therapy. Homologous and other recombination methods may be used to modify a cell that contains a normally transcriptionally-silent FGF-23 gene, or an under-expressed gene, and thereby produce a cell which expresses therapeutically efficacious amounts of FGF-23 polypeptides.

Homologous recombination is a technique originally developed for targeting genes to induce or correct mutations in transcriptionally active genes. Kucherlapati, 1989, *Prog. in Nucl. Acid Res. & Mol. Biol.* 36:301. The basic technique was developed as a method for introducing specific mutations into specific regions of the mammalian genome (Thomas *et al.*, 1986, *Cell* 44:419-28; Thomas and Capecchi, 1987, *Cell* 51:503-12; Doetschman *et al.*, 1988, *Proc. Natl. Acad. Sci. U.S.A.* 85:8583-87) or to correct specific mutations within defective genes (Doetschman *et al.*, 1987, *Nature* 330:576-78). Exemplary homologous recombination techniques are described in U.S. Patent No. 5,272,071; European Patent Nos. 9193051 and 505500; PCT/US90/07642, and PCT Pub No. WO 91/09955).

Through homologous recombination, the DNA sequence to be inserted into the genome can be directed to a specific region of the gene of interest by attaching it to targeting DNA. The targeting DNA is a nucleotide sequence that is complementary (homologous) to a region of the genomic DNA. Small pieces of targeting DNA that are complementary to a specific region of the genome are put in contact with the parental strand during the DNA replication process. It is a general property of DNA that has been inserted into a cell to hybridize, and therefore, recombine with other pieces of endogenous DNA through shared homologous regions. If this complementary strand is attached to an oligonucleotide that contains a mutation or a different sequence or an additional nucleotide, it too is incorporated into the newly synthesized strand as a result of the recombination. As a result of the proofreading function, it is possible for the new sequence of DNA to serve as the template. Thus, the transferred DNA is incorporated into the genome.

Attached to these pieces of targeting DNA are regions of DNA that may interact with or control the expression of an FGF-23 polypeptide, *e.g.*, flanking sequences. For example, a promoter/enhancer element, a suppressor, or an exogenous transcription modulatory element is inserted in the genome of the intended host cell in proximity and orientation sufficient to influence the transcription of DNA encoding the desired FGF-23 polypeptide. The control element controls a portion of the DNA present in the host cell genome. Thus, the expression of the desired FGF-23 polypeptide may be achieved not by transfection of DNA that encodes the FGF-23 gene itself, but rather by the use of targeting DNA (containing regions of homology with the endogenous gene of interest) coupled with DNA regulatory segments that provide the endogenous gene sequence with recognizable signals for transcription of an FGF-23 gene.

In an exemplary method, the expression of a desired targeted gene in a cell (*i.e.*, a desired endogenous cellular gene) is altered via homologous recombination into the cellular genome at a preselected site, by the introduction of DNA which includes at least a regulatory sequence, an exon, and a splice donor site. These components are introduced into the chromosomal (genomic) DNA in such a manner that this, in effect, results in the production of a new transcription unit (in which the regulatory sequence, the exon, and the splice donor site present in the DNA construct are operatively linked to the endogenous gene). As a result of the introduction of these components into the chromosomal DNA, the expression of the desired endogenous gene is altered.

Altered gene expression, as described herein, encompasses activating (or causing to be expressed) a gene which is normally silent (unexpressed) in the cell as obtained, as well as increasing the expression of a gene which is not expressed at physiologically significant levels in the cell as obtained. The embodiments further encompass changing the pattern of regulation or induction such that it is different from the pattern of regulation or induction that occurs in the cell as obtained, and reducing (including eliminating) the expression of a gene which is expressed in the cell as obtained.

One method by which homologous recombination can be used to increase, or cause, FGF-23 polypeptide production from a cell's endogenous FGF-23 gene involves first using homologous recombination to place a recombination sequence from a site-specific recombination system (e.g., Cre/loxP, FLP/FRT) (Sauer, 1994, *Curr. Opin. Biotechnol.*, 5:521-27; Sauer, 1993, *Methods Enzymol.*, 225:890-900) upstream of (i.e., 5' to) the cell's endogenous genomic FGF-23 polypeptide coding region. A plasmid containing a recombination site homologous to the site that was placed just upstream of the genomic FGF-23 polypeptide coding region is introduced into the modified cell line along with the appropriate recombinase enzyme. This recombinase causes the plasmid to integrate, via the plasmid's recombination site, into the recombination site located just upstream of the genomic FGF-23 polypeptide coding region in the cell line (Baubonis and Sauer, 1993, *Nucleic Acids Res.* 21:2025-29; O'Gorman *et al.*, 1991, *Science* 251:1351-55). Any flanking sequences known to increase transcription (e.g., enhancer/promoter, intron, translational enhancer), if properly positioned in this plasmid, would integrate in such a manner as to create a new or modified transcriptional unit resulting in *de novo* or increased FGF-23 polypeptide production from the cell's endogenous FGF-23 gene.

A further method to use the cell line in which the site specific recombination sequence had been placed just upstream of the cell's endogenous genomic FGF-23 polypeptide coding region is to use homologous recombination to introduce a second recombination site elsewhere in the cell line's genome. The appropriate recombinase enzyme is then introduced into the two-recombination-site cell line, causing a recombination event (deletion, inversion, and translocation) (Sauer, 1994, *Curr. Opin. Biotechnol.*, 5:521-27; Sauer, 1993, *Methods Enzymol.*, 225:890-900) that would create a new or modified transcriptional unit resulting in *de novo* or increased FGF-23 polypeptide production from the cell's endogenous FGF-23 gene.

An additional approach for increasing, or causing, the expression of FGF-23 polypeptide from a cell's endogenous FGF-23 gene involves increasing, or causing, the expression of a gene or genes (e.g., transcription factors) and/or

decreasing the expression of a gene or genes (e.g., transcriptional repressors) in a manner which results in *de novo* or increased FGF-23 polypeptide production from the cell's endogenous FGF-23 gene. This method includes the introduction of a non-naturally occurring polypeptide (e.g., a polypeptide comprising a site
5 specific DNA binding domain fused to a transcriptional factor domain) into the cell such that *de novo* or increased FGF-23 polypeptide production from the cell's endogenous FGF-23 gene results.

The present invention further relates to DNA constructs useful in the method of altering expression of a target gene. In certain embodiments, the
10 exemplary DNA constructs comprise: (a) one or more targeting sequences, (b) a regulatory sequence, (c) an exon, and (d) an unpaired splice-donor site. The targeting sequence in the DNA construct directs the integration of elements (a) - (d) into a target gene in a cell such that the elements (b) - (d) are operatively linked to sequences of the endogenous target gene. In another embodiment, the
15 DNA constructs comprise: (a) one or more targeting sequences, (b) a regulatory sequence, (c) an exon, (d) a splice-donor site, (e) an intron, and (f) a splice-acceptor site, wherein the targeting sequence directs the integration of elements (a) - (f) such that the elements of (b) - (f) are operatively linked to the endogenous gene. The targeting sequence is homologous to the preselected site in the cellular
20 chromosomal DNA with which homologous recombination is to occur. In the construct, the exon is generally 3' of the regulatory sequence and the splice-donor site is 3' of the exon.

If the sequence of a particular gene is known, such as the nucleic acid sequence of FGF-23 polypeptide presented herein, a piece of DNA that is
25 complementary to a selected region of the gene can be synthesized or otherwise obtained, such as by appropriate restriction of the native DNA at specific recognition sites bounding the region of interest. This piece serves as a targeting sequence upon insertion into the cell and will hybridize to its homologous region within the genome. If this hybridization occurs during DNA replication, this piece
30 of DNA, and any additional sequence attached thereto, will act as an Okazaki fragment and will be incorporated into the newly synthesized daughter strand of

DNA. The present invention, therefore, includes nucleotides encoding an FGF-23 polypeptide, which nucleotides may be used as targeting sequences.

FGF-23 polypeptide cell therapy, *e.g.*, the implantation of cells producing FGF-23 polypeptides, is also contemplated. This embodiment involves
5 implanting cells capable of synthesizing and secreting a biologically active form of FGF-23 polypeptide. Such FGF-23 polypeptide-producing cells can be cells that are natural producers of FGF-23 polypeptides or may be recombinant cells whose ability to produce FGF-23 polypeptides has been augmented by transformation with a gene encoding the desired FGF-23 polypeptide or with a
10 gene augmenting the expression of FGF-23 polypeptide. Such a modification may be accomplished by means of a vector suitable for delivering the gene as well as promoting its expression and secretion. In order to minimize a potential immunological reaction in patients being administered an FGF-23 polypeptide, as may occur with the administration of a polypeptide of a foreign species, it is
15 preferred that the natural cells producing FGF-23 polypeptide be of human origin and produce human FGF-23 polypeptide. Likewise, it is preferred that the recombinant cells producing FGF-23 polypeptide be transformed with an expression vector containing a gene encoding a human FGF-23 polypeptide.

Implanted cells may be encapsulated to avoid the infiltration of
20 surrounding tissue. Human or non-human animal cells may be implanted in patients in biocompatible, semipermeable polymeric enclosures or membranes that allow the release of FGF-23 polypeptide, but that prevent the destruction of the cells by the patient's immune system or by other detrimental factors from the surrounding tissue. Alternatively, the patient's own cells, transformed to produce
25 FGF-23 polypeptides *ex vivo*, may be implanted directly into the patient without such encapsulation.

Techniques for the encapsulation of living cells are known in the art, and the preparation of the encapsulated cells and their implantation in patients may be routinely accomplished. For example, Baetge *et al.* (PCT Pub. No. WO 95/05452
30 and PCT/US94/09299) describe membrane capsules containing genetically engineered cells for the effective delivery of biologically active molecules. The

capsules are biocompatible and are easily retrievable. The capsules encapsulate cells transfected with recombinant DNA molecules comprising DNA sequences coding for biologically active molecules operatively linked to promoters that are not subject to down-regulation *in vivo* upon implantation into a mammalian host.

5 The devices provide for the delivery of the molecules from living cells to specific sites within a recipient. In addition, *see* U.S. Patent Nos. 4,892,538; 5,011,472; and 5,106,627. A system for encapsulating living cells is described in PCT Pub. No. WO 91/10425 (Aebischer *et al.*). *See also*, PCT Pub. No. WO 91/10470 (Aebischer *et al.*); Winn *et al.*, 1991, *Exper. Neurol.* 113:322-29; Aebischer *et al.*,
10 1991, *Exper. Neurol.* 111:269-75; and Tresco *et al.*, 1992, *ASAIO* 38:17-23.

In vivo and *in vitro* gene therapy delivery of FGF-23 polypeptides is also envisioned. One example of a gene therapy technique is to use the FGF-23 gene (either genomic DNA, cDNA, and/or synthetic DNA) encoding an FGF-23 polypeptide which may be operably linked to a constitutive or inducible promoter
15 to form a "gene therapy DNA construct." The promoter may be homologous or heterologous to the endogenous FGF-23 gene, provided that it is active in the cell or tissue type into which the construct will be inserted. Other components of the gene therapy DNA construct may optionally include DNA molecules designed for site-specific integration (*e.g.*, endogenous sequences useful for homologous
20 recombination), tissue-specific promoters, enhancers or silencers, DNA molecules capable of providing a selective advantage over the parent cell, DNA molecules useful as labels to identify transformed cells, negative selection systems, cell specific binding agents (as, for example, for cell targeting), cell-specific internalization factors, transcription factors enhancing expression from a vector,
25 and factors enabling vector production.

A gene therapy DNA construct can then be introduced into cells (either *ex vivo* or *in vivo*) using viral or non-viral vectors. One means for introducing the gene therapy DNA construct is by means of viral vectors as described herein. Certain vectors, such as retroviral vectors, will deliver the DNA construct to the
30 chromosomal DNA of the cells, and the gene can integrate into the chromosomal DNA. Other vectors will function as episomes, and the gene therapy DNA

construct will remain in the cytoplasm.

In yet other embodiments, regulatory elements can be included for the controlled expression of the FGF-23 gene in the target cell. Such elements are turned on in response to an appropriate effector. In this way, a therapeutic polypeptide can be expressed when desired. One conventional control means involves the use of small molecule dimerizers or rapalogs to dimerize chimeric proteins which contain a small molecule-binding domain and a domain capable of initiating a biological process, such as a DNA-binding protein or transcriptional activation protein (*see* PCT Pub. Nos. WO 96/41865, WO 97/31898, and WO 97/31899). The dimerization of the proteins can be used to initiate transcription of the transgene.

An alternative regulation technology uses a method of storing proteins expressed from the gene of interest inside the cell as an aggregate or cluster. The gene of interest is expressed as a fusion protein that includes a conditional aggregation domain that results in the retention of the aggregated protein in the endoplasmic reticulum. The stored proteins are stable and inactive inside the cell. The proteins can be released, however, by administering a drug (*e.g.*, small molecule ligand) that removes the conditional aggregation domain and thereby specifically breaks apart the aggregates or clusters so that the proteins may be secreted from the cell. *See* Aridor *et al.*, 2000, *Science* 287:816-17 and Rivera *et al.*, 2000, *Science* 287:826-30.

Other suitable control means or gene switches include, but are not limited to, the systems described herein. Mifepristone (RU486) is used as a progesterone antagonist. The binding of a modified progesterone receptor ligand-binding domain to the progesterone antagonist activates transcription by forming a dimer of two transcription factors that then pass into the nucleus to bind DNA. The ligand-binding domain is modified to eliminate the ability of the receptor to bind to the natural ligand. The modified steroid hormone receptor system is further described in U.S. Patent No. 5,364,791 and PCT Pub. Nos. WO 96/40911 and WO 97/10337.

Yet another control system uses ecdysone (a fruit fly steroid hormone) which binds to and activates an ecdysone receptor (cytoplasmic receptor). The receptor then translocates to the nucleus to bind a specific DNA response element (promoter from ecdysone-responsive gene). The ecdysone receptor includes a transactivation domain, DNA-binding domain, and ligand-binding domain to initiate transcription. The ecdysone system is further described in U.S. Patent No. 5,514,578 and PCT Pub. Nos. WO 97/38117, WO 96/37609, and WO 93/03162.

Another control means uses a positive tetracycline-controllable transactivator. This system involves a mutated tet repressor protein DNA-binding domain (mutated tet R-4 amino acid changes which resulted in a reverse tetracycline-regulated transactivator protein, *i.e.*, it binds to a tet operator in the presence of tetracycline) linked to a polypeptide which activates transcription. Such systems are described in U.S. Patent Nos. 5,464,758, 5,650,298, and 5,654,168.

Additional expression control systems and nucleic acid constructs are described in U.S. Patent Nos. 5,741,679 and 5,834,186, to Innovir Laboratories Inc.

In vivo gene therapy may be accomplished by introducing the gene encoding FGF-23 polypeptide into cells via local injection of an FGF-23 nucleic acid molecule or by other appropriate viral or non-viral delivery vectors. Hefti, 1994, *Neurobiology* 25:1418-35. For example, a nucleic acid molecule encoding an FGF-23 polypeptide may be contained in an adeno-associated virus (AAV) vector for delivery to the targeted cells (*see, e.g.*, Johnson, PCT Pub. No. WO 95/34670; PCT App. No. PCT/US95/07178). The recombinant AAV genome typically contains AAV inverted terminal repeats flanking a DNA sequence encoding an FGF-23 polypeptide operably linked to functional promoter and polyadenylation sequences.

Alternative suitable viral vectors include, but are not limited to, retrovirus, adenovirus, herpes simplex virus, lentivirus, hepatitis virus, parvovirus, papovavirus, poxvirus, alphavirus, coronavirus, rhabdovirus, paramyxovirus, and papilloma virus vectors. U.S. Patent No. 5,672,344 describes an *in vivo* viral-

mediated gene transfer system involving a recombinant neurotrophic HSV-1 vector. U.S. Patent No. 5,399,346 provides examples of a process for providing a patient with a therapeutic protein by the delivery of human cells which have been treated *in vitro* to insert a DNA segment encoding a therapeutic protein.

5 Additional methods and materials for the practice of gene therapy techniques are described in U.S. Patent Nos. 5,631,236 (involving adenoviral vectors), 5,672,510 (involving retroviral vectors), 5,635,399 (involving retroviral vectors expressing cytokines).

Nonviral delivery methods include, but are not limited to, liposome-mediated transfer, naked DNA delivery (direct injection), receptor-mediated transfer (ligand-DNA complex), electroporation, calcium phosphate precipitation, and microparticle bombardment (*e.g.*, gene gun). Gene therapy materials and methods may also include inducible promoters, tissue-specific enhancer-promoters, DNA sequences designed for site-specific integration, DNA sequences

15 capable of providing a selective advantage over the parent cell, labels to identify transformed cells, negative selection systems and expression control systems (safety measures), cell-specific binding agents (for cell targeting), cell-specific internalization factors, and transcription factors to enhance expression by a vector as well as methods of vector manufacture. Such additional methods and materials

20 for the practice of gene therapy techniques are described in U.S. Patent Nos. 4,970,154 (involving electroporation techniques), 5,679,559 (describing a lipoprotein-containing system for gene delivery), 5,676,954 (involving liposome carriers), 5,593,875 (describing methods for calcium phosphate transfection), and 4,945,050 (describing a process wherein biologically active particles are propelled

25 at cells at a speed whereby the particles penetrate the surface of the cells and become incorporated into the interior of the cells), and PCT Pub. No. WO 96/40958 (involving nuclear ligands).

It is also contemplated that FGF-23 gene therapy or cell therapy can further include the delivery of one or more additional polypeptide(s) in the same

30 or a different cell(s). Such cells may be separately introduced into the patient, or the cells may be contained in a single implantable device, such as the

encapsulating membrane described above, or the cells may be separately modified by means of viral vectors.

A means to increase endogenous FGF-23 polypeptide expression in a cell via gene therapy is to insert one or more enhancer elements into the FGF-23 polypeptide promoter, where the enhancer elements can serve to increase transcriptional activity of the FGF-23 gene. The enhancer elements used will be selected based on the tissue in which one desires to activate the gene – enhancer elements known to confer promoter activation in that tissue will be selected. For example, if a gene encoding an FGF-23 polypeptide is to be “turned on” in T-cells, the *lck* promoter enhancer element may be used. Here, the functional portion of the transcriptional element to be added may be inserted into a fragment of DNA containing the FGF-23 polypeptide promoter (and optionally, inserted into a vector and/or 5’ and/or 3’ flanking sequences) using standard cloning techniques. This construct, known as a “homologous recombination construct,” can then be introduced into the desired cells either *ex vivo* or *in vivo*.

Gene therapy also can be used to decrease FGF-23 polypeptide expression by modifying the nucleotide sequence of the endogenous promoter. Such modification is typically accomplished via homologous recombination methods. For example, a DNA molecule containing all or a portion of the promoter of the FGF-23 gene selected for inactivation can be engineered to remove and/or replace pieces of the promoter that regulate transcription. For example, the TATA box and/or the binding site of a transcriptional activator of the promoter may be deleted using standard molecular biology techniques; such deletion can inhibit promoter activity thereby repressing the transcription of the corresponding FGF-23 gene. The deletion of the TATA box or the transcription activator binding site in the promoter may be accomplished by generating a DNA construct comprising all or the relevant portion of the FGF-23 polypeptide promoter (from the same or a related species as the FGF-23 gene to be regulated) in which one or more of the TATA box and/or transcriptional activator binding site nucleotides are mutated via substitution, deletion and/or insertion of one or more nucleotides. As a result, the TATA box and/or activator binding site has decreased activity or is rendered

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completely inactive. This construct, which also will typically contain at least about 500 bases of DNA that correspond to the native (endogenous) 5' and 3' DNA sequences adjacent to the promoter segment that has been modified, may be introduced into the appropriate cells (either *ex vivo* or *in vivo*) either directly or via a viral vector as described herein. Typically, the integration of the construct into the genomic DNA of the cells will be via homologous recombination, where the 5' and 3' DNA sequences in the promoter construct can serve to help integrate the modified promoter region via hybridization to the endogenous chromosomal DNA.

Therapeutic Uses

FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof can be used to treat, diagnose, ameliorate, or prevent a number of diseases, disorders, or conditions, including those recited herein.

FGF-23 polypeptide agonists and antagonists include those molecules which regulate FGF-23 polypeptide activity and either increase or decrease at least one activity of the mature form of the FGF-23 polypeptide. Agonists or antagonists may be co-factors, such as a protein, peptide, carbohydrate, lipid, or small molecular weight molecule, which interact with FGF-23 polypeptide and thereby regulate its activity. Potential polypeptide agonists or antagonists include antibodies that react with either soluble or membrane-bound forms of FGF-23 polypeptides that comprise part or all of the extracellular domains of the said proteins. Molecules that regulate FGF-23 polypeptide expression typically include nucleic acids encoding FGF-23 polypeptide that can act as anti-sense regulators of expression.

The FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof of the present invention are useful for the same purposes for which members of the FGF family of polypeptides are known to be useful. Thus, the FGF-23 polypeptides of this invention are potent mitogens for a variety of cells of the mesodermal, ectodermal, and endodermal origin, including fibroblasts, corneal and vascular endothelial cells, granulocytes, adrenal cortical cells,

chondrocytes, myoblasts, vascular smooth muscle cells, lens epithelial cells, melanocytes, keratinocytes, oligodendrocytes, astrocytes, osteoblasts, and hematopoietic cells. Included among these biological activities are the ability to stimulate the proliferation of vascular endothelial cells and to enable endothelial cells to penetrate the basement membrane. Consistent with these properties, the FGF-23 polypeptides of this invention may stimulate angiogenesis and promote wound healing (*i.e.*, facilitate the repair or replacement of damages of diseased tissue resulting from burns, traumatic injuries, surgery, or ulcers). These polypeptides may also induce mesoderm formation and modulate the differentiation of neuronal cells, adipocytes, and skeletal muscle cells. The polypeptides may also be employed to prevent or ameliorate skin aging due to sun exposure by stimulating keratinocyte growth. Further, the polypeptides of this invention may be employed to maintain organs before transplantation or for supporting cultures of primary cells and tissues. In addition, these polypeptides may be utilized to prevent hair loss since FGF family members activate hair-forming cells and promote melanocyte growth. They may also be used to stimulate the growth and differentiation of hematopoietic cells and bone marrow cells when used in combination with other cytokines.

FGF-23 has been linked with a human autosomal dominant genetic disease, hypophosphatemic rickets (ADHR) (The ADHR Consortium, 2000, *Nature Genetics* 26:345-48). Accordingly, the FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof of the present invention may be used to treat, diagnose, ameliorate, or prevent ADHR.

The FGF-23 gene has been shown to be most closely related to human FGF-21 (Yamashita *et al.*, 2000, *Biochem. Biophys. Res. Commun.* 277:494-98), a gene which is expressed most abundantly in the liver and at lower levels in the thymus (Nishimura *et al.*, 2000, *Biochim. Biophys. Acta* 1492:203-06). Accordingly, FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof may be used to treat, diagnose, ameliorate, or prevent diseases, disorders, or conditions involving the liver or thymus.

A non-exclusive list of other diseases, disorders, or conditions which may be treated, diagnosed, ameliorated, or prevented with the FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof of the present invention include: dermal wounds, epidermalysis bullosa, male pattern alopecia,
5 gastric ulcer, duodenal ulcer, erosive gastritis, esophagitis, esophageal reflux disease, inflammatory bowel disease, radiation- or chemotherapy-induced gut toxicity, hyaline membrane disease, necrosis of the respiratory epithelium, emphysema, pulmonary inflammation, pulmonary fibrosis, hepatic cirrhosis, fulminant liver failure, viral hepatitis, and diabetes.

10 Agonists or antagonists of FGF-23 polypeptide function may be used (simultaneously or sequentially) in combination with one or more cytokines, growth factors, antibiotics, anti-inflammatories, and/or chemotherapeutic agents as is appropriate for the condition being treated.

Other diseases caused by or mediated by undesirable levels of FGF-23
15 polypeptides are encompassed within the scope of the invention. Undesirable levels include excessive levels of FGF-23 polypeptides and sub-normal levels of FGF-23 polypeptides.

Uses of FGF-23 Nucleic Acids and Polypeptides

20 Nucleic acid molecules of the invention (including those that do not themselves encode biologically active polypeptides) may be used to map the locations of the FGF-23 gene and related genes on chromosomes. Mapping may be done by techniques known in the art, such as PCR amplification and *in situ* hybridization.

25 FGF-23 nucleic acid molecules (including those that do not themselves encode biologically active polypeptides), may be useful as hybridization probes in diagnostic assays to test, either qualitatively or quantitatively, for the presence of an FGF-23 nucleic acid molecule in mammalian tissue or bodily fluid samples.

Other methods may also be employed where it is desirable to inhibit the
30 activity of one or more FGF-23 polypeptides. Such inhibition may be effected by nucleic acid molecules that are complementary to and hybridize to expression

control sequences (triple helix formation) or to FGF-23 mRNA. For example, antisense DNA or RNA molecules, which have a sequence that is complementary to at least a portion of an FGF-23 gene can be introduced into the cell. Anti-sense probes may be designed by available techniques using the sequence of the FGF-23 gene disclosed herein. Typically, each such antisense molecule will be complementary to the start site (5' end) of each selected FGF-23 gene. When the antisense molecule then hybridizes to the corresponding FGF-23 mRNA, translation of this mRNA is prevented or reduced. Anti-sense inhibitors provide information relating to the decrease or absence of an FGF-23 polypeptide in a cell or organism.

Alternatively, gene therapy may be employed to create a dominant-negative inhibitor of one or more FGF-23 polypeptides. In this situation, the DNA encoding a mutant polypeptide of each selected FGF-23 polypeptide can be prepared and introduced into the cells of a patient using either viral or non-viral methods as described herein. Each such mutant is typically designed to compete with endogenous polypeptide in its biological role.

In addition, an FGF-23 polypeptide, whether biologically active or not, may be used as an immunogen, that is, the polypeptide contains at least one epitope to which antibodies may be raised. Selective binding agents that bind to an FGF-23 polypeptide (as described herein) may be used for *in vivo* and *in vitro* diagnostic purposes, including, but not limited to, use in labeled form to detect the presence of FGF-23 polypeptide in a body fluid or cell sample. The antibodies may also be used to prevent, treat, or diagnose a number of diseases and disorders, including those recited herein. The antibodies may bind to an FGF-23 polypeptide so as to diminish or block at least one activity characteristic of an FGF-23 polypeptide, or may bind to a polypeptide to increase at least one activity characteristic of an FGF-23 polypeptide (including by increasing the pharmacokinetics of the FGF-23 polypeptide).

The FGF-23 polypeptides of the present invention can be used to clone FGF-23 polypeptide receptors, using an expression cloning strategy. Radiolabeled (¹²⁵Iodine) FGF-23 polypeptide or affinity/activity-tagged FGF-23

polypeptide (such as an Fc fusion or an alkaline phosphatase fusion) can be used in binding assays to identify a cell type or cell line or tissue that expresses FGF-23 polypeptide receptors. RNA isolated from such cells or tissues can be converted to cDNA, cloned into a mammalian expression vector, and transfected
5 into mammalian cells (such as COS or 293 cells) to create an expression library. A radiolabeled or tagged FGF-23 polypeptide can then be used as an affinity ligand to identify and isolate from this library the subset of cells that express the FGF-23 polypeptide receptors on their surface. DNA can then be isolated from these cells and transfected into mammalian cells to create a secondary expression
10 library in which the fraction of cells expressing FGF-23 polypeptide receptors is many-fold higher than in the original library. This enrichment process can be repeated iteratively until a single recombinant clone containing an FGF-23 polypeptide receptor is isolated. Isolation of the FGF-23 polypeptide receptors is useful for identifying or developing novel agonists and antagonists of the FGF-23
15 polypeptide signaling pathway. Such agonists and antagonists include soluble FGF-23 polypeptide receptors, anti-FGF-23 polypeptide receptor antibodies, small molecules, or antisense oligonucleotides, and they may be used for treating, preventing, or diagnosing one or more of the diseases or disorders described herein.

20 A deposit of cDNA encoding human FGF-23 polypeptide, subcloned into the pGEM-t vector, and having Accession No. PTA-1617, was made with the American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209 on March 31, 2000.

The following examples are intended for illustration purposes only, and
25 should not be construed as limiting the scope of the invention in any way.

Example 1: Cloning of the Human FGF-23 Polypeptide Gene

To isolate cDNA sequences encoding human FGF-23 polypeptide, homology-based BLAST searches of a human genomic database were performed.
30 A putative coding sequence sharing homology with the Fibroblast Growth Factor (FGF) family was identified in a human genomic clone (GenBank accession no.

AC008012). The putative coding sequence consisted of three potential exons separated by introns of 6.6 kb and 1.87 kb. This sequence was used to design gene specific oligonucleotides for the identification of cDNA sources and the generation of cDNA clones, using various PCR strategies.

5 A number of cDNA libraries were analyzed in amplification reactions containing 10 pmol each of the amplimers (5'-C-T-A-T-C-C-C-A-A-T-G-C-C-T-C-C-C-C-A-C-T-G-3'; SEQ ID NO: 42, and 5'-C-G-C-C-C-C-T-G-A-C-C-A-C-C-C-T-A-A-T-G-3'; SEQ ID NO: 43) and Ready-To-Go PCR beads (Pharmacia, Piscataway, NJ), in a total reaction volume of 250 μ l. Reactions were
10 performed at 95°C for 5 minutes for one cycle; 95°C for 30 seconds, 68°C for 15 seconds, and 72°C for 1 minute for 35 cycles; and 72°C for 7 minutes for one cycle. A PCR product of the expected size (616 bp) was identified in a number of cDNA libraries, including libraries derived from colon tumor T25 (random primed), fetal mesentery (oligo-dT primed), fetal gall bladder (random primed),
15 and fetal heart (oligo-dT primed). The PCR product generated from the fetal mesentery cDNA library was subcloned using a TopoTA 4.0 cloning kit (Invitrogen) and four clones were sequenced to verify that the clones contained the predicted FGF-23 cDNA sequence. The fetal mesentery cDNA library was selected for further amplification experiments to isolate full-length cDNA
20 sequences encoding FGF-23 polypeptide.

 The fetal mesentery cDNA library was prepared as follows. Total RNA was extracted from human fetal mesentery using standard RNA extraction procedures and poly-A⁺ RNA was selected from this total RNA using standard procedures. Oligo-dT primed cDNA was synthesized from this poly-A⁺ RNA
25 using the Superscript Plasmid System for cDNA Synthesis and Plasmid Cloning kit (Gibco-BRL), according to the manufacturer's suggested protocols. The resulting cDNA was digested with the restriction endonucleases Sal I and Not I and was then ligated into pSPORT-1. Ligation products were transformed into *E. coli* using standard techniques, and bacterial transformants were selected on
30 culture plates containing ampicillin. The cDNA library consisted of all, or a subset, of these transformants.

Both 5'RACE and 3'RACE reactions were performed in order to generate the full-length cDNA sequence for FGF-23 polypeptide. To isolate cDNA sequences corresponding to the 5' end of the cDNA sequence for FGF-23 polypeptide, 5'RACE was performed using the Smart RACE cDNA Amplification kit (Clontech), random-primed human fetal mesentery cDNA library in pSPORT1, and the primers 5'-G-T-G-T-G-G-A-A-T-T-G-T-G-A-G-C-G-G-A-T-A-A-C-3' (SEQ ID NO: 44) and 5'-C-T-G-A-T-G-G-G-G-T-G-C-G-C-A-T-C-C-A-C-A-3' (SEQ ID NO: 45). Reactions were performed at 94°C for 1 minute for one cycle; 94°C for 5 seconds, 68°C for 10 seconds, and 72°C for 3 minutes for 35 cycles; and 72°C for 7 minutes for one cycle. Nested PCR was performed using a portion of the 5'RACE amplification product (diluted 1/100) and the primers 5'-C-T-A-T-G-A-C-C-A-T-G-A-T-T-A-C-G-C-C-A-A-G-C-3' (SEQ ID NO: 46) and 5'-C-A-T-T-C-T-T-G-T-G-G-A-T-C-T-G-C-A-G-G-T-G-G-T-3' (SEQ ID NO: 47). Nested PCR Reactions were performed at 94°C for 5 minutes for one cycle; 94°C for 15 seconds, 68°C for 15 seconds, and 72°C for 3 minutes for 30 cycles; and 72°C for 7 minutes for one cycle. The amplification products were analyzed by agarose gel electrophoresis and a prominent PCR product of 200 bp was isolated and subcloned using the TopoTA 4.0 cloning kit. Sequencing analysis of isolated clones indicated that the 5' PCR product did not extend the known sequence.

Further 5'RACE experiments were performed to isolate additional cDNA sequences corresponding to the 5' end of the cDNA sequence for FGF-23 polypeptide. Additional 5'RACE experiments were performed using the Advantage-2 PCR kit (Clontech), a Marathon™ human heart cDNA library (Clontech), and the primers 5'-C-T-G-A-T-G-G-G-G-T-G-C-G-C-C-A-T-C-C-A-C-A-3' (SEQ ID NO: 45) and AP1 (Clontech). Reactions were performed at 94°C for 30 seconds for one cycle and 94°C for 5 seconds and 68°C for 4 minutes for 30 cycles. Nested PCR was performed using a portion of the 5'RACE amplification product (diluted 1/100) and the primers 5'-C-A-T-T-C-T-T-G-T-G-G-A-T-C-T-G-C-A-G-G-T-G-G-T-3' (SEQ ID NO: 47) and AP2 (Clontech). Nested PCR Reactions were performed at 94°C for 30 seconds for one cycle and

94°C for 30 seconds, 68°C for 4 minutes for 30 cycles. The amplification products were analyzed by agarose gel electrophoresis and the most prominent PCR product (350 bp) was isolated and subcloned using the TopoTA 4.0 cloning kit. Sequencing analysis of isolated clones indicated that the 5' PCR product
5 extended the known sequence by approximately 143 bp.

To isolate cDNA sequences corresponding to the 3' end of the cDNA sequence for FGF-23 polypeptide, 3'RACE was performed using the Smart RACE cDNA Amplification kit (Clontech), random-primed human fetal mesentery cDNA library in pSPORT1, and the primers 5'-C-G-G-C-C-T-C-C-T-
10 G-T-T-C-A-C-A-G-G-A-G-C-T-C-3' (SEQ ID NO: 48) and 5'-C-G-G-G-C-C-T-C-T-T-C-G-C-T-A-T-T-A-C-G-C-3' (SEQ ID NO: 49). Reactions were performed at 94°C for 1 minute for one cycle; 94°C for 5 seconds, 68°C for 10 seconds, and 72°C for 3 minutes for 35 cycles; and 72°C for 7 minutes for one cycle. Nested PCR was performed using a portion of the 5'RACE amplification
15 product (diluted 1/100) and the primers 5'-G-C-G-C-C-G-A-G-G-A-C-A-A-C-A-G-C-C-C-G-A-3' (SEQ ID NO: 50) and 5'-T-G-G-C-G-A-A-A-G-G-G-G-G-A-T-G-T-G-C-T-G-3' (SEQ ID NO: 51). Nested PCR Reactions were performed at 94°C for 5 minutes for one cycle; 94°C for 15 seconds, 68°C for 15 seconds, and 72°C for 3 minutes for 30 cycles; and 72°C for 7 minutes for one cycle. The
20 amplification products were analyzed by agarose gel electrophoresis and a prominent PCR product of 650 bp was isolated and subcloned using the TopoTA 4.0 cloning kit. Sequencing analysis of isolated clones indicated that the 3' PCR product extended the known sequence by 433 bp, including the poly-A region.

A contiguous sequence that appears to contain the full-length open reading
25 frame for the FGF-23 gene was generated using the sequence derived from the initial PCR amplification and the 5' and 3'RACE amplifications. Sequence analysis of this consensus sequence indicated that the FGF-23 gene comprises a 753 bp open reading frame encoding a protein of 251 amino acids (Figures 1A-1B).

30 Sequence analysis also revealed that FGF-23 polypeptide shares homology with the Fibroblast Growth Factor (FGF) family. Figures 2A-2G illustrate the

amino acid sequence alignment of human FGF-1 (hu FGF-1; SEQ ID NO: 4), human FGF-2 (hu FGF-2; SEQ ID NO: 5), human FGF-3 (hu FGF-3; SEQ ID NO: 6), human FGF-4 (hu FGF-4; SEQ ID NO: 7), human FGF-5 (hu FGF-5; SEQ ID NO: 8), human FGF-6 (hu FGF-6; SEQ ID NO: 9), human FGF-7 (hu FGF-7; SEQ ID NO: 10), human FGF-8 (hu FGF-8; SEQ ID NO: 11), human FGF-9 (hu FGF-9; SEQ ID NO: 12), human FGF-10 (hu FGF-10; SEQ ID NO: 13), human FGF-11 (hu FGF-11; SEQ ID NO: 14), human FGF-12 (hu FGF-12; SEQ ID NO: 15), human FGF-13 (hu FGF-13; SEQ ID NO: 16), human FGF-14 (hu FGF-14; SEQ ID NO: 17), human FGF-16 (hu FGF-16; SEQ ID NO: 18), human FGF-17 (hu FGF-17; SEQ ID NO: 19), human FGF-18 (hu FGF-18; SEQ ID NO: 20), human FGF-19 (hu FGF-19; SEQ ID NO: 21), human FGF-23 (hu FGF-23; SEQ ID NO: 22), murine FGF-1 (mu FGF-1; SEQ ID NO: 23), murine FGF-2 (mu FGF-2; SEQ ID NO: 24), murine FGF-3 (mu FGF-3; SEQ ID NO: 25), murine FGF-4 (mu FGF-4; SEQ ID NO: 26), murine FGF-5 (mu FGF-5; SEQ ID NO: 27), murine FGF-6 (mu FGF-6; SEQ ID NO: 28), murine FGF-7 (mu FGF-7; SEQ ID NO: 29), murine FGF-8 (mu FGF-8; SEQ ID NO: 30), murine FGF-9 (mu FGF-9; SEQ ID NO: 31), murine FGF-10 (mu FGF-10; SEQ ID NO: 32), murine FGF-11 (mu FGF-11; SEQ ID NO: 33), murine FGF-12 (mu FGF-12; SEQ ID NO: 34), murine FGF-13 (mu FGF-13; SEQ ID NO: 35), murine FGF-14 (mu FGF-14; SEQ ID NO: 36), murine FGF-15 (mu FGF-15; SEQ ID NO: 37), rat FGF-16 (rat FGF-16; SEQ ID NO: 38), murine FGF-17 (mu FGF-17; SEQ ID NO: 39).

From the amino acid sequence analysis shown in Figures 2A-2G, the FGF-23 gene appears to be closely related to murine FGF-15 and human FGF-19. The regionally restricted pattern of FGF-15 expression in the developing nervous system suggests that FGF-15 may play an important role in regulating cell division and patterning within specific regions of the embryonic brain, spinal cord, and sensory organs (McWhirter *et al.*, 1997, *Development* 124:3221-32). Accordingly, FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof may be useful for the diagnosis or treatment of diseases involving the developing nervous system. Human FGF-19, which is expressed in

fetal cartilage, skin, and retina, adult gall bladder, and a colon adenocarcinoma cell line, maps to a region of chromosome 11 that is associated with an osteoporosis-pseudoglioma syndrome of skeletal and retinal defects (Xie *et al.*, 1999, *Cytokine* 11:729-35). Accordingly, FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof may be useful for the diagnosis or treatment of diseases involving the skeletal system or retina.

The FGF-23 gene has been shown to be most closely related to human FGF-21 (Yamashita *et al.*, 2000, *Biochem. Biophys. Res. Commun.* 277:494-98), a gene which is expressed most abundantly in the liver and at lower levels in the thymus (Nishimura *et al.*, 2000, *Biochim. Biophys. Acta* 1492:203-06). Accordingly, FGF-23 nucleic acid molecules, polypeptides, and agonists and antagonists thereof may be useful for the diagnosis or treatment of diseases involving the liver or thymus.

Example 2: FGF-23 mRNA Expression

The expression of FGF-23 was analyzed by RT-PCR. Total RNA was prepared from various human fetal tissues using standard techniques. Template and primer mixtures were prepared using 2 µg of total RNA and 50 ng of random primer (Gibco-BRL) in a volume of 12 µl. The mixtures were heated to 70°C for 10 minutes and then chilled on ice. Reverse transcription was performed by adding 4 µl of 5X first strand buffer (Gibco-BRL), 2 µl of 0.1 M DTT, and 1 µl of 10 mM dNTPs to the template-primer mixture, warming the reaction mixture to 37°C for 2 minutes, adding 1 µl of Superscript II RT (Gibco-BRL), and then incubating the reaction mixture at 37°C for 1 hour.

Differences in RNA concentration and cDNA conversion efficiency were normalized by performing control PCR amplifications on each cDNA sample using primers specific for glyceraldehyde-3-phosphate dehydrogenase (G3PDH), a gene expected to be expressed at about the same level in all of the tissues to be examined. Control PCR amplifications were performed using the amplimers 5'-T-C-C-A-C-C-A-C-C-C-T-G-T-T-G-C-T-G-T-A-G-3' (SEQ ID NO: 52) and 5'-G-A-C-C-A-CA-G-T-C-C-A-T-G-C-C-A-T-C-A-C-T-3' (SEQ ID NO: 53) and

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Ready-To-Go PCR Beads (Amersham Pharmacia Biotech, Piscataway, NJ). Reactions were performed at 95°C for 1 minute for one cycle; 92°C for 30 seconds, 55°C for 45 seconds, and 72°C for 1 minute for 25 cycles; and 72°C for 5 minutes for one cycle. Control reaction products were analyzed on 2% agarose gels, the relative intensities of the control products were estimated, and the concentration of cDNA samples was adjusted so that the cDNA samples would generate G3PDH bands of equal intensity. FGF-23 expression analysis was carried out using concentration-normalized cDNA samples.

The expression of FGF-23 was analyzed in PCR amplifications containing the amplimers 5'-C-T-A-T-C-C-CA-A-T-G-C-C-T-C-C-C-A-C-T-G-3' (SEQ ID NO: 54) and 5'-C-G-C-C-C-C-T-G-A-C-C-A-C-C-C-C-T-A-A-T-G-3' (SEQ ID NO: 55) and Ready-To-Go PCR Beads. Reactions were performed at 95°C for 5 minutes for one cycle; 95°C for 30 seconds, 68°C for 30 seconds, and 72°C for 1 minute for 30 cycles; and 72°C for 7 minutes for one cycle. PCR products were analyzed on 2% agarose gels and the relative intensities of the PCR products were estimated (using the faintest PCR product as a baseline). The results of this analysis are shown in Table III.

Table III

Relative FGF-23 Expression

Tissue	Relative Expression Level
Spinal cord	+++
Bladder	+++
Adrenal	+
Bone	+/-
Placenta	++
Intestine	++++
Mesentery	++
Lung	++
Thymus	+/-

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Pancreas	+
Cord Blood	+++
Uterus	+/-
Heart	++
Testes	+++
Eye	-

FGF-23 mRNA expression is analyzed on Northern blots. Multiple human tissue Northern blots (Clontech) are probed with a suitable restriction fragment isolated from a human FGF-23 polypeptide cDNA clone. The probe is labeled
 5 with ^{32}P -dCTP using standard techniques.

Northern blots are prehybridized for 2 hours at 42°C in hybridization solution (5X SSC, 50% deionized formamide, 5X Denhardt's solution, 0.5% SDS, and 100 mg/ml denatured salmon sperm DNA) and then hybridized at 42°C overnight in fresh hybridization solution containing 5 ng/ml of the labeled probe.
 10 Following hybridization, the filters are washed twice for 10 minutes at room temperature in 2X SSC and 0.1% SDS, and then twice for 30 minutes at 65°C in 0.1X SSC and 0.1% SDS. The blots are then exposed to autoradiography.

The expression of FGF-23 mRNA in normal adult mouse tissue and in 3-week-old high expressing and non-expressing transgenic mouse tissue (see
 15 Example 5) was localized by *in situ* hybridization. Normal embryonic and adult mouse tissues were immersion fixed, embedded in paraffin, and sectioned at 5 μm . *In situ* hybridization was performed using standard techniques. Sectioned tissues were hybridized overnight at 60°C in hybridization solution containing a ^{33}P -labeled antisense riboprobe complementary to the human FGF-23 gene. The
 20 riboprobe was obtained by *in vitro* transcription of a clone containing human FGF-23 cDNA sequences using standard techniques.

Following hybridization, sections were treated with RNaseA to digest unhybridized probe and washed in 0.1X SSC at 55°C for 30 minutes. Sections were then immersed in NTB-2 emulsion (Kodak, Rochester, NY), exposed for 3
 25 weeks at 4°C, developed, and counterstained with hematoxylin and eosin. Tissue

morphology and hybridization signal were simultaneously analyzed by darkfield and standard illumination for brain, gastrointestinal system (parotid, submandibular, and sublingual glands; esophagus; stomach; duodenum; jejunum; ileum; proximal and distal colon; liver; and pancreas), cardiopulmonary system
5 (heart, lung, trachea, and blood vessels); hematolymphoid system (lymph nodes, spleen, thymus, and bone marrow), urinary system (kidney and bladder), endocrine system (adrenal gland, thyroid gland, and pituitary gland), reproductive system (testis, prostate, and glands; ovary, uterus, and oviduct; placenta); and musculoskeletal system (bone, skeletal muscle, skin, and adipose tissue). Normal
10 mouse embryos at E18 and E14.5 with placenta were also analyzed by *in situ* hybridization.

A low to moderate, diffuse expression of FGF-23 was detected throughout the tissues of normal adult mouse. An RNAase protection assay was performed on a limited sample of mouse tissues in which a diffuse signal was detected by *in*
15 *situ* hybridization, to determine whether this signal due to non-specific binding. The results of the RNAase protection assay indicated that FGF-23 is expressed only weakly in the heart and brain, and not at all in liver, thyroid/parathyroid, and stomach. These results suggest that the diffuse signal detected by *in situ* hybridization is mostly due to non-specific binding, especially in epithelial cell
20 types. Since the RNAase protection assay indicated that FGF-23 is weakly expressed in the heart and brain, the low signal observed in these tissues by *in situ* hybridization may be real. In the brain, generally low expression was seen in most neurons including areas of the thalamus, caudate putamen, septum, and hypothalamus. Moderate labeling, however, was noted in the hippocampal
25 granule and pyramidal cells, the neocortex (Figure 3), and the piriform cortex. Low expression was found in the ependyma and choroid plexus. Both cardiac and skeletal muscle also exhibited low levels of FGF-23 expression. In cardiac muscle, low diffuse signal was evident throughout the left and right ventricles with a somewhat greater signal in the atrium (Figure 3). A low diffuse signal was
30 also found in skeletal muscle.

No expression was found in either the E14.5 or the E18 mouse embryos.

Neither of the 3-week-old transgenic mouse littermates showed the diffuse non-specific signal that was detected in normal adult mouse. In the non-expressing transgenic mouse, strong FGF-23 expression was detected in scattered cells in the lymph nodes (Figure 4), thymic medulla (Figure 4), and bone (Figure 4). While positive identification of the labeled cells was not possible, the expression in bone seemed to be in the mesenchymal cells scattered in the lacunae and trabeculae in the bones of the hindlimb, vertebrae, ribs, and nasal cavities. In the high expressing transgenic mouse, the distribution of labeled cells was more widespread. In the liver, strong FGF-23 expression was found throughout the hepatocytes (Figure 5). Strong labeling was also detected in scattered cells of the thymic medulla (Figure 5), as was observed in the non-expressing transgenic mouse. However, in the high expressing transgenic mouse, well-labeled cells were also found in the red pulp of the spleen (Figure 5), smooth muscle adjoining the prostate gland (Figure 6), and striated muscle of the jaw (Figure 6). Strong expression was also detected in a few identified megakaryocytes in the bone marrow and in numerous chondrocytes in the hindlimb and in the vertebrae (Figure 6).

Example 3: Production of FGF-23 Polypeptides

A. Expression of FGF-23 Polypeptides in Bacteria

PCR is used to amplify template DNA sequences encoding an FGF-23 polypeptide using primers corresponding to the 5' and 3' ends of the sequence. The amplified DNA products may be modified to contain restriction enzyme sites to allow for insertion into expression vectors. PCR products are gel purified and inserted into expression vectors using standard recombinant DNA methodology. An exemplary vector, such as pAMG21 (ATCC no. 98113) containing the lux promoter and a gene encoding kanamycin resistance is digested with Bam HI and Nde I for directional cloning of inserted DNA. The ligated mixture is transformed into an *E. coli* host strain by electroporation and transformants are selected for kanamycin resistance. Plasmid DNA from selected colonies is isolated and subjected to DNA sequencing to confirm the presence of the insert.

Transformed host cells are incubated in 2xYT medium containing 30 $\mu\text{g/mL}$ kanamycin at 30°C prior to induction. Gene expression is induced by the addition of N-(3-oxohexanoyl)-dl-homoserine lactone to a final concentration of 30 ng/mL followed by incubation at either 30°C or 37°C for six hours. The expression of FGF-23 polypeptide is evaluated by centrifugation of the culture, resuspension and lysis of the bacterial pellets, and analysis of host cell proteins by SDS-polyacrylamide gel electrophoresis.

Inclusion bodies containing FGF-23 polypeptide are purified as follows. Bacterial cells are pelleted by centrifugation and resuspended in water. The cell suspension is lysed by sonication and pelleted by centrifugation at 195,000 $\times g$ for 5 to 10 minutes. The supernatant is discarded, and the pellet is washed and transferred to a homogenizer. The pellet is homogenized in 5 mL of a Percoll solution (75% liquid Percoll and 0.15 M NaCl) until uniformly suspended and then diluted and centrifuged at 21,600 $\times g$ for 30 minutes. Gradient fractions containing the inclusion bodies are recovered and pooled. The isolated inclusion bodies are analyzed by SDS-PAGE.

A single band on an SDS polyacrylamide gel corresponding to *E. coli*-produced FGF-23 polypeptide is excised from the gel, and the N-terminal amino acid sequence is determined essentially as described by Matsudaira *et al.*, 1987, *J. Biol. Chem.* 262:10-35.

B. Expression of FGF-23 Polypeptide in Mammalian Cells

PCR is used to amplify template DNA sequences encoding an FGF-23 polypeptide using primers corresponding to the 5' and 3' ends of the sequence. The amplified DNA products may be modified to contain restriction enzyme sites to allow for insertion into expression vectors. PCR products are gel purified and inserted into expression vectors using standard recombinant DNA methodology. An exemplary expression vector, pCEP4 (Invitrogen, Carlsbad, CA), that contains an Epstein-Barr virus origin of replication, may be used for the expression of FGF-23 polypeptides in 293-EBNA-1 cells. Amplified and gel purified PCR products are ligated into pCEP4 vector and introduced into 293-EBNA cells by

lipofection. The transfected cells are selected in 100 µg/mL hygromycin and the resulting drug-resistant cultures are grown to confluence. The cells are then cultured in serum-free media for 72 hours. The conditioned media is removed and FGF-23 polypeptide expression is analyzed by SDS-PAGE.

5 FGF-23 polypeptide expression may be detected by silver staining. Alternatively, FGF-23 polypeptide is produced as a fusion protein with an epitope tag, such as an IgG constant domain or a FLAG epitope, which may be detected by Western blot analysis using antibodies to the peptide tag.

FGF-23 polypeptides may be excised from an SDS-polyacrylamide gel, or
10 FGF-23 fusion proteins are purified by affinity chromatography to the epitope tag, and subjected to N-terminal amino acid sequence analysis as described herein.

C. Purification of FGF-23 Polypeptide from Mammalian Cells

FGF-23 polypeptide expression constructs are introduced into 293 EBNA
15 or CHO cells using either a lipofection or calcium phosphate protocol.

To conduct functional studies on the FGF-23 polypeptides that are produced, large quantities of conditioned media are generated from a pool of hygromycin selected 293 EBNA clones. The cells are cultured in 500 cm Nunc Triple Flasks to 80% confluence before switching to serum-free media a week
20 prior to harvesting the media. Conditioned media is harvested and frozen at -20°C until the protein is to be purified.

Conditioned media is purified by affinity chromatography as described below. The media is thawed and then passed through a 0.2 µm filter. A Protein G column is equilibrated with PBS at pH 7.0, and then loaded with the filtered
25 media. The column is washed with PBS until the absorbance at A₂₈₀ reaches a baseline. FGF-23 polypeptide is eluted from the column with 0.1 M Glycine-HCl at pH 2.7 and immediately neutralized with 1 M Tris-HCl at pH 8.5. Fractions containing FGF-23 polypeptide are pooled, dialyzed in PBS, and stored at -70°C.

For Factor Xa cleavage of the human FGF-23 polypeptide-Fc fusion
30 polypeptide, affinity chromatography-purified protein is dialyzed in 50 mM Tris-HCl, 100 mM NaCl, 2 mM CaCl₂ at pH 8.0. The restriction protease Factor Xa is

added to the dialyzed protein at 1/100 (w/w) and the sample digested overnight at room temperature.

Example 4: Production of Anti-FGF-23 Polypeptide Antibodies

- 5 Antibodies to FGF-23 polypeptides may be obtained by immunization with purified protein or with FGF-23 peptides produced by biological or chemical synthesis. Suitable procedures for generating antibodies include those described in Hudson and Bay, *Practical Immunology* (2nd ed., Blackwell Scientific Publications).
- 10 In one procedure for the production of antibodies, animals (typically mice or rabbits) are injected with an FGF-23 antigen (such as an FGF-23 polypeptide), and those with sufficient serum titer levels as determined by ELISA are selected for hybridoma production. Spleens of immunized animals are collected and prepared as single cell suspensions from which splenocytes are recovered. The
- 15 splenocytes are fused to mouse myeloma cells (such as Sp2/0-Ag14 cells), are first incubated in DMEM with 200 U/mL penicillin, 200 µg/mL streptomycin sulfate, and 4 mM glutamine, and are then incubated in HAT selection medium (hypoxanthine, aminopterin, and thymidine). After selection, the tissue culture supernatants are taken from each fusion well and tested for anti- FGF-23 antibody
- 20 production by ELISA.

Alternative procedures for obtaining anti-FGF-23 antibodies may also be employed, such as the immunization of transgenic mice harboring human Ig loci for production of human antibodies, and the screening of synthetic antibody libraries, such as those generated by mutagenesis of an antibody variable domain.

25

Example 5: Expression of FGF-23 Polypeptide in Transgenic Mice

- To assess the biological activity of FGF-23 polypeptide, a construct encoding FGF-23 polypeptide under the control of the ApoE promoter (TH00-026) was prepared. The expression of the FGF-23 gene was expected to cause
- 30 pathological changes in the transgenic mice that would be informative as to the function of FGF-23 polypeptide.

A distinctive phenotype was produced in 3-week-old BDF1 mice following transfer of the TH00-026 construct in that litters had an unusually high number of runts. While not all of the expressor mice were runted and some of the non-expressing littermates were runted, the proportion of runts was higher among the expressor mice. All runts, including a number of the non-expressing runts, were taken down before weaning for examination. The skull of all expressing mice was shortened and more rounded, with the lower jaw developing properly. As a result, all expressing mice had protruding lower teeth. This condition is obvious by external examination and radiographic evaluation. In addition, the two highest expressing mice were found to have low serum phosphorous and low serum calcium levels. However, other signs of rickets – such as inadequate mineralization, overgrowth of epiphyseal cartilage, deranged organization of cartilage, and overgrowth of capillaries (*Pathologic Basis of Disease* (Cotran ed., 1994)) – were not observed. Bone morphology in expressing runts was no different than that in non-expressing runts, and both types of runts differed from non-runt littermates.

Partial hepatectomy was performed on 22 DNA positive mice and 4 DNA negative mice. All hepatectomized mice were bled and serum calcium, phosphorous, and alkaline phosphatase measurements were performed. The animals were also examined for the protruding lower teeth phenotype. The groups of mice that were evaluated included: controls, macroscopically phenotypic expressors, non-phenotypic high expressors, and moderate expressors. All phenotypic mice were found to be strong or very strong expressors. Serum calcium levels were somewhat lower in the phenotypic mice than in the other groups, but this difference was not statistically significant. However, serum phosphorous levels were found to be significantly lower in the phenotypic expressors and the non-phenotypic high expressors than in the moderate expressors and the controls. Serum alkaline phosphatase (ALP) was significantly elevated in the phenotypic group versus the controls and moderate expressors (see Table IV). The variability in the phenotype of the expressors may be due to

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genetic variation in the BDF1 mice, an outbred line that is a cross between the C57/B6 and DBA mouse strains.

Table IV

5 Serum Calcium, Phosphorous, and ALP in TH00-026 Transgenic Mice

	Control	Phenotypic expressor	Non- phenotypic high expressor	Moderate expressor	P value
Se Ca (mg/dL)	8.97 ± 0.26	8.64 ± 0.27	8.87 ± 0.32	9.00 ± 0.27	P<0.14
Se P (mg/dL)	6.67 ± 0.52	4.96 ± 1.52	5.22 ± 1.58	7.03 ± 0.58	P<0.0016
Se ALP (IU)	126.5 ± 33	255.4 ± 75	206.4 ± 117	146.3 ± 27	P<0.015

While the present invention has been described in terms of the preferred
embodiments, it is understood that variations and modifications will occur to
10 those skilled in the art. Therefore, it is intended that the appended claims cover
all such equivalent variations that come within the scope of the invention as
claimed.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:
 - 5 (a) the nucleotide sequence as set forth in SEQ ID NO: 1;
 - (b) the nucleotide sequence of the DNA insert in ATCC Deposit No. PTA-1617;
 - (c) a nucleotide sequence encoding the polypeptide as set forth in SEQ ID NO: 2;
 - 10 (d) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (c); and
 - (e) a nucleotide sequence complementary to any of (a) - (c).
2. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:
 - 15 (a) a nucleotide sequence encoding a polypeptide which is at least about 70 percent identical to the polypeptide as set forth in SEQ ID NO: 2, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;
 - 20 (b) a nucleotide sequence encoding an allelic variant or splice variant of the nucleotide sequence as set forth in SEQ ID NO: 1, the nucleotide sequence of the DNA insert in ATCC Deposit No. PTA-1617, or (a);
 - (c) a region of the nucleotide sequence of SEQ ID NO: 1, the DNA insert in ATCC Deposit No. PTA-1617, (a), or (b) encoding a polypeptide
25 fragment of at least about 25 amino acid residues, wherein the polypeptide fragment has an activity of the encoded polypeptide as set forth in SEQ ID NO: 2, or is antigenic;
 - (d) a region of the nucleotide sequence of SEQ ID NO: 1, the DNA insert in ATCC Deposit No. PTA-1617, or any of (a) - (c) comprising a fragment
30 of at least about 16 nucleotides;

(e) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (d); and

(f) a nucleotide sequence complementary to any of (a) - (d).

5 3. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:

(a) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one conservative amino acid substitution, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

10 (b) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid insertion, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(c) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one amino acid deletion, wherein the encoded polypeptide
15 has an activity of the polypeptide set forth in SEQ ID NO: 2;

(d) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 which has a C- and/or N- terminal truncation, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(e) a nucleotide sequence encoding a polypeptide as set forth in SEQ ID NO: 2 with at least one modification selected from the group consisting of amino acid substitutions, amino acid insertions, amino acid deletions, C-terminal truncation, and N-terminal truncation, wherein the encoded polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

(f) a nucleotide sequence of any of (a) - (e) comprising a fragment of
25 at least about 16 nucleotides;

(g) a nucleotide sequence which hybridizes under moderately or highly stringent conditions to the complement of any of (a) - (f); and

(h) a nucleotide sequence complementary to any of (a) - (e).

30 4. A vector comprising the nucleic acid molecule of any of Claims 1, 2, or 3.

5. A host cell comprising the vector of Claim 4.
6. The host cell of Claim 5 that is a eukaryotic cell.
- 5 7. The host cell of Claim 5 that is a prokaryotic cell.
8. A process of producing an FGF-23 polypeptide comprising culturing the host cell of Claim 5 under suitable conditions to express the
10 polypeptide, and optionally isolating the polypeptide from the culture.
9. A polypeptide produced by the process of Claim 8.
10. The process of Claim 8, wherein the nucleic acid molecule
15 comprises promoter DNA other than the promoter DNA for the native FGF-23 polypeptide operatively linked to the DNA encoding the FGF-23 polypeptide.
11. The isolated nucleic acid molecule according to Claim 2, wherein the percent identity is determined using a computer program selected from the
20 group consisting of GAP, BLASTN, FASTA, BLASTA, BLASTX, BestFit, and the Smith-Waterman algorithm.
12. A process for determining whether a compound inhibits FGF-23 polypeptide activity or FGF-23 polypeptide production comprising exposing a
25 cell according to any of Claims 5, 6, or 7 to the compound and measuring FGF-23 polypeptide activity or FGF-23 polypeptide production in said cell.
13. An isolated polypeptide comprising the amino acid sequence selected from the group consisting of:
30 (a) the amino acid sequence as set forth in SEQ ID NO: 2; and

(b) the amino acid sequence encoded by the DNA insert in ATCC Deposit No. PTA-1617.

14. An isolated polypeptide comprising the amino acid sequence
5 selected from the group consisting of:

- (a) the amino acid sequence as set forth in SEQ ID NO: 3, optionally further comprising an amino-terminal methionine;
- (b) an amino acid sequence for an ortholog of SEQ ID NO: 2;
- (c) an amino acid sequence which is at least about 70 percent identical
10 to the amino acid sequence of SEQ ID NO: 2, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;
- (d) a fragment of the amino acid sequence set forth in SEQ ID NO: 2 comprising at least about 25 amino acid residues, wherein the fragment has an activity of the polypeptide set forth in SEQ ID NO: 2, or is antigenic; and
- 15 (e) an amino acid sequence for an allelic variant or splice variant of the amino acid sequence as set forth in SEQ ID NO: 2, the amino acid sequence encoded by the DNA insert in ATCC Deposit No. PTA-1617, or (a) - (c).

15. An isolated polypeptide comprising the amino acid sequence
20 selected from the group consisting of:

- (a) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one conservative amino acid substitution, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;
- (b) the amino acid sequence as set forth in SEQ ID NO: 2 with at least
25 one amino acid insertion, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;
- (c) the amino acid sequence as set forth in SEQ ID NO: 2 with at least one amino acid deletion, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2;

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(d) the amino acid sequence as set forth in SEQ ID NO: 2 which has a C- and/or N- terminal truncation, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2; and

(e) the amino acid sequence as set forth in SEQ ID NO: 2 with at least
5 one modification selected from the group consisting of amino acid substitutions, amino acid insertions, amino acid deletions, C-terminal truncation, and N-terminal truncation, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2.

10 16. An isolated polypeptide encoded by the nucleic acid molecule of any of Claims 1, 2, or 3, wherein the polypeptide has an activity of the polypeptide set forth in SEQ ID NO: 2.

15 17. The isolated polypeptide according to Claim 14, wherein the percent identity is determined using a computer program selected from the group consisting of GAP, BLASTP, FASTA, BLASTA, BLASTX, BestFit, and the Smith-Waterman algorithm.

20 18. A selective binding agent or fragment thereof which specifically binds the polypeptide of any of Claims 13, 14, or 15.

25 19. The selective binding agent or fragment thereof of Claim 18 that specifically binds the polypeptide comprising the amino acid sequence as set forth in SEQ ID NO: 2, or a fragment thereof.

20. The selective binding agent of Claim 18 that is an antibody or fragment thereof.

30 21. The selective binding agent of Claim 18 that is a humanized antibody.

22. The selective binding agent of Claim 18 that is a human antibody or fragment thereof.

23. The selective binding agent of Claim 18 that is a polyclonal antibody or fragment thereof.

24. The selective binding agent Claim 18 that is a monoclonal antibody or fragment thereof.

25. The selective binding agent of Claim 18 that is a chimeric antibody or fragment thereof.

26. The selective binding agent of Claim 18 that is a CDR-grafted antibody or fragment thereof.

27. The selective binding agent of Claim 18 that is an antiidiotypic antibody or fragment thereof.

28. The selective binding agent of Claim 18 that is a variable region fragment.

29. The variable region fragment of Claim 28 that is a Fab or a Fab' fragment.

30. A selective binding agent or fragment thereof comprising at least one complementarity determining region with specificity for a polypeptide having the amino acid sequence of SEQ ID NO: 2.

31. The selective binding agent of Claim 18 that is bound to a detectable label.

32. The selective binding agent of Claim 18 that antagonizes FGF-23 polypeptide biological activity.

33. A method for treating, preventing, or ameliorating an FGF-23 polypeptide-related disease, condition, or disorder comprising administering to a patient an effective amount of a selective binding agent according to Claim 18.

34. A selective binding agent produced by immunizing an animal with a polypeptide comprising an amino acid sequence of SEQ ID NO: 2.

10

35. A hybridoma which produces a selective binding agent which is capable of binding a polypeptide according to any of Claims 1, 2, or 3.

36. A method of detecting or quantitating the amount of FGF-23 polypeptide using the anti-FGF-23 antibody or fragment of Claim 18.

15

37. A composition comprising the polypeptide of any of Claims 13, 14, or 15, and a pharmaceutically acceptable formulation agent.

38. The composition of Claim 37, wherein the pharmaceutically acceptable formulation agent is a carrier, adjuvant, solubilizer, stabilizer, or anti-oxidant.

20

39. The composition of Claim 37, wherein the polypeptide comprises the amino acid sequence as set forth in SEQ ID NO: 3.

25

40. A polypeptide comprising a derivative of the polypeptide of any of Claims 13, 14, or 15.

41. The polypeptide of Claim 40 that is covalently modified with a water-soluble polymer.

30

42. The polypeptide of Claim 41, wherein the water-soluble polymer is selected from the group consisting of polyethylene glycol, monomethoxy-polyethylene glycol, dextran, cellulose, poly-(N-vinyl pyrrolidone) polyethylene glycol, propylene glycol homopolymers, polypropylene oxide/ethylene oxide co-polymers, polyoxyethylated polyols, and polyvinyl alcohol.

43. A composition comprising a nucleic acid molecule of any of Claims 1, 2, or 3 and a pharmaceutically acceptable formulation agent.

10

44. The composition of Claim 43, wherein said nucleic acid molecule is contained in a viral vector.

45. A viral vector comprising a nucleic acid molecule of any of Claims 1, 2, or 3.

15

46. A fusion polypeptide comprising the polypeptide of any of Claims 13, 14, or 15 fused to a heterologous amino acid sequence.

47. The fusion polypeptide of Claim 46, wherein the heterologous amino acid sequence is an IgG constant domain or fragment thereof.

20

48. A method for treating, preventing, or ameliorating a medical condition comprising administering to a patient the polypeptide of any of Claims 13, 14, or 15, or the polypeptide encoded by the nucleic acid of any of Claims 1, 2, or 3.

25

49. A method for treating, preventing or ameliorating a medical condition comprising administering to a patient an agonist or antagonist of the biological activity of the polypeptide of any of Claims 13, 14, or 15 or of the polypeptide encoded by the nucleic acid of any of Claims 1, 2, or 3.

30

50. The method of either Claim 48 or 49 wherein the medical condition being treated, prevented, or ameliorated is autosomal dominant hypophosphatemic rickets (ADHR).

51. A method of diagnosing a pathological condition or a susceptibility to a pathological condition in a subject comprising:

(a) determining the presence or amount of expression of the polypeptide of any of Claims 13, 14, or 15, or the polypeptide encoded by the nucleic acid molecule of any of Claims 1, 2, or 3 in a sample; and

(b) diagnosing a pathological condition or a susceptibility to a pathological condition based on the presence or amount of expression of the polypeptide.

15

52. A device, comprising:

(a) a membrane suitable for implantation; and

(b) cells encapsulated within said membrane, wherein said cells secrete a protein of any of Claims 13, 14, or 15; and

said membrane is permeable to said protein and impermeable to materials detrimental to said cells.

53. A method of identifying a compound which binds to an FGF-23 polypeptide comprising:

(a) contacting the polypeptide of any of Claims 13, 14, or 15 with a compound; and

(b) determining the extent of binding of the FGF-23 polypeptide to the compound.

54. The method of Claim 53, further comprising determining the activity of the polypeptide when bound to the compound.

55. A method of modulating levels of a polypeptide in an animal comprising administering to the animal the nucleic acid molecule of any of Claims 1, 2, or 3.

5

56. A transgenic non-human mammal comprising the nucleic acid molecule of any of Claims 1, 2, or 3.

57. A process for determining whether a compound inhibits FGF-23 polypeptide activity or FGF-23 polypeptide production comprising exposing a transgenic mammal according to Claim 56 to the compound, and measuring FGF-23 polypeptide activity or FGF-23 polypeptide production in said mammal.

10

Figure 1A

atg ttg ggg gcc cgc ctc agg ctc tgg gtc tgt gcc ttg tgc agc gtc	48
<u>Met Leu Gly Ala Arg Leu Arg Leu Trp Val Cys Ala Leu Cys Ser Val</u>	
1 5 10 15	
tgc agc atg agc gtc ctc aga gcc tat ccc aat gcc tcc cca ctg ctc	96
<u>Cys Ser Met Ser Val Leu Arg Ala Tyr Pro Asn Ala Ser Pro Leu Leu</u>	
20 25 30	
ggc tcc agc tgg ggt ggc ctg atc cac ctg tac aca gcc aca gcc agg	144
Gly Ser Ser Trp Gly Gly Leu Ile His Leu Tyr Thr Ala Thr Ala Arg	
35 40 45	
aac agc tac cac ctg cag atc cac aag aat ggc cat gtg gat ggc gca	192
<u>Asn Ser Tyr His Leu Gln Ile His Lys Asn Gly His Val Asp Gly Ala</u>	
50 55 60	
ccc cat cag acc atc tac agt gcc ctg atg atc aga tca gag gat gct	240
<u>Pro His Gln Thr Ile Tyr Ser Ala Leu Met Ile Arg Ser Glu Asp Ala</u>	
65 70 75 80	
ggc ttt gtg gtg att aca ggt gtg atg agc aga aga tac ctc tgc atg	288
Gly Phe Val Val Ile Thr Gly Val Met Ser Arg Arg Tyr Leu Cys Met	
85 90 95	
gat ttc aga ggc aac att ttt gga tca cac tat ttc gac ccg gag aac	336
<u>Asp Phe Arg Gly Asn Ile Phe Gly Ser His Tyr Phe Asp Pro Glu Asn</u>	
100 105 110	
tgc agg ttc caa cac cag acg ctg gaa aac ggg tac gac gtc tac cac	384
<u>Cys Arg Phe Gln His Gln Thr Leu Glu Asn Gly Tyr Asp Val Tyr His</u>	
115 120 125	
tct cct cag tat cac ttc ctg gtc agt ctg ggc cgg gcg aag aga gcc	432
<u>Ser Pro Gln Tyr His Phe Leu Val Ser Leu Gly Arg Ala Lys Arg Ala</u>	
130 135 140	
ttc ctg cca ggc atg aac cca ccc ccg tac tcc cag ttc ctg tcc cgg	480
<u>Phe Leu Pro Gly Met Asn Pro Pro Pro Tyr Ser Gln Phe Leu Ser Arg</u>	
145 150 155 160	
agg aac gag atc ccc cta att cac ttc aac acc ccc ata cca cgg cgg	528
<u>Arg Asn Glu Ile Pro Leu Ile His Phe Asn Thr Pro Ile Pro Arg Arg</u>	
165 170 175	
cac acc cgg agc gcc gag gac gac tgg gag cgg gac ccc ctg aac gtg	576
<u>His Thr Arg Ser Ala Glu Asp Asp Ser Glu Arg Asp Pro Leu Asn Val</u>	
180 185 190	
ctg aag ccc cgg gcc cgg atg acc ccg gcc ccg gcc tcc tgt tca cag	624
<u>Leu Lys Pro Arg Ala Arg Met Thr Pro Ala Pro Ala Ser Cys Ser Gln</u>	
195 200 205	

Figure 1B

gag	ctc	ccg	agc	gcc	gag	gac	aac	agc	ccg	atg	gcc	agt	gac	cca	tta	672
Glu	Leu	Pro	Ser	Ala	Glu	Asp	Asn	Ser	Pro	Met	Ala	Ser	Asp	Pro	Leu	
210						215					220					
ggg	gtg	gtc	agg	ggc	ggt	cga	gtg	aac	acg	cac	gct	ggg	gga	acg	ggc	720
Gly	Val	Val	Arg	Gly	Gly	Arg	Val	Asn	Thr	His	Ala	Gly	Gly	Thr	Gly	
225					230				235					240		
ccg	gaa	ggc	tgc	cgc	ccc	ttc	gcc	aag	ttc	atc						753
Pro	Glu	Gly	Cys	Arg	Pro	Phe	Ala	Lys	Phe	Ile						
			245						250							

FIG. 2A

hu	FGF-1	MA...E....							.GE
hu	FGF-2	MA...A....							.GS
hu	FGF-3	M...G....	..LIWLL..					..LL SLL
hu	FGF-4	MS.....		.GPGTAVAL	LPAVL...	LA LLAPWAGRG			
hu	FGF-5	MSLSFL....	..LLLFFSHL	ILSAWAHGEK	R.....	..LAPKGQPG			
hu	FGF-6	MALGQK....	..LFITMSR.	.GAGRLOQT	WALVF...	LG ILVGMVVPSP			
hu	FGF-7	MH...K....	...WILTWI	LPITLYRS..	...CF...HI	IC.....L			
hu	FGF-8	MGSPRSALSC	LLLHLVLCL	QAQ...EGPG	RGPALGRELA	SL.....			
hu	FGF-9	MA...P....	..LGEVGNYP	GVOQD...		AV PF.....GN			
hu	FGF-10	MW...K....	...WILTHC	ASAFPHLPGC	CCCCF...	LL LP.....L			
hu	FGF-11	MAA.....	...LASSL	IRQKREVRE.		...PGGSRP			
hu	FGF-12	MAAA.....	...IASSL	IRQKRQARE.		...SNSDRV			
hu	FGF-13	MAAA.....	...IASSL	IRQKRQARE.		...R...EKS			
hu	FGF-14	MAAA.....	...IASGL	IRQKRQARE.		...QHWRDP			
hu	FGF-16	MA.....	...EVGGVF	ASLDWDLHG.	...F...SS	SL.....GN			
hu	FGF-17	MGAAR....	..LLPNLTLC	QLLI.....		LC COTQGEN...			
hu	FGF-18	MYSAPSACTC	LCLHPILLCF	QVQVLVA...		...BENV			
hu	FGF-19	M...RS...GC	VVVHVVI...	LAGWLVA...	...VAGRPLA				
hu	FGF-23	MSVLRA....							
mu	FGF-1	MA...E....							.GE
mu	FGF-2	MA...A....							.SG
mu	FGF-3	M...G....	..LIWLL..			..LL SL		L
mu	FGF-4	MA.....		.KRGPTTGTL	LPRVL...	LA LVVALADRGT			
mu	FGF-5	MSLSLL....	..FLIPCSHL	IHSANAHGEK	R.....	..LTPEGQPAP			
mu	FGF-6	MALGQR....	..LFITMSR.	.GAGRVOQT	QALVF...	LG VLVGMVVPSP			
mu	FGF-7	MR...K....	...WILTRI	LPITLYRS..	...CF...HL	VC.....L			
mu	FGF-8	MGSPRSALSC	LLLHLVLCL	QAQVRSAAQK	RGPGAGNPAD	TGQGGHEDRP			
mu	FGF-9	MA...P....	..LGEVGSYF	GVOQD...		AV PF.....GN			
mu	FGF-10	MW...K....	...WILTHC	ASAFPHLPGC	CCC.F...LL	LP.....L			
mu	FGF-11	MAA.....	...LASSL	IRQKREVRE.		...PGGSRP			
mu	FGF-12	MAAA.....	...IASSL	IRQKRQARE.		...SNSDRV			
mu	FGF-13	MTAA.....	...IASSL	IRQKRQARE.		...R...EKS			
mu	FGF-14	MAAA.....	...IASGL	IRQKRQARE.		...QHWRDP			
mu	FGF-15	MARKWN...GR	AVARALV...	LATLWLA...	...VSGRPLA				
rat	FGF-16	MA.....	...EVGGVF	ASLDWDLQG.	...F...SS	SL.....GN			
mu	FGF-17	MYSAPSACTC	LCLHPILLCF	QVQVLAA...		...BENV			

FIG. 2B

hu FGF-1	ITTTFA....LT.EK	FN...LPPGN	YKKP.....
hu FGF-2	ITTLPA....LP.ED	GGSGAFPPGH	FKDP.....
hu FGF-3	EPGWPAA...GPGAR	LR.RDAGGRG	G.VYEHL...
hu FGF-4	AAAPTAPNGT	LEAELER.RW	ESLVALSLAR	LPVAAQPKKA	AVQSGAGDYL
hu FGF-5	AATDRNPIGS	S.SRQSS.SS	AMSSSSASSS	PAASLGSQGS	GLEQSSSQWS
hu FGF-6	AGTR..ANNT	LLD..SR.GW	GTLLSRSRAG	LAGEI....A	GVNWESG.YL
hu FGF-7	VGTTISLACND	MTPEQMA.TN	V.NCSSPE..RH	TRSYDYME..
hu FGF-8	F....RAGREPQ	G.....	...VSQQHVR	EQSLVTDQLS
hu FGF-9	VPVLPVDSPV	LLSDHLG.QS	EA.....	..GGLPRGPA	VTDLHLK..
hu FGF-10	VSSVPVTCQA	LGQDMVS.PE	ATN..SSSS	FSSPSSAGRH	VRSYNHLQ..
hu FGF-11	VSAQRRVCP.	RGTKSLC.QK	QLLILLSKVR	LCGGRPARPD	RGPEPQLK..
hu FGF-12	SASKRRSSPS	KDGRSLC.ER	HVLGVFSKVR	FCSGRKRPVR	RRPEPQLK..
hu FGF-13	NACKCVSSPS	K.GKTSC.DK	NKLNVSFRVK	LFGSKKRR..R	RRPEPQLK..
hu FGF-14	SASRRRSSPS	KN.RGLC.NG	NLVDIFSKVR	IFGLKKRRLR	RQ.DPQLK..
hu FGF-16	VPL..ADSPG	FLNERLG.QI	E.....	..GKLQRGSP	.TDFAHLK..
hu FGF-17HPSPNF..NQYVR	DQGAMTDQLS
hu FGF-18	F.....RIHVE	NQTRARDDVS
hu FGF-19FS	DAGPHVHYGW
hu FGF-23YP	NASPLLGSW
mu FGF-1	ITTFAA....LT.ER	FN...LPLGN	YKKP.....
mu FGF-2	ITSLPA....LP.ED	GGA.AFPPGH	FKDP.....
mu FGF-3	EPSWPTT...GPGTR	LR.RDAGGRG	G.VYEHL...
mu FGF-4	AA....PNGT	RHAELGH.GW	DGLVARSLAR	LPVAAQPPQA	AVRSGAGDYL
mu FGF-5	P...RNPGRS	SGSRGRS.SA	TFSSSSASSP	VAASPGSQGS	GSEHSSSQWS
mu FGF-6	AGAR..ANGT	LLD..SR.GW	GTLLSRSRAG	LAGEI....S	GVNWESG.YL
mu FGF-7	VGTTISLACND	MSPEQTA.TS	V.NCSSPE..RH	TRSYDYME..
mu FGF-8	FGQSRAGKN	FTNPAPNYPE	EGSKEQRDSV	LPKVTQRHVR	EQSLVTDQLS
mu FGF-9	VPVLPVDSPV	LLNDHLG.QS	EA.....	..GGLPRGPA	VTDLHLK..
mu FGF-10	VSSFPVTCQA	LGQDMVS.QE	ATNCSSSSSS	FSSPSSAGRH	VRSYNHLQ..
mu FGF-11	VSAQRRVCP.	RGTKSLC.QK	QLLILLSKVR	LCGGRPTRQD	RGPEPQLK..
mu FGF-12	SASKRRSSPS	KDGRSLC.ER	HVLGVFSKVR	FCSGRKRPVR	RRPEPQLK..
mu FGF-13	NACKCVSSPS	K.GKTSC.DK	NKLNVSFRVK	LFGSKKRR..R	RRPEPQLK..
mu FGF-14	SASRRRSSPS	KN.RGLF.NG	NLVDIFSKVR	IFGLKKRRLR	RQ.DPQLK..
mu FGF-15	...QQSQSVS	DEDPLFLYGW
rat FGF-16	VPL..ADSPG	FLNERLG.QI	E.....	..GKLQRGSP	.TDFAHLK..
mu FGF-17	F.....RIHVE	NQTRARDDVS

FIG. 2C

hu FGF-1KLLY	CSNG.GH...	FLRILPDGTV	DGTRDRSDQH	IQLQLSAESV
hu FGF-2KRLY	CKNG.GF...	FLRIHPDGRV	DGVREKSDPH	IKLQLQAEER
hu FGF-3	GGAPRRRKLY	CATK..Y...	HLQLHPSGRV	NGSL.ENSAY	SILEITAVEV
hu FGF-4	LGIKRLRRLY	CNVGIGF...	HLQALPDGRI	GGAHADT.RD	SLELSPVER
hu FGF-5	PSGRRTGSLY	CRVGIGF...	HLQIYPDGKV	NGSHEAN.ML	SVLEIFAVSQ
hu FGF-6	VGIKRQRRLY	CNVGIGF...	HLQVLPDGRI	SGTHEEN.PY	SLEISTVER
hu FGF-7	GGDIRVRLRF	CRTQ..W...	YLRIDKRGKV	KGTQEMKNNY	NIMEIRTVAV
hu FGF-8	RRLIRTYQLY	SRTS.GK...	HVQVLANKRI	NAMAEDGDPF	AKLIVETDTF
hu FGF-9	G.ILRRRQLY	CRTG..F...	HLEIFPNGTI	QGTRKDHSRF	GILEFISIAV
hu FGF-10	G.DVRWRKLF	SPTK..Y...	FLKIEKNGKV	SGTKKENCYP	SLEITSVEI
hu FGF-11	.GIV..TKLF	CRQ..GF...	YLQANPDGSI	QGTPEDTSSF	THFNLI PVGL
hu FGF-12	.GIV..TRLF	SQ..GY...	FLQMHPDGTI	DGTDKEDSDY	TLFNLI PVGL
hu FGF-13	.GIV..TKLY	SRQ..GY...	HLQLQADGTI	DGTDKEDSTY	TLFNLI PVGL
hu FGF-14	.GIV..TRLY	CRQ..GY...	YLQMHPDGAL	DGTDKDDSTNS	TLFNLI PVGL
hu FGF-16	G.ILRRRQLY	CRTG..F...	HLEIFPNGTV	HGTRHDHSRF	GILEFISLAV
hu FGF-17	RRQIREYQLY	SRTS.GK...	HVQV.TGRR	SATAEDGNKF	AKLIVETDTF
hu FGF-18	RKQLRLYQLY	SRTS.GK...	HIQVL.GRRI	SARGEDGDY	AQLLVETDTF
hu FGF-19	GDPIRLRLY	TSGPHGLSSC	FLRIRADGVV	DCARGQSA.H	SLEIKAV
hu FGF-23	GGLIH...LY	TATARN...SY	HLQIHKNHGV	DGAPHOTI.Y	SALMIRSEDA
mu FGF-1KLLY	CSNG.GH...	FLRILPDGTV	DGTRDRSDQH	IQLQLSAESA
mu FGF-2KRLY	CKNG.GF...	FLRIHPDGRV	DGVREKSDPH	VKLQLQAEER
mu FGF-3	GGAPRRRKLY	CATK..Y...	HLQLHPSGRV	NGSL.ENSAY	SILEITAVEV
mu FGF-4	LGLKRLRRLY	CNVGIGF...	HLQVLPDGRI	GGVHADT.RD	SLELSPVQR
mu FGF-5	PSGRRTGSLY	CRVGIGF...	HLQIYPDGKV	NGSHEAS.VL	SILEIFAVSQ
mu FGF-6	VGIKRQRRLY	CNVGIGF...	HLQVPPDGRI	SGTHEEN.PY	SLEISTVER
mu FGF-7	GGDIRVRLRF	CRTQ..W...	YLRIDKRGKV	KGTQEMKNSY	NIMEIRTVAV
mu FGF-8	RRLIRTYQLY	SRTS.GK...	HVQVLANKRI	NAMAEDGDPF	AKLIVETDTF
mu FGF-9	G.ILRRRQLY	CRTG..F...	HLEIFPNGTI	QGTRKDHSRF	GILEFISIAV
mu FGF-10	G.DVRWRRLRF	SPTK..Y...	FLTIEKNGKV	SGTKNEDCPY	SVLEITSVEI
mu FGF-11	.GIV..TKLF	CRQ..GF...	YLQANPDGSI	QGTPEDTSSF	THFNLI PVGL
mu FGF-12	.GIV..TRLF	SQ..GY...	FLQMHPDGTI	DGTDKEDSDY	TLFNLI PVGL
mu FGF-13	.GIV..TKLY	SRQ..GY...	HLQLQADGTI	DGTDKEDSTY	TLFNLI PVGL
mu FGF-14	.GIV..TRLY	CRQ..GY...	YLQMHPDGAL	DGTDKDDSTNS	TLFNLI PVGL
mu FGF-15	GKITRLQYLY	SAGPY.VSNC	FLRIRSDGSV	DCEEDQNE.R	NLEFRAVAL
rat FGF-16	G.ILRRRQLY	CRTG..F...	HLEIFPNGTV	HGTRHDHSRF	GILEFISLAV
mu FGF-17	RKQLRLYQLY	SRTS.GK...	HIQVL.GRRI	SARGEDGDY	AQLLVETDTF

FIG. 2D

hu FGF-1	GE.VYIKSTE	TGQYLAMDTD	GLLYGSQTPN	E.ECLFLERL	EENHYNTYIS
hu FGF-2	GV.VSIKVC	ANRYLAMKED	GRLLASKCVT	D.ECFFFERL	ESNNYNTYRS
hu FGF-3	GI.VAIRGLF	SGRYLAMNKR	GRLYASEHYS	A.ECEFVERI	HELGYNTYAS
hu FGF-4	GV.VSIFGVA	SRPFVAMSSK	GKLYGSPFFT	D.ECTFKEIL	LPNNYNAYES
hu FGF-5	GI.VGIRGVF	SNKFLAMSKK	GKLHASAKFT	D.DCKFRERF	QENSYNTYAS
hu FGF-6	GV.VSLFGVR	SALFVAMNSK	GRLYATPSFQ	E.ECKFRETL	LPNNYNAYES
hu FGF-7	GI.VAIKGVF	SEFYLAMNKE	GKLYAKKECN	E.DCNFKELI	LENHYNTYAS
hu FGF-8	GSRVRVRGAE	TGLYICMNKK	GKLIASNGK	GKDCVFTEIV	LENNYTALQN
hu FGF-9	GL.VSIRGVD	SGLYLGMNEK	GELYGSEKLT	Q.ECVFREOF	EENWYNTYSS
hu FGF-10	GV.VAVKAIN	SNYYLAMNKK	GKLYGSKEFN	N.DCKLKERI	EENGYNTYAS
hu FGF-11	RV.VTIQSAK	LGHYAMNAB	GLLYSSPHFT	A.ECRFKECV	FENYYVLYAS
hu FGF-12	RV.VAIQGVK	ASLYVAMNGE	GYLYSSDVFT	P.ECKFKESV	FENYYVIYSS
hu FGF-13	RV.VAIQGVQ	TKLYLAMNSE	GYLYTSELFT	P.ECKFKESV	FENYYVTYSS
hu FGF-14	RV.VAIQGVK	TGLYIAMNGE	GYLYPSELFT	P.ECKFKESV	FENYYVIYSS
hu FGF-16	GL.ISIRGVD	SGLYLGMNER	GELYGSKKLT	R.ECVFREOF	EENWYNTYAS
hu FGF-17	GSRVRIKGAE	SBKYICMNKR	GKLIGKPSGK	SKDCVFTEIV	LENNYTAFQN
hu FGF-18	GSQVRIKGE	TEFYLCMNRK	GKLVGKPDGT	SKECVFIEKV	LENNYTALMS
<u>hu FGF-19</u>	<u>RT.VAIKGVH</u>	<u>SVRYLCMGAD</u>	<u>GKMOGLLOYS</u>	<u>EEDCAFREEI</u>	<u>RPDGYNVYRS</u>
<u>hu FGF-23</u>	<u>GF.VVITGVM</u>	<u>SRRYLCMDPR</u>	<u>GNI FGSHYFD</u>	<u>PENCRFOHOT</u>	<u>LENGYDVYHS</u>
mu FGF-1	GE.VYIKGTE	TGQYLAMDTE	GLLYGSQTPN	E.ECLFLERL	EENHYNTYTS
mu FGF-2	GV.VSIKVC	ANRYLAMKED	GRLLASKCVT	E.ECFFFERL	ESNNYNTYRS
mu FGF-3	GV.VAIKGLF	SGRYLAMNKR	GRLYASDHYN	A.ECEFVERI	HELGYNTYAS
mu FGF-4	GV.VSIFGVA	SRPFVAMSSR	GKLFGVPFFT	D.ECKFKEIL	LPNNYNAYEA
mu FGF-5	GI.VGIRGVF	SNKFLAMSKK	GKLHASAKFT	D.DCKFRERF	QENSYNTYAS
mu FGF-6	GV.VSLFGVK	SALFIAMNSK	GRLYTTPSPH	D.ECKFRETL	LPNNYNAYES
mu FGF-7	GI.VAIKGVF	SEYYLAMNKE	GKLYAKKECN	E.DCNFKELI	LENHYNTYAS
mu FGF-8	GSRVRVRGAE	TGLYICMNKK	GKLIASNGK	GKDCVFTEIV	LENNYTALQN
mu FGF-9	GL.VSIRGVD	SGLYLGMNEK	GELYGSEKLT	Q.ECVFREOF	EENWYNTYSS
mu FGF-10	GV.VAVKAIN	SNYYLAMNKK	GKLYGSKEFN	N.DCKLKERI	EENGYNTYAS
mu FGF-11	RV.VTIQSAK	LGHYAMNAB	GLLYSSPHFT	A.ECRFKECV	FENYYVLYAS
mu FGF-12	RV.VAIQGVK	ASLYVAMNGE	GYLYSSDVFT	P.ECKFKESV	FENYYVIYSS
mu FGF-13	RV.VAIQGVQ	TKLYLAMNSE	GYLYTSEHFT	P.ECKFKESV	FENYYVTYSS
mu FGF-14	RV.VAIQGVK	TGLYIAMNGE	GYLYPSELFT	P.ECKFKESV	FENYYVIYSS
mu FGF-15	KT.IAIKDV	SVRYLCMSAD	GKIYGLIRYS	EEDCTFREEM	DCLGYNQYRS
rat FGF-16	GL.ISIRGVD	SGLYLGMNER	GELFGSKKLT	R.ECVFREOF	EENWYNTYAS
mu FGF-17	GSQVRIKGE	TEFYLCMNRK	GKLVGKPDGT	SKECVFIEKV	LENNYTALMS

FIG. 2E

hu FGF-1	KKH.....A	EKNWFVGLKK	NGSCKRG..P	RTHYGQKAIL
hu FGF-2	RKY.....T	..SWYVALKR	TGQYKLG..S	KTGPGQKAIL
hu FGF-3	RLYRTVSSTP	GARRQ..PSA	ERLWYVSVNG	KGRPRRG..F	KTRRTQKSSL
hu FGF-4	YKYPGM....FIALSK	NGTKKKG..N	RVSPTMKVTH
hu FGF-5	AIHRTEKTG.REWYVALNK	RGKAKRGCS	P RVKPDHISTH
hu FGF-6	DLYQGT....YIALSK	YGRVKRG..S	KVSPIMTVTH
hu FGF-7	A.....	..KWT..HNG	GEM.FVALNQ	KGIPVRG..K	KTKEQKTAH
hu FGF-8	AKYEG.....WYMAFTR	KGRPRKG..S	KTRQHOREVH
hu FGF-9	NLYK.....H..VDT	GRRYYVALNK	DGTPREG..T	RTKRHQKPTH
hu FGF-10	F.....	..NWQ..HNG	RQM.YVALNG	KGAPRRG..Q	KTRRKNTSAH
hu FGF-11	ALYR.....Q..RRS	GRAWYLGDK	EGQVMKG..N	RVKKTAAAH
hu FGF-12	TLYR.....Q..QES	GRAWFLGLNK	EGQIMKG..N	RVKKTKPSSH
hu FGF-13	MIYR.....Q..QES	GRGWYLGDK	EGEIMKG..N	HVKKNKPAAH
hu FGF-14	MLYR.....Q..QES	GRAWFLGLNK	EGQAMKG..N	RVKKTKPAAH
hu FGF-16	TLYK.....H..SDS	ERQYYVALNK	DGSPREG..Y	RTKRHQKPTH
hu FGF-17	ARHEG.....WFMAFTR	QGRPRQA..S	RSRQONREAH
hu FGF-18	AKYSG.....WYVGFTK	KGRPRKG..P	KTRENQDQVH
hu FGF-19	EKHRLPVSL	SAKO.....RQLYK	NRGFLPLSHF	...LPMLPMV
hu FGF-23	POYHFLVSLG	RAKRAFLPGM	NPPPYSOFLS	RRNEIPLIHF	NTPIPRRHTR
mu FGF-1	KKH.....A	EKNWFVGLKK	NGSCKRG..P	RTHYGQKAIL
mu FGF-2	RKY.....S	..SWYVALKR	TGQYKLG..S	KTGPGQKAIL
mu FGF-3	RLYRTGSSGP	GAQRQ..PGA	QRPWYVSVNG	KGRPRRG..F	KTRRTQKSSL
mu FGF-4	YAYPGM....FMALSK	NGRTKKG..N	RVSPTMKVTH
mu FGF-5	AIHRTEKTG.REWYVALNK	RGKAKRGCS	P RVKPDHISTH
mu FGF-6	DLYRGT....YIALSK	YGRVKRG..S	KVSPIMTVTH
mu FGF-7	A.....	..KWT..HSG	GEM.FVALNQ	KGIPVKG..K	KTKEQKTAH
mu FGF-8	AKYEG.....WYMAFTR	KGRPRKG..S	KTRQHOREVH
mu FGF-9	NLYK.....H..VDT	GRRYYVALNK	DGTPREG..T	RTKRHQKPTH
mu FGF-10	F.....	..NWQ..HNG	RQM.YVALNG	KGAPRRG..Q	KTRRKNTSAH
mu FGF-11	ALYR.....Q..RRS	GRAWYLGDK	EGRVMKG..N	RVKKTAAAH
mu FGF-12	TLYR.....Q..QES	GRAWFLGLNK	EGQIMKG..N	RVKKTKPSSH
mu FGF-13	MIYR.....Q..QES	GRGWYLGDK	EGEIMKG..N	HVKKNKPAAH
mu FGF-14	MLYR.....Q..QES	GRAWFLGLNK	EGQVMKG..N	RVKKTKPAAH
mu FGF-15	MKHHLHIIFI	QAKP.....REQLO	DQ...KPSNF	...IPVFHRS
rat FGF-16	TLYK.....H..SDS	ERQYYVALNK	DGSPREG..Y	RTKRHQKPTH
mu FGF-17	AKYSG.....WYVGFTK	KGRPRKG..P	KTRENQDQVH

FIG. 2F

hu FGF-1	FLPLPVSSD.
hu FGF-2	FLPMSAKS..
hu FGF-3	FLPRVLDHRD	HEM....VRQ	LQSGLP RP PG	KGVP RRRRQ	K.QSPDNLEP
hu FGF-4	FLPRL.....
hu FGF-5	FLPRFKQSEQ	PEL....SFT	VTVP EKKNPP	SPIKSK....
hu FGF-6	FLPRI.....
hu FGF-7	FLPMAI.....
hu FGF-8	FMKRLPRG..	.H.....	HTTEQSLRFE	FLNYPPFTRS	LRGSQRTWAP
hu FGF-9	FLPRPVD...PDKV..
hu FGF-10	FLPMVVH...
hu FGF-11	FLPKLLEVAM	YQE....PSL	HSVPEASPSS	PPAP.....
hu FGF-12	FVPKPIEVCM	YRE....PSL	HEIG EKQGR.SRKSS.
hu FGF-13	FLPKPLKVAM	YKE....PSL	HDLTEFSRSG	SGTPTKSRS.
hu FGF-14	FLPKPLEVAM	YRE....PSL	HDVGETV PKP	GVTPSKSTS.
hu FGF-16	FLPRPVD...PSKL..
hu FGF-17	FIKRLYQG..	.QL....PFP	NHAEKQKQFB	FVGSAPTRRT	...KTRRRP
hu FGF-18	FMKRYPKG..	.Q.....	..PELQKPFK	YTTVTKRSRR	IRPTHPA...
<u>hu FGF-19</u>	<u>PKEPEDLRGH</u>	<u>LR.....</u>	<u>.....SDM</u>	<u>FSSPLETDSM</u>	<u>..DPFGLVTG</u>
<u>hu FGF-23</u>	<u>SAEDDSE RDP</u>	<u>LNVLKPRARM</u>	<u>TPAPASCSOR</u>	<u>LPSAEDNSPM</u>	<u>ASDPLGVVRG</u>
mu FGF-1	FLPLPVSSD.
mu FGF-2	FLPMSAKS..
mu FGF-3	FLPRVLGHKD	HEM....VRL	LQSSQPRAPG	EGSQPRQRQ	KKQSPGDHGK
mu FGF-4	FLPRL.....
mu FGF-5	FLPRFKQSEQ	PEL....SFT	VTVP EKKKPP	..VKPK....
mu FGF-6	FLPRI.....
mu FGF-7	FLPMAI.....
mu FGF-8	FMKRLPRG..	.H.....	HTTEQSLRFE	FLNYPPFTRS	LRGSQRTWAP
mu FGF-9	FLPRPVD...PDKV..
mu FGF-10	FLPMTIQ...
mu FGF-11	FVPKLEEVAM	YRE....PSL	HSVPE TSPSS	PPAH.....
mu FGF-12	FVPKPIEVCM	YRE....PSL	HEIG EKQGR.SRKSS.
mu FGF-13	FLPKPLKVAM	YKE....PSL	HDLTEFSRSG	SGTPTKSRS.
mu FGF-14	FLPKPLEVAM	YRE....PSL	HDVGETV PKA	GVTPSKSTS.
mu FGF-15	FFETGD...Q	LR.....SKM	FSLPLESDSM	..DPFRMVED
rat FGF-16	FLPRPVD...PSKL..
mu FGF-17	FMKRYPKG..	.Q.....	..AELQKPFK	YTTVTKRSRR	IRPTHPG...

FIG. 2G

```

hu FGF-1 ..... ..
hu FGF-2 ..... ..
hu FGF-3 SHVQASRL.. GSQLEASA.. .H
hu FGF-4 ..... ..
hu FGF-5 IPLSAPRKNT NSVKYRLKFR FG
hu FGF-6 ..... ..
hu FGF-7 ..... ..T
hu FGF-8 EP..... .R
hu FGF-9 ..... .PELYKDILS QS
hu FGF-10 ..... .S
hu FGF-11 ..... ..
hu FGF-12 ...GTPTMNG GKVVNQ...D ST
hu FGF-13 ...VSGVLNG GKSM SHN..E ST
hu FGF-14 ...ASAIMNG GKPVNKS..K TT
hu FGF-16 ..... .PSMSRDLFH YR
hu FGF-17 QPL..... .T
hu FGF-18 ..... ..
hu FGF-19 LEA.VRS... ..PSFEK ..
hu FGF-23 GRVNT HAGGT GPEGCRPFAK FI
mu FGF-1 ..... ..
mu FGF-2 ..... ..
mu FGF-3 METLSTRATP STQLHTGGLA VA
mu FGF-4 ..... ..
mu FGF-5 VPLSQPRRSP SPVKYRLKFR FG
mu FGF-6 ..... ..
mu FGF-7 ..... ..T
mu FGF-8 EP..... .R
mu FGF-9 ..... .PELYKDILS QS
mu FGF-10 ..... ..T
mu FGF-11 ..... ..
mu FGF-12 ...GTPTMNG GKVVNQ...D ST
mu FGF-13 ...VSGVLNG GKSM SHN..E ST
mu FGF-14 ...ASAIMNG GKPVNKC..K TT
mu FGF-15 VDHLVKS... ..PSFQK ..
rat FGF-16 ..... .PSMSRDLFR YR
mu FGF-17 ..... ..

```

FIG. 3

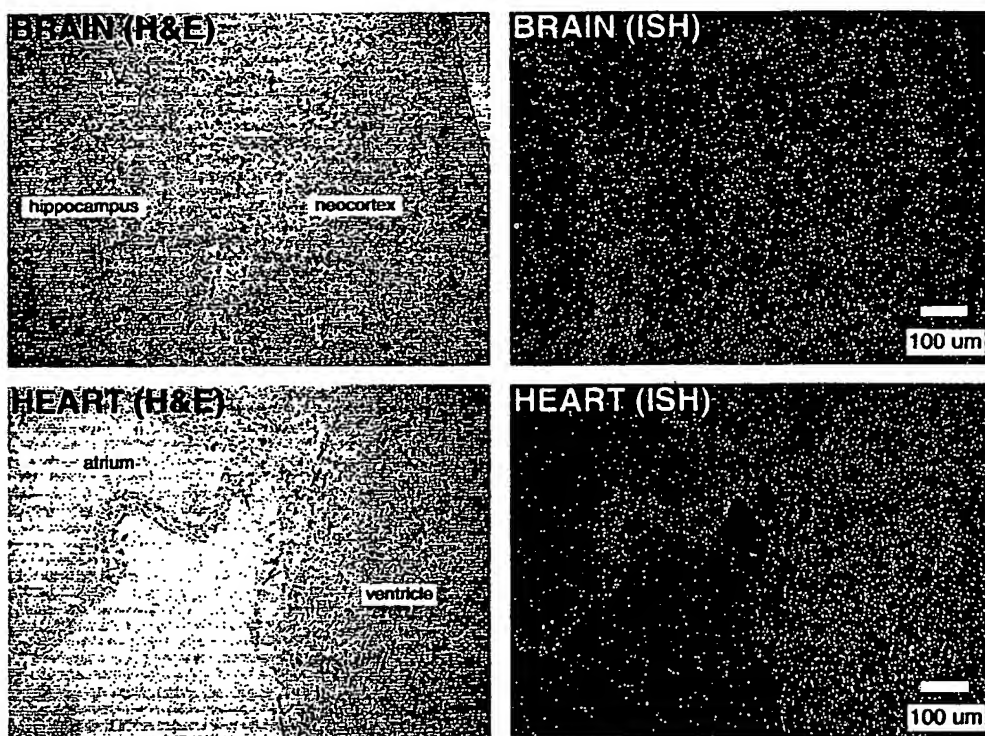


FIG. 4

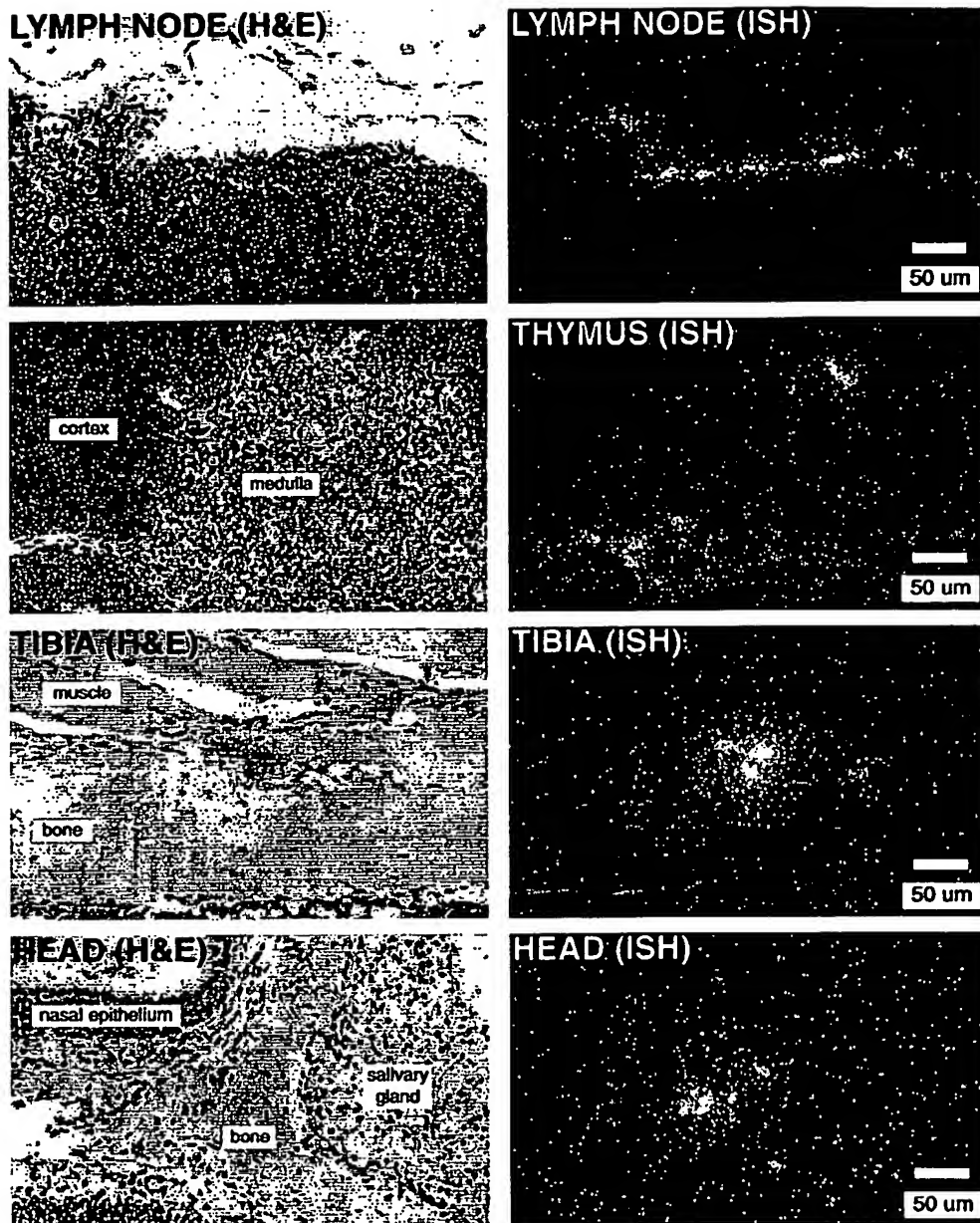


FIG. 5

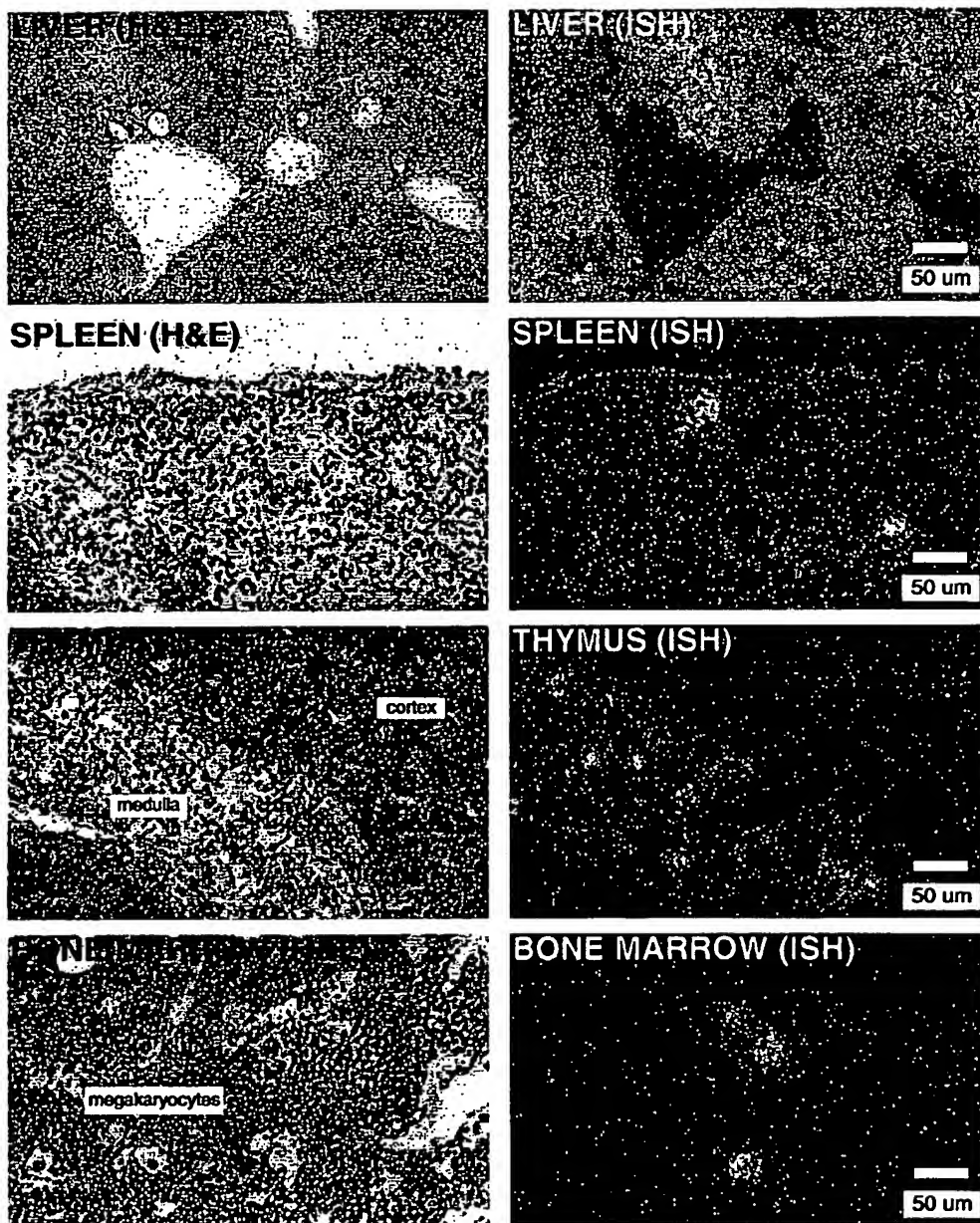
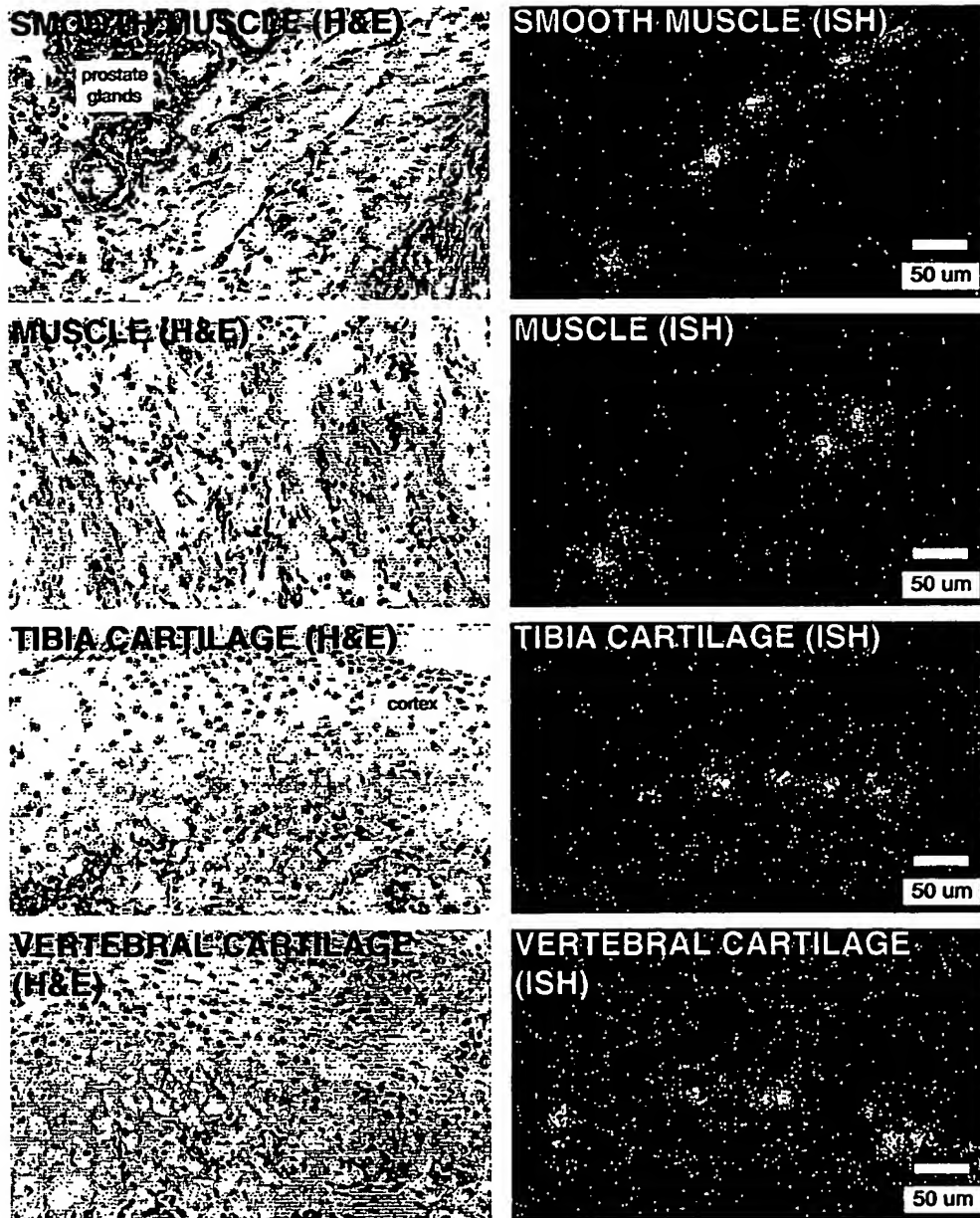


FIG. 6



SEQUENCE LISTING

<110> Luethy, Roland
 Yang, Robert
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<120> Fibroblast Growth Factor-23 Molecules and Uses Thereof

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His	Leu	Tyr	Thr	Ala	Thr	Ala	Arg	Asn	Ser	Tyr	His	Leu	Gln	Ile	His
			20					25						30	
Lys	Asn	Gly	His	Val	Asp	Gly	Ala	Pro	His	Gln	Thr	Ile	Tyr	Ser	Ala
		35					40					45			
Leu	Met	Ile	Arg	Ser	Glu	Asp	Ala	Gly	Phe	Val	Val	Ile	Thr	Gly	Val
	50					55					60				
Met	Ser	Arg	Arg	Tyr	Leu	Cys	Met	Asp	Phe	Arg	Gly	Asn	Ile	Phe	Gly
	65				70					75					80
Ser	His	Tyr	Phe	Asp	Pro	Glu	Asn	Cys	Arg	Phe	Gln	His	Gln	Thr	Leu
				85					90					95	
Glu	Asn	Gly	Tyr	Asp	Val	Tyr	His	Ser	Pro	Gln	Tyr	His	Phe	Leu	Val
		100						105					110		
Ser	Leu	Gly	Arg	Ala	Lys	Arg	Ala	Phe	Leu	Pro	Gly	Met	Asn	Pro	Pro
		115					120					125			
Pro	Tyr	Ser	Gln	Phe	Leu	Ser	Arg	Arg	Asn	Glu	Ile	Pro	Leu	Ile	His

130 135 140
 Phe Asn Thr Pro Ile Pro Arg Arg His Thr Arg Ser Ala Glu Asp Asp
 145 150 155 160
 Ser Glu Arg Asp Pro Leu Asn Val Leu Lys Pro Arg Ala Arg Met Thr
 165 170 175
 Pro Ala Pro Ala Ser Cys Ser Gln Glu Leu Pro Ser Ala Glu Asp Asn
 180 185 190
 Ser Pro Met Ala Ser Asp Pro Leu Gly Val Val Arg Gly Gly Arg Val
 195 200 205
 Asn Thr His Ala Gly Gly Thr Gly Pro Glu Gly Cys Arg Pro Phe Ala
 210 215 220
 Lys Phe Ile
 225

<210> 4
 <211> 155
 <212> PRT
 <213> Homo sapiens

<400> 4
 Met Ala Glu Gly Glu Ile Thr Thr Phe Thr Ala Leu Thr Glu Lys Phe
 1 5 10 15
 Asn Leu Pro Pro Gly Asn Tyr Lys Lys Pro Lys Leu Leu Tyr Cys Ser
 20 25 30
 Asn Gly Gly His Phe Leu Arg Ile Leu Pro Asp Gly Thr Val Asp Gly
 35 40 45
 Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu Ser Ala Glu
 50 55 60
 Ser Val Gly Glu Val Tyr Ile Lys Ser Thr Glu Thr Gly Gln Tyr Leu
 65 70 75 80
 Ala Met Asp Thr Asp Gly Leu Leu Tyr Gly Ser Gln Thr Pro Asn Glu
 85 90 95
 Glu Cys Leu Phe Leu Glu Arg Leu Glu Glu Asn His Tyr Asn Thr Tyr
 100 105 110
 Ile Ser Lys Lys His Ala Glu Lys Asn Trp Phe Val Gly Leu Lys Lys
 115 120 125
 Asn Gly Ser Cys Lys Arg Gly Pro Arg Thr His Tyr Gly Gln Lys Ala
 130 135 140
 Ile Leu Phe Leu Pro Leu Pro Val Ser Ser Asp
 145 150 155

<210> 5
 <211> 155
 <212> PRT

<213> Homo sapiens

<400> 5

```

Met Ala Ala Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly
 1              5              10              15

Gly Ser Gly Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu
              20              25              30

Tyr Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg
              35              40              45

Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile Lys Leu Gln Leu
              50              55              60

Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn
              65              70              75              80

Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys
              85              90              95

Val Thr Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr
              100              105              110

Asn Thr Tyr Arg Ser Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys
              115              120              125

Arg Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys
              130              135              140

Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145              150              155

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<210> 6

<211> 239

<212> PRT

<213> Homo sapiens

<400> 6

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Met Gly Leu Ile Trp Leu Leu Leu Leu Ser Leu Leu Glu Pro Gly Trp
 1              5              10              15

Pro Ala Ala Gly Pro Gly Ala Arg Leu Arg Arg Asp Ala Gly Gly Arg
              20              25              30

Gly Gly Val Tyr Glu His Leu Gly Gly Ala Pro Arg Arg Arg Lys Leu
              35              40              45

Tyr Cys Ala Thr Lys Tyr His Leu Gln Leu His Pro Ser Gly Arg Val
              50              55              60

Asn Gly Ser Leu Glu Asn Ser Ala Tyr Ser Ile Leu Glu Ile Thr Ala
              65              70              75              80

Val Glu Val Gly Ile Val Ala Ile Arg Gly Leu Phe Ser Gly Arg Tyr
              85              90              95

Leu Ala Met Asn Lys Arg Gly Arg Leu Tyr Ala Ser Glu His Tyr Ser
              100              105              110

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Ala Glu Cys Glu Phe Val Glu Arg Ile His Glu Leu Gly Tyr Asn Thr
 115 120 125
 Tyr Ala Ser Arg Leu Tyr Arg Thr Val Ser Ser Thr Pro Gly Ala Arg
 130 135 140
 Arg Gln Pro Ser Ala Glu Arg Leu Trp Tyr Val Ser Val Asn Gly Lys
 145 150 155 160
 Gly Arg Pro Arg Arg Gly Phe Lys Thr Arg Arg Thr Gln Lys Ser Ser
 165 170 175
 Leu Phe Leu Pro Arg Val Leu Asp His Arg Asp His Glu Met Val Arg
 180 185 190
 Gln Leu Gln Ser Gly Leu Pro Arg Pro Pro Gly Lys Gly Val Gln Pro
 195 200 205
 Arg Arg Arg Arg Gln Lys Gln Ser Pro Asp Asn Leu Glu Pro Ser His
 210 215 220
 Val Gln Ala Ser Arg Leu Gly Ser Gln Leu Glu Ala Ser Ala His
 225 230 235

<210> 7

<211> 206

<212> PRT

<213> Homo sapiens

<400> 7

Met Ser Gly Pro Gly Thr Ala Ala Val Ala Leu Leu Pro Ala Val Leu
 1 5 10 15
 Leu Ala Leu Leu Ala Pro Trp Ala Gly Arg Gly Gly Ala Ala Ala Pro
 20 25 30
 Thr Ala Pro Asn Gly Thr Leu Glu Ala Glu Leu Glu Arg Arg Trp Glu
 35 40 45
 Ser Leu Val Ala Leu Ser Leu Ala Arg Leu Pro Val Ala Ala Gln Pro
 50 55 60
 Lys Glu Ala Ala Val Gln Ser Gly Ala Gly Asp Tyr Leu Leu Gly Ile
 65 70 75 80
 Lys Arg Leu Arg Arg Leu Tyr Cys Asn Val Gly Ile Gly Phe His Leu
 85 90 95
 Gln Ala Leu Pro Asp Gly Arg Ile Gly Gly Ala His Ala Asp Thr Arg
 100 105 110
 Asp Ser Leu Leu Glu Leu Ser Pro Val Glu Arg Gly Val Val Ser Ile
 115 120 125
 Phe Gly Val Ala Ser Arg Phe Phe Val Ala Met Ser Ser Lys Gly Lys
 130 135 140
 Leu Tyr Gly Ser Pro Phe Phe Thr Asp Glu Cys Thr Phe Lys Glu Ile
 145 150 155 160

Leu Leu Pro Asn Asn Tyr Asn Ala Tyr Glu Ser Tyr Lys Tyr Pro Gly
 165 170 175

Met Phe Ile Ala Leu Ser Lys Asn Gly Lys Thr Lys Lys Gly Asn Arg
 180 185 190

Val Ser Pro Thr Met Lys Val Thr His Phe Leu Pro Arg Leu
 195 200 205

<210> 8

<211> 268

<212> PRT

<213> Homo sapiens

<400> 8

Met Ser Leu Ser Phe Leu Leu Leu Leu Phe Phe Ser His Leu Ile Leu
 1 5 10 15

Ser Ala Trp Ala His Gly Glu Lys Arg Leu Ala Pro Lys Gly Gln Pro
 20 25 30

Gly Pro Ala Ala Thr Asp Arg Asn Pro Ile Gly Ser Ser Ser Arg Gln
 35 40 45

Ser Ser Ser Ser Ala Met Ser Ser Ser Ser Ala Ser Ser Ser Pro Ala
 50 55 60

Ala Ser Leu Gly Ser Gln Gly Ser Gly Leu Glu Gln Ser Ser Phe Gln
 65 70 75 80

Trp Ser Pro Ser Gly Arg Arg Thr Gly Ser Leu Tyr Cys Arg Val Gly
 85 90 95

Ile Gly Phe His Leu Gln Ile Tyr Pro Asp Gly Lys Val Asn Gly Ser
 100 105 110

His Glu Ala Asn Met Leu Ser Val Leu Glu Ile Phe Ala Val Ser Gln
 115 120 125

Gly Ile Val Gly Ile Arg Gly Val Phe Ser Asn Lys Phe Leu Ala Met
 130 135 140

Ser Lys Lys Gly Lys Leu His Ala Ser Ala Lys Phe Thr Asp Asp Cys
 145 150 155 160

Lys Phe Arg Glu Arg Phe Gln Glu Asn Ser Tyr Asn Thr Tyr Ala Ser
 165 170 175

Ala Ile His Arg Thr Glu Lys Thr Gly Arg Glu Trp Tyr Val Ala Leu
 180 185 190

Asn Lys Arg Gly Lys Ala Lys Arg Gly Cys Ser Pro Arg Val Lys Pro
 195 200 205

Gln His Ile Ser Thr His Phe Leu Pro Arg Phe Lys Gln Ser Glu Gln
 210 215 220

Pro Glu Leu Ser Phe Thr Val Thr Val Pro Glu Lys Lys Asn Pro Pro
 225 230 235 240

Ser Pro Ile Lys Ser Lys Ile Pro Leu Ser Ala Pro Arg Lys Asn Thr
 245 250 255

Asn Ser Val Lys Tyr Arg Leu Lys Phe Arg Phe Gly
 260 265

<210> 9
 <211> 208
 <212> PRT
 <213> Homo sapiens

<400> 9
 Met Ala Leu Gly Gln Lys Leu Phe Ile Thr Met Ser Arg Gly Ala Gly
 1 5 10 15
 Arg Leu Gln Gly Thr Leu Trp Ala Leu Val Phe Leu Gly Ile Leu Val
 20 25 30
 Gly Met Val Val Pro Ser Pro Ala Gly Thr Arg Ala Asn Asn Thr Leu
 35 40 45
 Leu Asp Ser Arg Gly Trp Gly Thr Leu Leu Ser Arg Ser Arg Ala Gly
 50 55 60
 Leu Ala Gly Glu Ile Ala Gly Val Asn Trp Glu Ser Gly Tyr Leu Val
 65 70 75 80
 Gly Ile Lys Arg Gln Arg Arg Leu Tyr Cys Asn Val Gly Ile Gly Phe
 85 90 95
 His Leu Gln Val Leu Pro Asp Gly Arg Ile Ser Gly Thr His Glu Glu
 100 105 110
 Asn Pro Tyr Ser Leu Leu Glu Ile Ser Thr Val Glu Arg Gly Val Val
 115 120 125
 Ser Leu Phe Gly Val Arg Ser Ala Leu Phe Val Ala Met Asn Ser Lys
 130 135 140
 Gly Arg Leu Tyr Ala Thr Pro Ser Phe Gln Glu Glu Cys Lys Phe Arg
 145 150 155 160
 Glu Thr Leu Leu Pro Asn Asn Tyr Asn Ala Tyr Glu Ser Asp Leu Tyr
 165 170 175
 Gln Gly Thr Tyr Ile Ala Leu Ser Lys Tyr Gly Arg Val Lys Arg Gly
 180 185 190
 Ser Lys Val Ser Pro Ile Met Thr Val Thr His Phe Leu Pro Arg Ile
 195 200 205

<210> 10
 <211> 194
 <212> PRT
 <213> Homo sapiens

<400> 10

Met His Lys Trp Ile Leu Thr Trp Ile Leu Pro Thr Leu Leu Tyr Arg
 1 5 10 15

Ser Cys Phe His Ile Ile Cys Leu Val Gly Thr Ile Ser Leu Ala Cys
 20 25 30

Asn Asp Met Thr Pro Glu Gln Met Ala Thr Asn Val Asn Cys Ser Ser
 35 40 45

Pro Glu Arg His Thr Arg Ser Tyr Asp Tyr Met Glu Gly Gly Asp Ile
 50 55 60

Arg Val Arg Arg Leu Phe Cys Arg Thr Gln Trp Tyr Leu Arg Ile Asp
 65 70 75 80

Lys Arg Gly Lys Val Lys Gly Thr Gln Glu Met Lys Asn Asn Tyr Asn
 85 90 95

Ile Met Glu Ile Arg Thr Val Ala Val Gly Ile Val Ala Ile Lys Gly
 100 105 110

Val Glu Ser Glu Phe Tyr Leu Ala Met Asn Lys Glu Gly Lys Leu Tyr
 115 120 125

Ala Lys Lys Glu Cys Asn Glu Asp Cys Asn Phe Lys Glu Leu Ile Leu
 130 135 140

Glu Asn His Tyr Asn Thr Tyr Ala Ser Ala Lys Trp Thr His Asn Gly
 145 150 155 160

Gly Glu Met Phe Val Ala Leu Asn Gln Lys Gly Ile Pro Val Arg Gly
 165 170 175

Lys Lys Thr Lys Lys Glu Gln Lys Thr Ala His Phe Leu Pro Met Ala
 180 185 190

Ile Thr

<210> 11

<211> 233

<212> PRT

<213> Homo sapiens

<400> 11

Met Gly Ser Pro Arg Ser Ala Leu Ser Cys Leu Leu Leu His Leu Leu
 1 5 10 15

Val Leu Cys Leu Gln Ala Gln Glu Gly Pro Gly Arg Gly Pro Ala Leu
 20 25 30

Gly Arg Glu Leu Ala Ser Leu Phe Arg Ala Gly Arg Glu Pro Gln Gly
 35 40 45

Val Ser Gln Gln His Val Arg Glu Gln Ser Leu Val Thr Asp Gln Leu
 50 55 60

Ser Arg Arg Leu Ile Arg Thr Tyr Gln Leu Tyr Ser Arg Thr Ser Gly
 65 70 75 80

Lys	His	Val	Gln	Val	Leu	Ala	Asn	Lys	Arg	Ile	Asn	Ala	Met	Ala	Glu	
				85					90					95		
Asp	Gly	Asp	Pro	Phe	Ala	Lys	Leu	Ile	Val	Glu	Thr	Asp	Thr	Phe	Gly	
				100					105					110		
Ser	Arg	Val	Arg	Val	Arg	Gly	Ala	Glu	Thr	Gly	Leu	Tyr	Ile	Cys	Met	
				115					120					125		
Asn	Lys	Lys	Gly	Lys	Leu	Ile	Ala	Lys	Ser	Asn	Gly	Lys	Gly	Lys	Asp	
				130					135					140		
Cys	Val	Phe	Thr	Glu	Ile	Val	Leu	Glu	Asn	Asn	Tyr	Thr	Ala	Leu	Gln	
				145					150					155		
Asn	Ala	Lys	Tyr	Glu	Gly	Trp	Tyr	Met	Ala	Phe	Thr	Arg	Lys	Gly	Arg	
				165					170					175		
Pro	Arg	Lys	Gly	Ser	Lys	Thr	Arg	Gln	His	Gln	Arg	Glu	Val	His	Phe	
				180					185					190		
Met	Lys	Arg	Leu	Pro	Arg	Gly	His	His	Thr	Thr	Glu	Gln	Ser	Leu	Arg	
				195					200					205		
Phe	Glu	Phe	Leu	Asn	Tyr	Pro	Pro	Phe	Thr	Arg	Ser	Leu	Arg	Gly	Ser	
				210					215					220		
Gln	Arg	Thr	Trp	Ala	Pro	Glu	Pro	Arg								
				225					230							

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<210> 12
<211> 208
<212> PRT
<213> Homo sapiens
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<400> 12
Met Ala Pro Leu Gly Glu Val Gly Asn Tyr Phe Gly Val Gln Asp Ala
  1              5              10              15

Val Pro Phe Gly Asn Val Pro Val Leu Pro Val Asp Ser Pro Val Leu
      20              25              30

Leu Ser Asp His Leu Gly Gln Ser Glu Ala Gly Gly Leu Pro Arg Gly
      35              40              45

Pro Ala Val Thr Asp Leu Asp His Leu Lys Gly Ile Leu Arg Arg Arg
      50              55              60

Gln Leu Tyr Cys Arg Thr Gly Phe His Leu Glu Ile Phe Pro Asn Gly
      65              70              75              80

Thr Ile Gln Gly Thr Arg Lys Asp His Ser Arg Phe Gly Ile Leu Glu
      85              90              95

Phe Ile Ser Ile Ala Val Gly Leu Val Ser Ile Arg Gly Val Asp Ser
      100              105              110

Gly Leu Tyr Leu Gly Met Asn Glu Lys Gly Glu Leu Tyr Gly Ser Glu
      115              120              125

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Lys Leu Thr Gln Glu Cys Val Phe Arg Glu Gln Phe Glu Glu Asn Trp
 130 135 140
 Tyr Asn Thr Tyr Ser Ser Asn Leu Tyr Lys His Val Asp Thr Gly Arg
 145 150 155 160
 Arg Tyr Tyr Val Ala Leu Asn Lys Asp Gly Thr Pro Arg Glu Gly Thr
 165 170 175
 Arg Thr Lys Arg His Gln Lys Phe Thr His Phe Leu Pro Arg Pro Val
 180 185 190
 Asp Pro Asp Lys Val Pro Glu Leu Tyr Lys Asp Ile Leu Ser Gln Ser
 195 200 205

<210> 13
 <211> 208
 <212> PRT
 <213> Homo sapiens

<400> 13
 Met Trp Lys Trp Ile Leu Thr His Cys Ala Ser Ala Phe Pro His Leu
 1 5 10 15
 Pro Gly Cys Cys Cys Cys Cys Phe Leu Leu Phe Leu Val Ser Ser
 20 25 30
 Val Pro Val Thr Cys Gln Ala Leu Gly Gln Asp Met Val Ser Pro Glu
 35 40 45
 Ala Thr Asn Ser Ser Ser Ser Ser Phe Ser Ser Pro Ser Ser Ala Gly
 50 55 60
 Arg His Val Arg Ser Tyr Asn His Leu Gln Gly Asp Val Arg Trp Arg
 65 70 75 80
 Lys Leu Phe Ser Phe Thr Lys Tyr Phe Leu Lys Ile Glu Lys Asn Gly
 85 90 95
 Lys Val Ser Gly Thr Lys Lys Glu Asn Cys Pro Tyr Ser Ile Leu Glu
 100 105 110
 Ile Thr Ser Val Glu Ile Gly Val Val Ala Val Lys Ala Ile Asn Ser
 115 120 125
 Asn Tyr Tyr Leu Ala Met Asn Lys Lys Gly Lys Leu Tyr Gly Ser Lys
 130 135 140
 Glu Phe Asn Asn Asp Cys Lys Leu Lys Glu Arg Ile Glu Glu Asn Gly
 145 150 155 160
 Tyr Asn Thr Tyr Ala Ser Phe Asn Trp Gln His Asn Gly Arg Gln Met
 165 170 175
 Tyr Val Ala Leu Asn Gly Lys Gly Ala Pro Arg Arg Gly Gln Lys Thr
 180 185 190

Arg Arg Lys Asn Thr Ser Ala His Phe Leu Pro Met Val Val His Ser
 195 200 205

<210> 14
 <211> 225
 <212> PRT
 <213> Homo sapiens

<400> 14
 Met Ala Ala Leu Ala Ser Ser Leu Ile Arg Gln Lys Arg Glu Val Arg
 1 5 10 15
 Glu Pro Gly Gly Ser Arg Pro Val Ser Ala Gln Arg Arg Val Cys Pro
 20 25 30
 Arg Gly Thr Lys Ser Leu Cys Gln Lys Gln Leu Leu Ile Leu Leu Ser
 35 40 45
 Lys Val Arg Leu Cys Gly Gly Arg Pro Ala Arg Pro Asp Arg Gly Pro
 50 55 60
 Glu Pro Gln Leu Lys Gly Ile Val Thr Lys Leu Phe Cys Arg Gln Gly
 65 70 75 80
 Phe Tyr Leu Gln Ala Asn Pro Asp Gly Ser Ile Gln Gly Thr Pro Glu
 85 90 95
 Asp Thr Ser Ser Phe Thr His Phe Asn Leu Ile Pro Val Gly Leu Arg
 100 105 110
 Val Val Thr Ile Gln Ser Ala Lys Leu Gly His Tyr Met Ala Met Asn
 115 120 125
 Ala Glu Gly Leu Leu Tyr Ser Ser Pro His Phe Thr Ala Glu Cys Arg
 130 135 140
 Phe Lys Glu Cys Val Phe Glu Asn Tyr Tyr Val Leu Tyr Ala Ser Ala
 145 150 155 160
 Leu Tyr Arg Gln Arg Arg Ser Gly Arg Ala Trp Tyr Leu Gly Leu Asp
 165 170 175
 Lys Glu Gly Gln Val Met Lys Gly Asn Arg Val Lys Lys Thr Lys Ala
 180 185 190
 Ala Ala His Phe Leu Pro Lys Leu Leu Glu Val Ala Met Tyr Gln Glu
 195 200 205
 Pro Ser Leu His Ser Val Pro Glu Ala Ser Pro Ser Ser Pro Pro Ala
 210 215 220

Pro
 225

<210> 15

<211> 243
 <212> PRT
 <213> Homo sapiens

<400> 15
 Met Ala Ala Ala Ile Ala Ser Ser Leu Ile Arg Gln Lys Arg Gln Ala
 1 5 10 15
 Arg Glu Ser Asn Ser Asp Arg Val Ser Ala Ser Lys Arg Arg Ser Ser
 20 25 30
 Pro Ser Lys Asp Gly Arg Ser Leu Cys Glu Arg His Val Leu Gly Val
 35 40 45
 Phe Ser Lys Val Arg Phe Cys Ser Gly Arg Lys Arg Pro Val Arg Arg
 50 55 60
 Arg Pro Glu Pro Gln Leu Lys Gly Ile Val Thr Arg Leu Phe Ser Gln
 65 70 75 80
 Gln Gly Tyr Phe Leu Gln Met His Pro Asp Gly Thr Ile Asp Gly Thr
 85 90 95
 Lys Asp Glu Asn Ser Asp Tyr Thr Leu Phe Asn Leu Ile Pro Val Gly
 100 105 110
 Leu Arg Val Val Ala Ile Gln Gly Val Lys Ala Ser Leu Tyr Val Ala
 115 120 125
 Met Asn Gly Glu Gly Tyr Leu Tyr Ser Ser Asp Val Phe Thr Pro Glu
 130 135 140
 Cys Lys Phe Lys Glu Ser Val Phe Glu Asn Tyr Tyr Val Ile Tyr Ser
 145 150 155 160
 Ser Thr Leu Tyr Arg Gln Gln Glu Ser Gly Arg Ala Trp Phe Leu Gly
 165 170 175
 Leu Asn Lys Glu Gly Gln Ile Met Lys Gly Asn Arg Val Lys Lys Thr
 180 185 190
 Lys Pro Ser Ser His Phe Val Pro Lys Pro Ile Glu Val Cys Met Tyr
 195 200 205
 Arg Glu Pro Ser Leu His Glu Ile Gly Glu Lys Gln Gly Arg Ser Arg
 210 215 220
 Lys Ser Ser Gly Thr Pro Thr Met Asn Gly Gly Lys Val Val Asn Gln
 225 230 235 240
 Asp Ser Thr

<210> 16
 <211> 245
 <212> PRT
 <213> Homo sapiens

<400> 16
 Met Ala Ala Ala Ile Ala Ser Ser Leu Ile Arg Gln Lys Arg Gln Ala

1 5 10 15
 Arg Glu Arg Glu Lys Ser Asn Ala Cys Lys Cys Val Ser Ser Pro Ser
 20 25 30
 Lys Gly Lys Thr Ser Cys Asp Lys Asn Lys Leu Asn Val Phe Ser Arg
 35 40 45
 Val Lys Leu Phe Gly Ser Lys Lys Arg Arg Arg Arg Pro Glu Pro
 50 55 60
 Gln Leu Lys Gly Ile Val Thr Lys Leu Tyr Ser Arg Gln Gly Tyr His
 65 70 75 80
 Leu Gln Leu Gln Ala Asp Gly Thr Ile Asp Gly Thr Lys Asp Glu Asp
 85 90 95
 Ser Thr Tyr Thr Leu Phe Asn Leu Ile Pro Val Gly Leu Arg Val Val
 100 105 110
 Ala Ile Gln Gly Val Gln Thr Lys Leu Tyr Leu Ala Met Asn Ser Glu
 115 120 125
 Gly Tyr Leu Tyr Thr Ser Glu Leu Phe Thr Pro Glu Cys Lys Phe Lys
 130 135 140
 Glu Ser Val Phe Glu Asn Tyr Tyr Val Thr Tyr Ser Ser Met Ile Tyr
 145 150 155 160
 Arg Gln Gln Gln Ser Gly Arg Gly Trp Tyr Leu Gly Leu Asn Lys Glu
 165 170 175
 Gly Glu Ile Met Lys Gly Asn His Val Lys Lys Asn Lys Pro Ala Ala
 180 185 190
 His Phe Leu Pro Lys Pro Leu Lys Val Ala Met Tyr Lys Glu Pro Ser
 195 200 205
 Leu His Asp Leu Thr Glu Phe Ser Arg Ser Gly Ser Gly Thr Pro Thr
 210 215 220
 Lys Ser Arg Ser Val Ser Gly Val Leu Asn Gly Gly Lys Ser Met Ser
 225 230 235 240
 His Asn Glu Ser Thr
 245

<210> 17

<211> 247

<212> PRT

<213> Homo sapiens

<400> 17

Met Ala Ala Ala Ile Ala Ser Gly Leu Ile Arg Gln Lys Arg Gln Ala
 1 5 10 15
 Arg Glu Gln His Trp Asp Arg Pro Ser Ala Ser Arg Arg Ser Ser
 20 25 30
 Pro Ser Lys Asn Arg Gly Leu Cys Asn Gly Asn Leu Val Asp Ile Phe

35 40 45
 Ser Lys Val Arg Ile Phe Gly Leu Lys Lys Arg Arg Leu Arg Arg Gln
 50 55 60
 Asp Pro Gln Leu Lys Gly Ile Val Thr Arg Leu Tyr Cys Arg Gln Gly
 65 70 75 80
 Tyr Tyr Leu Gln Met His Pro Asp Gly Ala Leu Asp Gly Thr Lys Asp
 85 90 95
 Asp Ser Thr Asn Ser Thr Leu Phe Asn Leu Ile Pro Val Gly Leu Arg
 100 105 110
 Val Val Ala Ile Gln Gly Val Lys Thr Gly Leu Tyr Ile Ala Met Asn
 115 120 125
 Gly Glu Gly Tyr Leu Tyr Pro Ser Glu Leu Phe Thr Pro Glu Cys Lys
 130 135 140
 Phe Lys Glu Ser Val Phe Glu Asn Tyr Tyr Val Ile Tyr Ser Ser Met
 145 150 155 160
 Leu Tyr Arg Gln Gln Glu Ser Gly Arg Ala Trp Phe Leu Gly Leu Asn
 165 170 175
 Lys Glu Gly Gln Ala Met Lys Gly Asn Arg Val Lys Lys Thr Lys Pro
 180 185 190
 Ala Ala His Phe Leu Pro Lys Pro Leu Glu Val Ala Met Tyr Arg Glu
 195 200 205
 Pro Ser Leu His Asp Val Gly Glu Thr Val Pro Lys Pro Gly Val Thr
 210 215 220
 Pro Ser Lys Ser Thr Ser Ala Ser Ala Ile Met Asn Gly Gly Lys Pro
 225 230 235 240
 Val Asn Lys Ser Lys Thr Thr
 245

<210> 18

<211> 207

<212> PRT

<213> Homo sapiens

<400> 18

Met Ala Glu Val Gly Gly Val Phe Ala Ser Leu Asp Trp Asp Leu His
 1 5 10 15
 Gly Phe Ser Ser Ser Leu Gly Asn Val Pro Leu Ala Asp Ser Pro Gly
 20 25 30
 Phe Leu Asn Glu Arg Leu Gly Gln Ile Glu Gly Lys Leu Gln Arg Gly
 35 40 45
 Ser Pro Thr Asp Phe Ala His Leu Lys Gly Ile Leu Arg Arg Arg Gln
 50 55 60
 Leu Tyr Cys Arg Thr Gly Phe His Leu Glu Ile Phe Pro Asn Gly Thr

65	70	75	80
Val His Gly Thr Arg	His Asp His Ser Arg	Phe Gly Ile Leu Glu Phe	
	85	90	95
Ile Ser Leu Ala Val Gly Leu Ile Ser Ile Arg Gly Val Asp Ser Gly			
	100	105	110
Leu Tyr Leu Gly Met Asn Glu Arg Gly Glu Leu Tyr Gly Ser Lys Lys			
	115	120	125
Leu Thr Arg Glu Cys Val Phe Arg Glu Gln Phe Glu Glu Asn Trp Tyr			
	130	135	140
Asn Thr Tyr Ala Ser Thr Leu Tyr Lys His Ser Asp Ser Glu Arg Gln			
	145	150	155
Tyr Tyr Val Ala Leu Asn Lys Asp Gly Ser Pro Arg Glu Gly Tyr Arg			
	165	170	175
Thr Lys Arg His Gln Lys Phe Thr His Phe Leu Pro Arg Pro Val Asp			
	180	185	190
Pro Ser Lys Leu Pro Ser Met Ser Arg Asp Leu Phe His Tyr Arg			
	195	200	205

<210> 19

<211> 207

<212> PRT

<213> Homo sapiens

<400> 19

Met Tyr Ser Ala Pro Ser Ala Cys Thr Cys Leu Cys Leu His Phe Leu			
1	5	10	15
Leu Leu Cys Phe Gln Val Gln Val Leu Val Ala Glu Glu Asn Val Asp			
	20	25	30
Phe Arg Ile His Val Glu Asn Gln Thr Arg Ala Arg Asp Asp Val Ser			
	35	40	45
Arg Lys Gln Leu Arg Leu Tyr Gln Leu Tyr Ser Arg Thr Ser Gly Lys			
	50	55	60
His Ile Gln Val Leu Gly Arg Arg Ile Ser Ala Arg Gly Glu Asp Gly			
	65	70	75
Asp Lys Tyr Ala Gln Leu Leu Val Glu Thr Asp Thr Phe Gly Ser Gln			
	85	90	95
Val Arg Ile Lys Gly Lys Glu Thr Glu Phe Tyr Leu Cys Met Asn Arg			
	100	105	110
Lys Gly Lys Leu Val Gly Lys Pro Asp Gly Thr Ser Lys Glu Cys Val			
	115	120	125
Phe Ile Glu Lys Val Leu Glu Asn Asn Tyr Thr Ala Leu Met Ser Ala			
	130	135	140
Lys Tyr Ser Gly Trp Tyr Val Gly Phe Thr Lys Lys Gly Arg Pro Arg			

145							150							155							160
Lys	Gly	Pro	Lys	Thr	Arg	Glu	Asn	Gln	Gln	Asp	Val	His	Phe	Met	Lys						
				165					170					175							
Arg	Tyr	Pro	Lys	Gly	Gln	Pro	Glu	Leu	Gln	Lys	Pro	Phe	Lys	Tyr	Thr						
			180					185					190								
Thr	Val	Thr	Lys	Arg	Ser	Arg	Arg	Ile	Arg	Pro	Thr	His	Pro	Ala							
		195					200					205									

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<210> 20
<211> 216
<212> PRT
<213> Homo sapiens
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[illegible]

<210> 21
 <211> 233
 <212> PRT
 <213> Homo sapiens

<400> 21

```

Met Ser Val Leu Arg Ala Tyr Pro Asn Ala Ser Pro Leu Leu Gly Ser
 1           5           10           15

Ser Trp Gly Gly Leu Ile His Leu Tyr Thr Ala Thr Ala Arg Asn Ser
           20           25           30

Tyr His Leu Gln Ile His Lys Asn Gly His Val Asp Gly Ala Pro His
           35           40           45

Gln Thr Ile Tyr Ser Ala Leu Met Ile Arg Ser Glu Asp Ala Gly Phe
           50           55           60

Val Val Ile Thr Gly Val Met Ser Arg Arg Tyr Leu Cys Met Asp Phe
           65           70           75           80

Arg Gly Asn Ile Phe Gly Ser His Tyr Phe Asp Pro Glu Asn Cys Arg
           85           90           95

Phe Gln His Gln Thr Leu Glu Asn Gly Tyr Asp Val Tyr His Ser Pro
           100          105          110

Gln Tyr His Phe Leu Val Ser Leu Gly Arg Ala Lys Arg Ala Phe Leu
           115          120          125

Pro Gly Met Asn Pro Pro Pro Tyr Ser Gln Phe Leu Ser Arg Arg Asn
           130          135          140

Glu Ile Pro Leu Ile His Phe Asn Thr Pro Ile Pro Arg Arg His Thr
           145          150          155          160

Arg Ser Ala Glu Asp Asp Ser Glu Arg Asp Pro Leu Asn Val Leu Lys
           165          170          175

Pro Arg Ala Arg Met Thr Pro Ala Pro Ala Ser Cys Ser Gln Glu Leu
           180          185          190

Pro Ser Ala Glu Asp Asn Ser Pro Met Ala Ser Asp Pro Leu Gly Val
           195          200          205

Val Arg Gly Gly Arg Val Asn Thr His Ala Gly Gly Thr Gly Pro Glu
           210          215          220

Gly Cys Arg Pro Phe Ala Lys Phe Ile
           225          230

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<210> 22
 <211> 155
 <212> PRT
 <213> Mus musculus

<400> 22

```

Met Ala Glu Gly Glu Ile Thr Thr Phe Ala Ala Leu Thr Glu Arg Phe
 1           5           10           15

```

Asn Leu Pro Leu Gly Asn Tyr Lys Lys Pro Lys Leu Leu Tyr Cys Ser
 20 25 30
 Asn Gly Gly His Phe Leu Arg Ile Leu Pro Asp Gly Thr Val Asp Gly
 35 40 45
 Thr Arg Asp Arg Ser Asp Gln His Ile Gln Leu Gln Leu Ser Ala Glu
 50 55 60
 Ser Ala Gly Glu Val Tyr Ile Lys Gly Thr Glu Thr Gly Gln Tyr Leu
 65 70 75 80
 Ala Met Asp Thr Glu Gly Leu Leu Tyr Gly Ser Gln Thr Pro Asn Glu
 85 90 95
 Glu Cys Leu Phe Leu Glu Arg Leu Glu Glu Asn His Tyr Asn Thr Tyr
 100 105 110
 Thr Ser Lys Lys His Ala Glu Lys Asn Trp Phe Val Gly Leu Lys Lys
 115 120 125
 Asn Gly Ser Cys Lys Arg Gly Pro Arg Thr His Tyr Gly Gln Lys Ala
 130 135 140
 Ile Leu Phe Leu Pro Leu Pro Val Ser Ser Asp
 145 150 155

<210> 23

<211> 154

<212> PRT

<213> Mus musculus

<400> 23

Met Ala Ala Ser Gly Ile Thr Ser Leu Pro Ala Leu Pro Glu Asp Gly
 1 5 10 15
 Gly Ala Ala Phe Pro Pro Gly His Phe Lys Asp Pro Lys Arg Leu Tyr
 20 25 30
 Cys Lys Asn Gly Gly Phe Phe Leu Arg Ile His Pro Asp Gly Arg Val
 35 40 45
 Asp Gly Val Arg Glu Lys Ser Asp Pro His Val Lys Leu Gln Leu Gln
 50 55 60
 Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala Asn Arg
 65 70 75 80
 Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys Val
 85 90 95
 Thr Glu Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr Asn
 100 105 110
 Thr Tyr Arg Ser Arg Lys Tyr Ser Ser Trp Tyr Val Ala Leu Lys Arg
 115 120 125
 Thr Gly Gln Tyr Lys Leu Gly Ser Lys Thr Gly Pro Gly Gln Lys Ala
 130 135 140

Ile Leu Phe Leu Pro Met Ser Ala Lys Ser
145 150

<210> 24

<211> 245

<212> PRT

<213> Mus musculus

<400> 24

Met Gly Leu Ile Trp Leu Leu Leu Leu Ser Leu Leu Glu Pro Ser Trp
1 5 10 15

Pro Thr Thr Gly Pro Gly Thr Arg Leu Arg Arg Asp Ala Gly Gly Arg
20 25 30

Gly Gly Val Tyr Glu His Leu Gly Gly Ala Pro Arg Arg Arg Lys Leu
35 40 45

Tyr Cys Ala Thr Lys Tyr His Leu Gln Leu His Pro Ser Gly Arg Val
50 55 60

Asn Gly Ser Leu Glu Asn Ser Ala Tyr Ser Ile Leu Glu Ile Thr Ala
65 70 75 80

Val Glu Val Gly Val Val Ala Ile Lys Gly Leu Phe Ser Gly Arg Tyr
85 90 95

Leu Ala Met Asn Lys Arg Gly Arg Leu Tyr Ala Ser Asp His Tyr Asn
100 105 110

Ala Glu Cys Glu Phe Val Glu Arg Ile His Glu Leu Gly Tyr Asn Thr
115 120 125

Tyr Ala Ser Arg Leu Tyr Arg Thr Gly Ser Ser Gly Pro Gly Ala Gln
130 135 140

Arg Gln Pro Gly Ala Gln Arg Pro Trp Tyr Val Ser Val Asn Gly Lys
145 150 155 160

Gly Arg Pro Arg Arg Gly Phe Lys Thr Arg Arg Thr Gln Lys Ser Ser
165 170 175

Leu Phe Leu Pro Arg Val Leu Gly His Lys Asp His Glu Met Val Arg
180 185 190

Leu Leu Gln Ser Ser Gln Pro Arg Ala Pro Gly Glu Gly Ser Gln Pro
195 200 205

Arg Gln Arg Arg Gln Lys Lys Gln Ser Pro Gly Asp His Gly Lys Met
210 215 220

Glu Thr Leu Ser Thr Arg Ala Thr Pro Ser Thr Gln Leu His Thr Gly
225 230 235 240

Gly Leu Ala Val Ala
245

<210> 25

<211> 202

<212> PRT

<213> Mus musculus

<400> 25

Met Ala Lys Arg Gly Pro Thr Thr Gly Thr Leu Leu Pro Arg Val Leu
 1 5 10 15

Leu Ala Leu Val Val Ala Leu Ala Asp Arg Gly Thr Ala Ala Pro Asn
 20 25 30

Gly Thr Arg His Ala Glu Leu Gly His Gly Trp Asp Gly Leu Val Ala
 35 40 45

Arg Ser Leu Ala Arg Leu Pro Val Ala Ala Gln Pro Pro Gln Ala Ala
 50 55 60

Val Arg Ser Gly Ala Gly Asp Tyr Leu Leu Gly Leu Lys Arg Leu Arg
 65 70 75 80

Arg Leu Tyr Cys Asn Val Gly Ile Gly Phe His Leu Gln Val Leu Pro
 85 90 95

Asp Gly Arg Ile Gly Gly Val His Ala Asp Thr Arg Asp Ser Leu Leu
 100 105 110

Glu Leu Ser Pro Val Gln Arg Gly Val Val Ser Ile Phe Gly Val Ala
 115 120 125

Ser Arg Phe Phe Val Ala Met Ser Ser Arg Gly Lys Leu Phe Gly Val
 130 135 140

Pro Phe Phe Thr Asp Glu Cys Lys Phe Lys Glu Ile Leu Leu Pro Asn
 145 150 155 160

Asn Tyr Asn Ala Tyr Glu Ala Tyr Ala Tyr Pro Gly Met Phe Met Ala
 165 170 175

Leu Ser Lys Asn Gly Arg Thr Lys Lys Gly Asn Arg Val Ser Pro Thr
 180 185 190

Met Lys Val Thr His Phe Leu Pro Arg Leu
 195 200

<210> 26

<211> 264

<212> PRT

<213> Mus musculus

<400> 26

Met Ser Leu Ser Leu Leu Phe Leu Ile Phe Cys Ser His Leu Ile His
 1 5 10 15

Ser Ala Trp Ala His Gly Glu Lys Arg Leu Thr Pro Glu Gly Gln Pro
 20 25 30

Ala Pro Pro Arg Asn Pro Gly Asp Ser Ser Gly Ser Arg Gly Arg Ser
 35 40 45

Ser Ala Thr Phe Ser Ser Ser Ser Ala Ser Ser Pro Val Ala Ala Ser
 50 55 60

Pro Gly Ser Gln Gly Ser Gly Ser Glu His Ser Ser Phe Gln Trp Ser
 65 70 75 80
 Pro Ser Gly Arg Arg Thr Gly Ser Leu Tyr Cys Arg Val Gly Ile Gly
 85 90 95
 Phe His Leu Gln Ile Tyr Pro Asp Gly Lys Val Asn Gly Ser His Glu
 100 105 110
 Ala Ser Val Leu Ser Ile Leu Glu Ile Phe Ala Val Ser Gln Gly Ile
 115 120 125
 Val Gly Ile Arg Gly Val Phe Ser Asn Lys Phe Leu Ala Met Ser Lys
 130 135 140
 Lys Gly Lys Leu His Ala Ser Ala Lys Phe Thr Asp Asp Cys Lys Phe
 145 150 155 160
 Arg Glu Arg Phe Gln Glu Asn Ser Tyr Asn Thr Tyr Ala Ser Ala Ile
 165 170 175
 His Arg Thr Glu Lys Thr Gly Arg Glu Trp Tyr Val Ala Leu Asn Lys
 180 185 190
 Arg Gly Lys Ala Lys Arg Gly Cys Ser Pro Arg Val Lys Pro Gln His
 195 200 205
 Val Ser Thr His Phe Leu Pro Arg Phe Lys Gln Ser Glu Gln Pro Glu
 210 215 220
 Leu Ser Phe Thr Val Thr Val Pro Glu Lys Lys Lys Pro Pro Val Lys
 225 230 235 240
 Pro Lys Val Pro Leu Ser Gln Pro Arg Arg Ser Pro Ser Pro Val Lys
 245 250 255
 Tyr Arg Leu Lys Phe Arg Phe Gly
 260

<210> 27
 <211> 208
 <212> PRT
 <213> Mus musculus

<400> 27
 Met Ala Leu Gly Gln Arg Leu Phe Ile Thr Met Ser Arg Gly Ala Gly
 1 5 10 15
 Arg Val Gln Gly Thr Leu Gln Ala Leu Val Phe Leu Gly Val Leu Val
 20 25 30
 Gly Met Val Val Pro Ser Pro Ala Gly Ala Arg Ala Asn Gly Thr Leu
 35 40 45
 Leu Asp Ser Arg Gly Trp Gly Thr Leu Leu Ser Arg Ser Arg Ala Gly
 50 55 60
 Leu Ala Gly Glu Ile Ser Gly Val Asn Trp Glu Ser Gly Tyr Leu Val
 65 70 75 80

Gly Ile Lys Arg Gln Arg Arg Leu Tyr Cys Asn Val Gly Ile Gly Phe
 85 90 95
 His Leu Gln Val Pro Pro Asp Gly Arg Ile Ser Gly Thr His Glu Glu
 100 105 110
 Asn Pro Tyr Ser Leu Leu Glu Ile Ser Thr Val Glu Arg Gly Val Val
 115 120 125
 Ser Leu Phe Gly Val Lys Ser Ala Leu Phe Ile Ala Met Asn Ser Lys
 130 135 140
 Gly Arg Leu Tyr Thr Thr Pro Ser Phe His Asp Glu Cys Lys Phe Arg
 145 150 155 160
 Glu Thr Leu Leu Pro Asn Asn Tyr Asn Ala Tyr Glu Ser Asp Leu Tyr
 165 170 175
 Arg Gly Thr Tyr Ile Ala Leu Ser Lys Tyr Gly Arg Val Lys Arg Gly
 180 185 190
 Ser Lys Val Ser Pro Ile Met Thr Val Thr His Phe Leu Pro Arg Ile
 195 200 205

<210> 28
 <211> 194
 <212> PRT
 <213> Mus musculus

<400> 28
 Met Arg Lys Trp Ile Leu Thr Arg Ile Leu Pro Thr Leu Leu Tyr Arg
 1 5 10 15
 Ser Cys Phe His Leu Val Cys Leu Val Gly Thr Ile Ser Leu Ala Cys
 20 25 30
 Asn Asp Met Ser Pro Glu Gln Thr Ala Thr Ser Val Asn Cys Ser Ser
 35 40 45
 Pro Glu Arg His Thr Arg Ser Tyr Asp Tyr Met Glu Gly Gly Asp Ile
 50 55 60
 Arg Val Arg Arg Leu Phe Cys Arg Thr Gln Trp Tyr Leu Arg Ile Asp
 65 70 75 80
 Lys Arg Gly Lys Val Lys Gly Thr Gln Glu Met Lys Asn Ser Tyr Asn
 85 90 95
 Ile Met Glu Ile Arg Thr Val Ala Val Gly Ile Val Ala Ile Lys Gly
 100 105 110
 Val Glu Ser Glu Tyr Tyr Leu Ala Met Asn Lys Glu Gly Lys Leu Tyr
 115 120 125
 Ala Lys Lys Glu Cys Asn Glu Asp Cys Asn Phe Lys Glu Leu Ile Leu
 130 135 140

Glu Asn His Tyr Asn Thr Tyr Ala Ser Ala Lys Trp Thr His Ser Gly
 145 150 155 160

Gly Glu Met Phe Val Ala Leu Asn Gln Lys Gly Ile Pro Val Lys Gly
 165 170 175

Lys Lys Thr Lys Lys Glu Gln Lys Thr Ala His Phe Leu Pro Met Ala
 180 185 190

Ile Thr

<210> 29

<211> 268

<212> PRT

<213> Mus musculus

<400> 29

Met Gly Ser Pro Arg Ser Ala Leu Ser Cys Leu Leu Leu His Leu Leu
 1 5 10 15

Val Leu Cys Leu Gln Ala Gln Val Arg Ser Ala Ala Gln Lys Arg Gly
 20 25 30

Pro Gly Ala Gly Asn Pro Ala Asp Thr Leu Gly Gln Gly His Glu Asp
 35 40 45

Arg Pro Phe Gly Gln Arg Ser Arg Ala Gly Lys Asn Phe Thr Asn Pro
 50 55 60

Ala Pro Asn Tyr Pro Glu Glu Gly Ser Lys Glu Gln Arg Asp Ser Val
 65 70 75 80

Leu Pro Lys Val Thr Gln Arg His Val Arg Glu Gln Ser Leu Val Thr
 85 90 95

Asp Gln Leu Ser Arg Arg Leu Ile Arg Thr Tyr Gln Leu Tyr Ser Arg
 100 105 110

Thr Ser Gly Lys His Val Gln Val Leu Ala Asn Lys Arg Ile Asn Ala
 115 120 125

Met Ala Glu Asp Gly Asp Pro Phe Ala Lys Leu Ile Val Glu Thr Asp
 130 135 140

Thr Phe Gly Ser Arg Val Arg Val Arg Gly Ala Glu Thr Gly Leu Tyr
 145 150 155 160

Ile Cys Met Asn Lys Lys Gly Lys Leu Ile Ala Lys Ser Asn Gly Lys
 165 170 175

Gly Lys Asp Cys Val Phe Thr Glu Ile Val Leu Glu Asn Asn Tyr Thr
 180 185 190

Ala Leu Gln Asn Ala Lys Tyr Glu Gly Trp Tyr Met Ala Phe Thr Arg
 195 200 205

Lys Gly Arg Pro Arg Lys Gly Ser Lys Thr Arg Gln His Gln Arg Glu
 210 215 220

Val His Phe Met Lys Arg Leu Pro Arg Gly His His Thr Thr Glu Gln
 225 230 235 240

Ser Leu Arg Phe Glu Phe Leu Asn Tyr Pro Pro Phe Thr Arg Ser Leu
 245 250 255

Arg Gly Ser Gln Arg Thr Trp Ala Pro Glu Pro Arg
 260 265

<210> 30

<211> 208

<212> PRT

<213> Mus musculus

<400> 30

Met Ala Pro Leu Gly Glu Val Gly Ser Tyr Phe Gly Val Gln Asp Ala
 1 5 10 15

Val Pro Phe Gly Asn Val Pro Val Leu Pro Val Asp Ser Pro Val Leu
 20 25 30

Leu Asn Asp His Leu Gly Gln Ser Glu Ala Gly Gly Leu Pro Arg Gly
 35 40 45

Pro Ala Val Thr Asp Leu Asp His Leu Lys Gly Ile Leu Arg Arg Arg
 50 55 60

Gln Leu Tyr Cys Arg Thr Gly Phe His Leu Glu Ile Phe Pro Asn Gly
 65 70 75 80

Thr Ile Gln Gly Thr Arg Lys Asp His Ser Arg Phe Gly Ile Leu Glu
 85 90 95

Phe Ile Ser Ile Ala Val Gly Leu Val Ser Ile Arg Gly Val Asp Ser
 100 105 110

Gly Leu Tyr Leu Gly Met Asn Glu Lys Gly Glu Leu Tyr Gly Ser Glu
 115 120 125

Lys Leu Thr Gln Glu Cys Val Phe Arg Glu Gln Phe Glu Glu Asn Trp
 130 135 140

Tyr Asn Thr Tyr Ser Ser Asn Leu Tyr Lys His Val Asp Thr Gly Arg
 145 150 155 160

Arg Tyr Tyr Val Ala Leu Asn Lys Asp Gly Thr Pro Arg Glu Gly Thr
 165 170 175

Arg Thr Lys Arg His Gln Lys Phe Thr His Phe Leu Pro Arg Pro Val
 180 185 190

Asp Pro Asp Lys Val Pro Glu Leu Tyr Lys Asp Ile Leu Ser Gln Ser
 195 200 205

<210> 31

<211> 209

<212> PRT

<213> Mus musculus

<400> 31

Met Trp Lys Trp Ile Leu Thr His Cys Ala Ser Ala Phe Pro His Leu
 1 5 10 15

Pro Gly Cys Cys Cys Cys Phe Leu Leu Leu Phe Leu Val Ser Ser Phe
 20 25 30

Pro Val Thr Cys Gln Ala Leu Gly Gln Asp Met Val Ser Gln Glu Ala
 35 40 45

Thr Asn Cys Ser Ser Ser Ser Ser Ser Phe Ser Ser Pro Ser Ser Ala
 50 55 60

Gly Arg His Val Arg Ser Tyr Asn His Leu Gln Gly Asp Val Arg Trp
 65 70 75 80

Arg Arg Leu Phe Ser Phe Thr Lys Tyr Phe Leu Thr Ile Glu Lys Asn
 85 90 95

Gly Lys Val Ser Gly Thr Lys Asn Glu Asp Cys Pro Tyr Ser Val Leu
 100 105 110

Glu Ile Thr Ser Val Glu Ile Gly Val Val Ala Val Lys Ala Ile Asn
 115 120 125

Ser Asn Tyr Tyr Leu Ala Met Asn Lys Lys Gly Lys Leu Tyr Gly Ser
 130 135 140

Lys Glu Phe Asn Asn Asp Cys Lys Leu Lys Glu Arg Ile Glu Glu Asn
 145 150 155 160

Gly Tyr Asn Thr Tyr Ala Ser Phe Asn Trp Gln His Asn Gly Arg Gln
 165 170 175

Met Tyr Val Ala Leu Asn Gly Lys Gly Ala Pro Arg Arg Gly Gln Lys
 180 185 190

Thr Arg Arg Lys Asn Thr Ser Ala His Phe Leu Pro Met Thr Ile Gln
 195 200 205

Thr

<210> 32

<211> 225

<212> PRT

<213> Mus musculus

<400> 32

Met Ala Ala Leu Ala Ser Ser Leu Ile Arg Gln Lys Arg Glu Val Arg
 1 5 10 15

Glu Pro Gly Gly Ser Arg Pro Val Ser Ala Gln Arg Arg Val Cys Pro
 20 25 30

Arg Gly Thr Lys Ser Leu Cys Gln Lys Gln Leu Leu Ile Leu Leu Ser

35 40 45
 Lys Val Arg Leu Cys Gly Gly Arg Pro Thr Arg Gln Asp Arg Gly Pro
 50 55 60
 Glu Pro Gln Leu Lys Gly Ile Val Thr Lys Leu Phe Cys Arg Gln Gly
 65 70 75 80
 Phe Tyr Leu Gln Ala Asn Pro Asp Gly Ser Ile Gln Gly Thr Pro Glu
 85 90 95
 Asp Thr Ser Ser Phe Thr His Phe Asn Leu Ile Pro Val Gly Leu Arg
 100 105 110
 Val Val Thr Ile Gln Ser Ala Lys Leu Gly His Tyr Met Ala Met Asn
 115 120 125
 Ala Glu Gly Leu Leu Tyr Ser Ser Pro His Phe Thr Ala Glu Cys Arg
 130 135 140
 Phe Lys Glu Cys Val Phe Glu Asn Tyr Tyr Val Leu Tyr Ala Ser Ala
 145 150 155 160
 Leu Tyr Arg Gln Arg Arg Ser Gly Arg Ala Trp Tyr Leu Gly Leu Asp
 165 170 175
 Lys Glu Gly Arg Val Met Lys Gly Asn Arg Val Lys Lys Thr Lys Ala
 180 185 190
 Ala Ala His Phe Val Pro Lys Leu Leu Glu Val Ala Met Tyr Arg Glu
 195 200 205
 Pro Ser Leu His Ser Val Pro Glu Thr Ser Pro Ser Ser Pro Pro Ala
 210 215 220
 His
 225

<210> 33
 <211> 243
 <212> PRT
 <213> Mus musculus

<400> 33
 Met Ala Ala Ala Ile Ala Ser Ser Leu Ile Arg Gln Lys Arg Gln Ala
 1 5 10 15
 Arg Glu Ser Asn Ser Asp Arg Val Ser Ala Ser Lys Arg Arg Ser Ser
 20 25 30
 Pro Ser Lys Asp Gly Arg Ser Leu Cys Glu Arg His Val Leu Gly Val
 35 40 45
 Phe Ser Lys Val Arg Phe Cys Ser Gly Arg Lys Arg Pro Val Arg Arg
 50 55 60
 Arg Pro Glu Pro Gln Leu Lys Gly Ile Val Thr Arg Leu Phe Ser Gln
 65 70 75 80
 Gln Gly Tyr Phe Leu Gln Met His Pro Asp Gly Thr Ile Asp Gly Thr

85 90 95
 Lys Asp Glu Asn Ser Asp Tyr Thr Leu Phe Asn Leu Ile Pro Val Gly
 100 105 110
 Leu Arg Val Val Ala Ile Gln Gly Val Lys Ala Ser Leu Tyr Val Ala
 115 120 125
 Met Asn Gly Glu Gly Tyr Leu Tyr Ser Ser Asp Val Phe Thr Pro Glu
 130 135 140
 Cys Lys Phe Lys Glu Ser Val Phe Glu Asn Tyr Tyr Val Ile Tyr Ser
 145 150 155 160
 Ser Thr Leu Tyr Arg Gln Gln Glu Ser Gly Arg Ala Trp Phe Leu Gly
 165 170 175
 Leu Asn Lys Glu Gly Gln Ile Met Lys Gly Asn Arg Val Lys Lys Thr
 180 185 190
 Lys Pro Ser Ser His Phe Val Pro Lys Pro Ile Glu Val Cys Met Tyr
 195 200 205
 Arg Glu Pro Ser Leu His Glu Ile Gly Glu Lys Gln Gly Arg Ser Arg
 210 215 220
 Lys Ser Ser Gly Thr Pro Thr Met Asn Gly Gly Lys Val Val Asn Gln
 225 230 235 240
 Asp Ser Thr

<210> 34
 <211> 245
 <212> PRT
 <213> Mus musculus

<400> 34
 Met Thr Ala Ala Ile Ala Ser Ser Leu Ile Arg Gln Lys Arg Gln Ala
 1 5 10 15
 Arg Glu Arg Glu Lys Ser Asn Ala Cys Lys Cys Val Ser Ser Pro Ser
 20 25 30
 Lys Gly Lys Thr Ser Cys Asp Lys Asn Lys Leu Asn Val Phe Ser Arg
 35 40 45
 Val Lys Leu Phe Gly Ser Lys Lys Arg Arg Arg Arg Arg Pro Glu Pro
 50 55 60
 Gln Leu Lys Gly Ile Val Thr Lys Leu Tyr Ser Arg Gln Gly Tyr His
 65 70 75 80
 Leu Gln Leu Gln Ala Asp Gly Thr Ile Asp Gly Thr Lys Asp Glu Asp
 85 90 95
 Ser Thr Tyr Thr Leu Phe Asn Leu Ile Pro Val Gly Leu Arg Val Val
 100 105 110
 Ala Ile Gln Gly Val Gln Thr Lys Leu Tyr Leu Ala Met Asn Ser Glu

115 120 125
 Gly Tyr Leu Tyr Thr Ser Glu His Phe Thr Pro Glu Cys Lys Phe Lys
 130 135 140
 Glu Ser Val Phe Glu Asn Tyr Tyr Val Thr Tyr Ser Ser Met Ile Tyr
 145 150 155 160
 Arg Gln Gln Gln Ser Gly Arg Gly Trp Tyr Leu Gly Leu Asn Lys Glu
 165 170 175
 Gly Glu Ile Met Lys Gly Asn His Val Lys Lys Asn Lys Pro Ala Ala
 180 185 190
 His Phe Leu Pro Lys Pro Leu Lys Val Ala Met Tyr Lys Glu Pro Ser
 195 200 205
 Leu His Asp Leu Thr Glu Phe Ser Arg Ser Gly Ser Gly Thr Pro Thr
 210 215 220
 Lys Ser Arg Ser Val Ser Gly Val Leu Asn Gly Gly Lys Ser Met Ser
 225 230 235 240
 His Asn Glu Ser Thr
 245

<210> 35

<211> 247

<212> PRT

<213> Mus musculus

<400> 35

Met Ala Ala Ala Ile Ala Ser Gly Leu Ile Arg Gln Lys Arg Gln Ala
 1 5 10 15
 Arg Glu Gln His Trp Asp Arg Pro Ser Ala Ser Arg Arg Arg Ser Ser
 20 25 30
 Pro Ser Lys Asn Arg Gly Leu Phe Asn Gly Asn Leu Val Asp Ile Phe
 35 40 45
 Ser Lys Val Arg Ile Phe Gly Leu Lys Lys Arg Arg Leu Arg Arg Gln
 50 55 60
 Asp Pro Gln Leu Lys Gly Ile Val Thr Arg Leu Tyr Cys Arg Gln Gly
 65 70 75 80
 Tyr Tyr Leu Gln Met His Pro Asp Gly Ala Leu Asp Gly Thr Lys Asp
 85 90 95
 Asp Ser Thr Asn Ser Thr Leu Phe Asn Leu Ile Pro Val Gly Leu Arg
 100 105 110
 Val Val Ala Ile Gln Gly Val Lys Thr Gly Leu Tyr Ile Ala Met Asn
 115 120 125
 Gly Glu Gly Tyr Leu Tyr Pro Ser Glu Leu Phe Thr Pro Glu Cys Lys
 130 135 140
 Phe Lys Glu Ser Val Phe Glu Asn Tyr Tyr Val Ile Tyr Ser Ser Met

145 150 155 160
 Leu Tyr Arg Gln Gln Glu Ser Gly Arg Ala Trp Phe Leu Gly Leu Asn
 165 170 175
 Lys Glu Gly Gln Val Met Lys Gly Asn Arg Val Lys Lys Thr Lys Pro
 180 185 190
 Ala Ala His Phe Leu Pro Lys Pro Leu Glu Val Ala Met Tyr Arg Glu
 195 200 205
 Pro Ser Leu His Asp Val Gly Glu Thr Val Pro Lys Ala Gly Val Thr
 210 215 220
 Pro Ser Lys Ser Thr Ser Ala Ser Ala Ile Met Asn Gly Gly Lys Pro
 225 230 235 240
 Val Asn Lys Cys Lys Thr Thr
 245

<210> 36
 <211> 218
 <212> PRT
 <213> Mus musculus

<400> 36
 Met Ala Arg Lys Trp Asn Gly Arg Ala Val Ala Arg Ala Leu Val Leu
 1 5 10 15
 Ala Thr Leu Trp Leu Ala Val Ser Gly Arg Pro Leu Ala Gln Gln Ser
 20 25 30
 Gln Ser Val Ser Asp Glu Asp Pro Leu Phe Leu Tyr Gly Trp Gly Lys
 35 40 45
 Ile Thr Arg Leu Gln Tyr Leu Tyr Ser Ala Gly Pro Tyr Val Ser Asn
 50 55 60
 Cys Phe Leu Arg Ile Arg Ser Asp Gly Ser Val Asp Cys Glu Glu Asp
 65 70 75 80
 Gln Asn Glu Arg Asn Leu Leu Glu Phe Arg Ala Val Ala Leu Lys Thr
 85 90 95
 Ile Ala Ile Lys Asp Val Ser Ser Val Arg Tyr Leu Cys Met Ser Ala
 100 105 110
 Asp Gly Lys Ile Tyr Gly Leu Ile Arg Tyr Ser Glu Glu Asp Cys Thr
 115 120 125
 Phe Arg Glu Glu Met Asp Cys Leu Gly Tyr Asn Gln Tyr Arg Ser Met
 130 135 140
 Lys His His Leu His Ile Ile Phe Ile Gln Ala Lys Pro Arg Glu Gln
 145 150 155 160
 Leu Gln Asp Gln Lys Pro Ser Asn Phe Ile Pro Val Phe His Arg Ser
 165 170 175
 Phe Phe Glu Thr Gly Asp Gln Leu Arg Ser Lys Met Phe Ser Leu Pro

180	185	190
Leu Glu Ser Asp Ser Met Asp Pro Phe Arg Met Val Glu Asp Val Asp		
195	200	205
His Leu Val Lys Ser Pro Ser Phe Gln Lys		
210	215	

<210> 37
 <211> 207
 <212> PRT
 <213> Rattus norvegicus

<400> 37
 Met Ala Glu Val Gly Gly Val Phe Ala Ser Leu Asp Trp Asp Leu Gln
 1 5 10 15
 Gly Phe Ser Ser Ser Leu Gly Asn Val Pro Leu Ala Asp Ser Pro Gly
 20 25 30
 Phe Leu Asn Glu Arg Leu Gly Gln Ile Glu Gly Lys Leu Gln Arg Gly
 35 40 45
 Ser Pro Thr Asp Phe Ala His Leu Lys Gly Ile Leu Arg Arg Arg Gln
 50 55 60
 Leu Tyr Cys Arg Thr Gly Phe His Leu Glu Ile Phe Pro Asn Gly Thr
 65 70 75 80
 Val His Gly Thr Arg His Asp His Ser Arg Phe Gly Ile Leu Glu Phe
 85 90 95
 Ile Ser Leu Ala Val Gly Leu Ile Ser Ile Arg Gly Val Asp Ser Gly
 100 105 110
 Leu Tyr Leu Gly Met Asn Glu Arg Gly Glu Leu Phe Gly Ser Lys Lys
 115 120 125
 Leu Thr Arg Glu Cys Val Phe Arg Glu Gln Phe Glu Glu Asn Trp Tyr
 130 135 140
 Asn Thr Tyr Ala Ser Thr Leu Tyr Lys His Ser Asp Ser Glu Arg Gln
 145 150 155 160
 Tyr Tyr Val Ala Leu Asn Lys Asp Gly Ser Pro Arg Glu Gly Tyr Arg
 165 170 175
 Thr Lys Arg His Gln Lys Phe Thr His Phe Leu Pro Arg Pro Val Asp
 180 185 190
 Pro Ser Lys Leu Pro Ser Met Ser Arg Asp Leu Phe Arg Tyr Arg
 195 200 205

<210> 38
 <211> 207
 <212> PRT
 <213> Mus musculus
 <400> 38

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Met Tyr Ser Ala Pro Ser Ala Cys Thr Cys Leu Cys Leu His Phe Leu
 1             5             10             15

Leu Leu Cys Phe Gln Val Gln Val Leu Ala Ala Glu Glu Asn Val Asp
      20             25             30

Phe Arg Ile His Val Glu Asn Gln Thr Arg Ala Arg Asp Asp Val Ser
      35             40             45

Arg Lys Gln Leu Arg Leu Tyr Gln Leu Tyr Ser Arg Thr Ser Gly Lys
      50             55             60

His Ile Gln Val Leu Gly Arg Arg Ile Ser Ala Arg Gly Glu Asp Gly
      65             70             75             80

Asp Lys Tyr Ala Gln Leu Leu Val Glu Thr Asp Thr Phe Gly Ser Gln
      85             90             95

Val Arg Ile Lys Gly Lys Glu Thr Glu Phe Tyr Leu Cys Met Asn Arg
      100            105            110

Lys Gly Lys Leu Val Gly Lys Pro Asp Gly Thr Ser Lys Glu Cys Val
      115            120            125

Phe Ile Glu Lys Val Leu Glu Asn Asn Tyr Thr Ala Leu Met Ser Ala
      130            135            140

Lys Tyr Ser Gly Trp Tyr Val Gly Phe Thr Lys Lys Gly Arg Pro Arg
      145            150            155            160

Lys Gly Pro Lys Thr Arg Glu Asn Gln Gln Asp Val His Phe Met Lys
      165            170            175

Arg Tyr Pro Lys Gly Gln Ala Glu Leu Gln Lys Pro Phe Lys Tyr Thr
      180            185            190

Thr Val Thr Lys Arg Ser Arg Arg Ile Arg Pro Thr His Pro Gly
      195            200            205

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<210> 39
<211> 11
<212> PRT
<213> Human immunodeficiency virus type 1

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<400> 39
Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg
 1             5             10

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<210> 40
<211> 15
<212> PRT
<213> Artificial Sequence

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<220>
<223> Description of Artificial Sequence: internalizing
      domain derived from HIV tat protein

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<400> 40
Gly Gly Gly Gly Tyr Gly Arg Lys Lys Arg Arg Gln Arg Arg Arg

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1	5	10	15
<p><210> 41 <211> 22 <212> DNA <213> Artificial Sequence</p>			
<p><220> <223> Description of Artificial Sequence: oligonucleotide; PCR primer</p>			
<p><400> 41 ctatcccaat gcctcccccac tg 22</p>			
<p><210> 42 <211> 21 <212> DNA <213> Artificial Sequence</p>			
<p><220> <223> Description of Artificial Sequence: oligonucleotide; PCR primer</p>			
<p><400> 42 cgcccctgac caccctaataat g 21</p>			
<p><210> 43 <211> 23 <212> DNA <213> Artificial Sequence</p>			
<p><220> <223> Description of Artificial Sequence: oligonucleotide; 5'RACE primer</p>			
<p><400> 43 gtgtggaatt gtgagcggat aac 23</p>			
<p><210> 44 <211> 22 <212> DNA <213> Artificial Sequence</p>			
<p><220> <223> Description of Artificial Sequence: oligonucleotide; 5'RACE primer</p>			
<p><400> 44 ctgatgggggt gcgccatcca ca 22</p>			
<p><210> 45 <211> 23 <212> DNA <213> Artificial Sequence</p>			
<p><220></p>			

<223> Description of Artificial Sequence:
oligonucleotide; nested PCR primer

<400> 45
ctatgaccat gattacgcca agc

23

<210> 46
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; nested PCR primer

<400> 46
cattcttggt gatctgcagg tggt

24

<210> 47
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; 3'RACE primer

<400> 47
cggcctcctg ttcacaggag ctc

23

<210> 48
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; 3'RACE primer

<400> 48
cgggcctctt cgctattacg c

21

<210> 49
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; nested PCR primer

<400> 49
gcgccgagga caacagcccg a

21

<210> 50
<211> 21

<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; nested PCR primer

<400> 50
tggcgaaagg gggatgtgct g 21

<210> 51
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; PCR primer

<400> 51
tccaccaccc tgttgctgta g 21

<210> 52
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; PCR primer

<400> 52
gaccacagtc catgccatca ct 22

<210> 53
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; PCR primer

<400> 53
ctatcccaat gcctccccac tg 22

<210> 54
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
oligonucleotide; PCR primer

<400> 54
cgcccctgac caccctaat g 21